# Cardiovascular disease risk assessment in HIV-infected black South Africans: A longitudinal study

### **M** Duvenhage



Dissertation submitted in partial fulfilment of the requirements for the degree Master of Science in Physiology at the North-West University

Supervisor: Prof CMT Fourie

Co-Supervisor: Prof JM van Rooyen

Final Copy: May 2018
Student number: 23440848



#### **Acknowledgements**

First and foremost I want to thank my Heavenly Father for making **anything** possible (*Philippians 4:13*).

I would like to acknowledge the following people/institutions for the roles they played in making this dissertation a success and for their constant support:

- Prof. Carla MT Fourie and Prof. Johannes M van Rooyen for their willingness to be my supervisors, for their guidance, motivation and valuable insights throughout this project.
- To my husband and parents for their encouragement and love throughout this project.
- To Ms. C Vorster for the language editing of this dissertation.
- To all the participants, researchers, field workers and supporting staff who formed part of the PURE study and making the completion of this dissertation possible.
- The financial assistance of the National Research Foundation, (NRF-SARChI) towards this research.

#### **Affirmation by authors**

The following researchers contributed to this project:

#### Ms. Marlene Duvenhage

Responsible for literature research, statistical analyses, interpretation of results and writing of the manuscript and dissertation.

#### Prof. Carla MT Fourie (Supervisor)

Responsible for this study design and planning of the study, took part in PURE study data collection, supervised statistical analysis of data, as well as giving recommendations regarding the writing and construction of the script.

#### Prof. Johannes M van Rooyen (Co-supervisor)

Took part in the PURE study data collection and planning of this study and gave recommendations regarding interpretation of data, writing and construction of the manuscript and dissertation.

This is a statement from the co-authors confirming their individual role in the study and giving their permission that the manuscript may form part of this dissertation.

\_\_\_\_ Courie \_\_

Ms. M Duvenhage

Prof. CMT Fourie

Prof. JM van Rooyen

#### **Table of contents**

Summary	7
List of abbreviations	12
Chapter 1: Introduction	14
Background	15
Motivation	17
References	18
Chapter 2: Literature review	24
1. Cardiovascular disease in Africa	25
2. Cardiovascular disease and Human Immunodeficiency Virus in Africa	25
3. Cardiovascular risk factors	27
3.1 Non-modifiable cardiovascular risk factors	27
3.1.1 Age	27
3.1.2 Sex and Ethnicity	28
3.2 Modifiable cardiovascular disease risk factors	28
3.2.1 Lipids	28
3.2.2 Obesity	29
3.2.3 Hypertension	30
3.2.4 Hyperglycaemia	31
3.2.5 Smoking	32
3.2.6 Alcohol	32
4. Inflammation, cardiovascular disease and human immunodeficiency virus	33
5. End-organ damage and cardiovascular disease	34
5.1 Chronic kidney disease	34
5.2 Arterial stiffness	35
5.3 Left ventricular hypertrophy	36
5.4 Atherosclerosis	36
6. Cardiovascular risk assessment	37

7. The Framingham risk score	38
8. The Reynolds risk score	39
9. Cardiovascular risk assessment and end-organ damage	40
Summary	41
Aims	42
Objectives	42
Hypotheses	42
References	44
Chapter 3: Methodology	67
Materials and methods	68
References	76
Chapter 4: Manuscript for publication	78
Abstract	82
Introduction	83
Materials and methods	84
Results	86
Discussion	92
Conclusion	94
Conflict of interest	94
Acknowledgements	94
References	96
Chapter 5: General conclusions and recommendations	102
Introduction	103
Hypotheses and comparison to relevant literature	103
Discussion of main findings	104
Conclusion	105
Chance and confounding	105
Recommendations to future research	106

References	107
Appendices	109
Decleration by language editor	

Turn-it-in report

#### **Summary**

#### **Motivation**

A double burden of non-communicable -and communicable diseases exists in South Africa which include high prevalence of cardiovascular disease (CVD) and human immunodeficiency virus (HIV) infection. Coronary heart disease (CHD), a sub-category of CVD, affects more African-American individuals than white individuals. The high prevalence of CVD and HIV decreases the quality of life for those living with HIV. However, being treated with antiretroviral drugs are known to increase the life expectancy for the HIV-infected population. Those living with HIV presented with higher prevalence of arterial stiffness, atherosclerosis, kidney disease and left ventricular hypertrophy (LVH). Despite the advantages of antiretroviral therapy (ART) use, it was shown to have worsen CVD risk for the HIV-infected population. The need for risk assessment in the African HIV-infected population burdened by both coronary heart disease (CHD) and CVD are important.

#### Aim

The aim of this study was to determine the risk scores of 10-year CHD and CVD by making use of the Framingham -and Reynolds risk score models, respectively, and to determine associations of the risk score models with measures of end-organ damage.

#### Methodology

This study is embedded in the South African arm of the Prospective Urban and Rural Epidemiological (PURE) study and included African participants from the North-West Province of South Africa who were infected with HIV for at least 10-years. The PURE-South African study consisted of baseline (2005) and follow-up (2015) data. A number of 2010 individuals participated in the PURE study during baseline, where 322 participants were newly identified as being infected with HIV. Ten years later 100 of the 322 HIV-infected participants took part in the follow-up study of which 29 participants were excluded due to incomplete data sets. A number of 71 HIV-infected participants remained and they were matched to 71 HIV-free controls according to age, sex and locality.

Anthropometric measurements included height, weight and waist circumference (WC), followed by the calculation of body mass index (BMI). Regarding cardiovascular measurements, systolic and diastolic blood pressure (SBP and DBP), carotid-femoral

pulse wave velocity (cfPWV), only for follow-up, and intima-media thickness (IMT) were determined. Biochemical variables included total cholesterol (TC), high and low-density lipoprotein cholesterol (HDL-C and LDL-C), triglycerides (TG), glycosylated haemoglobin A1c (HbA1c), C-reactive protein (CRP) and HIV status. Creatinine clearance (CrCl) was calculated with the Cockcroft-Gault formula. The CD4 counts were determined, in whole blood, at baseline by the National Health Laboratory using flow cytometric analysis and at follow-up with finger-prick blood and a point-of-care device PIMA<sup>TM</sup> CD4 (Alere, Jena, Germany).

The statistical analyses were performed by using Statistica® 13 (StatSoft, Inc., Tulsa, OK, USA). The Framingham and Reynolds risk score were determined with excel spreadsheets, separately, during baseline and follow-up. Basic descriptive statistics were used to determine normal distribution of the data and logarithmic transformation was applied and presented as geometric mean with 5<sup>th</sup> and 95<sup>th</sup> percentiles if skewed. Groups were compared using independent t-tests and Chi-square tests as appropriate. Co-morbidity prevalence was defined by using cut-off values. Associations of measures of end-organ damage with the risk scores were determined by making use of Pearson and partial correlations. Odds ratios with 95% confidence intervals (CI) were calculated. Median values of risk scores and measures of end-organ damage were used as cut-off values.

#### **Results and conclusion**

The HIV-infected group presented with lower HDL-C (p<0.01) and CrCl levels (p=0.02) at baseline, compared to the HIV-free group. At follow-up the HIV-infected group had significant lower BMI (p<0.01), WC (p<0.01) and HbA1c (p=0.01) compared to the HIV-free control group.

The CD4 counts of the HIV-infected group was higher (p=0.03) at follow-up compared to baseline.

More HIV-free participants were overweight (p=0.02), had diabetes (p=0.04), had lower HDL-C (men) (p=0.013) and had a higher prevalence of microalbuminuria (p=0.04), compared to the HIV-infected group at follow-up.

No differences were seen between the HIV-infected and HIV-free group with either the Framingham or Reynolds risk score at baseline or follow-up. The Framingham risk score was higher in the HIV-infected group at follow-up when compared to the HIV-free controls, however both the HIV-free controls and HIV-infected group had a higher

Reynolds risk score at follow-up than at baseline. No differences were seen between the ART and no-ART group.

A borderline negative correlation (p=0.053) was seen between CHD and CrCl, while CVD risk correlated negatively with cfPWV in the HIV-infected group at follow-up, however after adjusting for age, sex, WC, SBP, CRP, CD4 and ART use no correlations were seen. The CVD risk correlated negatively with Cornell product in the HIV-free group, however after adjusting for age, sex, WC, SBP, CRP, tobacco use and alcohol use no correlations were seen. No significant odds ratios were found for having a higher than median CHD or CVD risk with higher than median PWV, IMT and Cornell product or lower than median CrCl.

To conclude, despite their HIV-status for at least 10-years and 80% of participants receiving ART, we found that the HIV-infected participants did not have higher CHD or CVD risk when compared to the HIV-free participants. Those infected with HIV did also not show associations of risk scores with measures of end-organ damage.

**Keywords:** Cardiovascular disease, Human immunodeficiency virus, Risk assessment, Africans, End-organ damage

#### **Preface**

The article format was used for the completion of this dissertation. The chosen journal for publication of the manuscript is *Heart, Lung and Circulation*. This dissertation is written in English.

The structure of this dissertation is as follows:

- **Chapter 1:** The introductory chapter consists of a background, motivation, aim, objectives and hypotheses of the study.
- Chapter 2: A complete literature study of the relevant topics.
- Chapter 3: A complete methods section.
- **Chapter 4:** The research manuscript which includes instructions for authors of the journal *Heart, Lung and Circulation*, an introduction, the materials and methods, results, discussion, conclusion and acknowledgements.
- Chapter 5: Concluding remarks, a critical discussion of the findings and recommendations.

A reference list is provided at the end of each chapter, according to the Vancouver referencing style, as prescribed by the journal *Heart, Lung and Circulation* and was used throughout the dissertation.

#### List of tables and figures

#### **Tables**

#### Chapter 2:

Table 1 - Cardiovascular disease risk factors.

#### Chapter 3:

Table 2 - The Framingham risk table implemented for South Africans.

#### Chapter 4:

- Table 3 Characteristics of HIV-infected participants (n=71) and HIV-free participants (n=71) in the baseline study (2005) and follow-up study (2015).
- Table 4 Prevalence of co-morbidities among the HIV-infected and HIV-free participants at follow-up.
- Table 5 Partial correlations of 10-year Framingham (CHD) risk score and Reynolds (CVD) risk score with measures of end-organ damage at 10-year follow-up in the HIV-free and HIV-infected group.
- Table 6 Odds Ratios of 10-year risk scores and measures of end-organ damage in the HIV-free and HIV-infected participants.

#### **Figures**

#### Chapter 4:

- Figure 1 Baseline and follow-up Reynolds (CVD) and Framingham (CHD) risk scores in HIV-free and HIV-infected groups with the influence of treatment.
- Figure 2 Scatterplots indicating the Pearson correlations between IMTf and CrCl with the Framingham risk score (CHD) and Cornell product with the Reynolds risk score (CVD) in the HIV-free and HIV-infected group at 10-year follow-up.

#### List of abbreviations

AIDS - Acquired Immunodeficiency Syndrome

ART - Antiretroviral therapy

BMI - Body mass index

cfPWV - Carotid-femoral pulse wave velocity

CDC - Centers for Disease Control and prevention

CHD - Coronary heart disease

CI - Confidence interval

CKD - Chronic kidney disease

CP - Cornell product

CrCl - Creatinine clearance

CVD - Cardiovascular disease

DBP - Diastolic blood pressure

ECG - Electrocardiography

FSGS - Focal Segmental Glomerulosclerosis

GFR - Glomerular filtration rate

GGT - Gamma-glutamyltransferase

HbA1c - Glycosylated haemoglobin A1c

HDL-C - High-density lipoprotein cholesterol

HIV - Human immunodeficiency virus

hsCRP - High-sensitivity C-reactive protein

IL-6 - Interleukin 6

IMT - Intima-media thickness

LDL-C - Low-density lipoprotein cholesterol

LVH - Left ventricular hypertrophy

NCEP - National Cholesterol Education Program

NRTI - Nucleoside Reverse Transcriptase Inhibitor

NNRTI - Non-nucleoside Reverse Transcriptase Inhibitor

PI - Protease inhibitor

PURE - Prospective Urban and Rural Epidemiological

SBP - Systolic blood pressure

TC - Total cholesterol

TG - Triglyceride

uACR - Urinary albumin to creatinine ratio

WC - Waist circumference

WHO - World Health Organization

## Chapter 1: Introduction

#### Background

Cardiovascular disease (CVD) is considered a leading cause of morbidity and mortality in the African population [1, 2]. Nearly 38.3% of non-communicable disease (NCD) deaths were due to CVD in sub-Saharan Africa in 2013 [3]. During 2015, statistics showed that the Eastern and Southern parts of Africa had the highest number of human immunodeficiency virus (HIV) infections, compared to other African regions, with the number being 19 million individuals [4]. It has been concluded by Triant et al. (2007) that HIV-infected individuals will most likely develop an increasing burden of CVD as they age and live longer [5]. Aging also leads to chronic inflammation development and this process may be described as "inflammaging" [6]. In addition to inflammaging, HIV infection as well as the aging process itself share cellular immunologic similarities such as reduced T-cell function [7].

Lipid profile disturbances, or better known as dyslipidaemia, contribute to CVD development in those infected with HIV and may be due to the virus itself, ART or both [8]. In 2009, Duprez et al. stated that, regardless of other CVD risk factors present in HIV-infected individuals, lower levels of high-density lipoprotein cholesterol (HDL-C) were highly associated with developing CVD [9]. Black HIV-infected individuals usually have lower levels of total cholesterol (TC), triglycerides (TG) [8] and higher levels of HDL-C which is seen to be a "protective" pattern against the development of ischaemic heart disease and atherosclerosis in this population [10]. However, being treated with ART may contribute to developing hyperlipidaemia [11, 12].

Hypertension is the most significant CVD risk factor and accounts for more than half of CVD morbidity and mortality [13]. It is also the main cause of 7.6 million deaths worldwide every year [14]. In a study conducted by Lloyd-Sherlock et al. (2014) it has been found that South Africa is the country with the highest prevalence of hypertension among the elderly [15]. Black individuals are also more prone to develop hypertension [16, 17].

Studies have indicated that the risk of developing CVD is even more likely to occur in individuals who are classified as being diabetic and as much as 80% of mortality in individuals with diabetes occur through CVD [18, 19]. Diabetes, a state of chronic inflammation [20], is higher amongst HIV-infected individuals compared to their HIV-free counterparts [21].

Smoking is, according to literature, known as the most established modifiable CVD risk factor and the relationship thereof with CVD development is well-described [22]. HIV-infected individuals are more likely to be smokers, when compared to their HIV-free counterparts [23, 24]. Up to 40%, if not more, of myocardial infarction risk may be prevented by the cessation of smoking among those infected with HIV [25]. In a study conducted by Ohsawa et al. (2005), it has been found that smokers have higher levels of C-reactive protein (CRP) [26]. Those living with HIV show elevated levels of CRP [27, 28] and it has also been suggested that CRP may be used as a marker for the atherosclerotic burden in the African-American population [29]. The use of ART in previously untreated HIV-infected female individuals is associated with an increase in CRP levels [30].

Microalbuminuria may be seen as an early marker of renal damage and it is known to be associated with CVD risk [31-34]. HIV-infected Africans have a higher prevalence of chronic kidney disease (CKD) [35, 36]. Low-grade albuminuria is associated with the progression of arterial stiffness [37]. It has been found that HIV-infected individuals without ART show early onset of aortic stiffness which correlates with atherosclerosis and is also associated with CVD [38]. Van Vonderen et al. (2009) concluded that HIV-infected individuals show lower compliance and distensibility components in the carotid, femoral and brachial arteries [39]. A recent study has found that an increase in the pulse wave velocity (PWV) is associated with the use of ART, especially efavirenz [40]. Schoffelen et al. (2015) concluded that intima-media thickness (IMT) of the HIV-infected South African women is increased when compared to their male counterparts and has a stronger association with cardiovascular risk factors [41].

Cardiovascular risk assessment is seen to be beneficial, and Blom (2011) concluded that it is a science in "rapid evolution" [42]. Many cardiovascular risk assessment methods exist of which the Framingham risk score is probably the most popular. The risk factors currently included in the Framingham risk score models (one for men and women) [43] are: age, TC, smoking, HDL-C and SBP. The Framingham risk model is used to calculate the 10-year coronary heart disease (CHD) score [42]. Cardiovascular risk also relates to family history, inflammation (CRP) and glycosylated haemoglobin A1c (HbA1c) (among diabetics) [44, 45]. The Reynolds risk score includes the same risk variables as those of the Framingham risk score, but CRP and HbA1c are also incorporated [44]. These risk factors are included in two separate models for men [45]

and women [44]. The Reynolds risk score is part of the risk scores which calculate the 10-year cardiovascular risk.

Both the Reynolds and Framingham risk models received class one recommendations from the American Heart Association as well as the American College of Cardiology [46] and both risk models were endorsed as part of the national guidelines for the CVD prevention programme in Canada [47]. Both these risk models were also mainly developed for Caucasians [43, 44, 48], however, in a recent study it was found that the application of the Framingham risk model to a large black study population was applicable to the black participants [49]. The Reynolds risk score on the other hand showed improved discrimination overall and in black and white women in particular [50]. Therefore this risk model was also used in the present study.

End-organ damage has an independent prognostic significance, irrespective of whether it involves the function and/or structure of the blood vessels, kidney, brain or heart [51]. According to Mancia et al. (2007), once organ damage has been detected, the patients usually already have a high cardiovascular risk [52].

#### **Motivation**

A dual burden of non-communicable (CVD) and communicable (HIV) diseases exist in South Africa and the literature suggests that HIV-infected individuals have higher prevalence of CVD. Literature regarding cardiovascular risk in HIV-infected Africans are sparse. Risk monitoring is also known to be a considerable component to achieve risk management, and therefore we thought it well to implement these risk scores in our study (longitudinal regard) in order to identify risk in the African HIV-infected population.

This data may be beneficial to all healthcare systems in order to make important decisions concerning the cardiovascular risk among HIV-infected black South Africans and to develop the necessary treatment.

#### References

- 1. Rehm J, Mathers C, Popova S, Thavorncharoensap M, Teerawattananon Y, Patra J. Global burden of disease and injury and economic cost attributable to alcohol use and alcohol-use disorders. Lancet 2009;373(9682):2223-2233.
- 2. Stewart S, Libhaber E, Carrington M, Damasceno A, Abbasi H, Hansen C, et al. The clinical consequences and challenges of hypertension in urban-dwelling black Africans: Insights from The Heart of Soweto study. Int J Cardiol 2011;146(1):22-27.
- 3. Mensah GA, Roth GA, Sampson UK, Moran AE, Feigin VL, Forouzanfar MH, et al. Mortality from cardiovascular diseases in sub-Saharan Africa, 1990-2013: A systematic analysis of data from the global burden of disease study 2013. Cardiovasc J Afr 2015;26:6-10.
- 4. Joint United Nations Programme on HIV/AIDS (UNAIDS). Global AIDS update [internet]. 2016 [cited 2016]. Available from:

  <a href="http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf">http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf</a>.
- 5. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. J Clin Endocrinol Metab 2007;92(7):2506-2512.
- 6. Franceschi C, Bonafe M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann N Y Acad Sci 2000;908:244-254.
- 7. Desai S, Landay A. Early immune senescence in HIV disease. Curr HIV/AIDS Rep 2010;7(1):4-10.
- 8. Dube MP, Lipshultz SE, Fichtenbaum CJ, Greenberg R, Schecter AD, Fisher SD. Effects of HIV infection and antiretroviral therapy on the heart and vasculature. Circulation 2008;118(2):36-40.
- 9. Duprez DA, Kuller LH, Tracy R, Otvos J, Cooper DA, Hoy J, et al. Lipoprotein particle subclasses, cardiovascular disease and HIV infection. Atherosclerosis 2009;207(2):524-529.

- 10. Steyn K, Damasceno A. Lifestyle and related risk factors for chronic diseases. Disease and mortality in sub-Saharan Africa 2006;2:247-265.
- 11. Grunfeld C, Pang M, Doerrler W, Shigenaga JK, Jensen P, Feingold KR. Lipids, lipoproteins, triglyceride clearance, and cytokines in human immunodeficiency virus infection and the acquired immunodeficiency syndrome. J Clin Endocrinol Metab 1992;74(5):1045-1052.
- 12. Feingold KR, Krauss RM, Pang M, Doerrler W, Jensen P, Grunfeld C. The hypertriglyceridemia of acquired immunodeficiency syndrome is associated with an increased prevalence of low density lipoprotein subclass pattern b. J Clin Endocrinol Metab 1993;76(6):1423-1427.
- 13. Ezzati M, Vander Hoorn S, Lawes CM, Leach R, James WP, Lopez AD, et al. Rethinking the "diseases of affluence" paradigm: Global patterns of nutritional risks in relation to economic development. PLoS Med 2005;2(5):133.
- 14. Lawes CMM, Hoorn SV, Rodgers A. Global burden of blood-pressure-related disease, 2001. Lancet 371(9623):1513-1518.
- 15. Lloyd-Sherlock P, Beard J, Minicuci N, Ebrahim S, Chatterji S. Hypertension among older adults in low- and middle-income countries: Prevalence, awareness and control. Int J Epidemiol 2014;43(1):116-128.
- 16. Fray JC, Douglas JG. Pathophysiology of hypertension in blacks: Springer; 2013.
- 17. Hajjar I, Kotchen TA. Trends in prevalence, awareness, treatment, and control of hypertension in the united states, 1988-2000. JAMA 2003;290(2):199-206.
- 18. Stamler J, Vaccaro O, Neaton JD, Wentworth D. Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the multiple risk factor intervention trial. Diabetes Care 1993;16(2):434-444.
- 19. Woodward M, Zhang X, Barzi F, Pan W, Ueshima H, Rodgers A, et al. The effects of diabetes on the risks of major cardiovascular diseases and death in the Asia-Pacific region. Diabetes Care 2003;26(2):360-366.

- 20. Pickup JC, Mattock MB, Chusney GD, Burt D. NIDDM as a disease of the innate immune system: Association of acute-phase reactants and interleukin-6 with metabolic syndrome x. Diabetologia 1997;40(11):1286-1292.
- 21. Brown TT, Cole SR, Li X, et al. Antiretroviral therapy and the prevalence and incidence of diabetes mellitus in The Multicenter AIDS Cohort study. Arch Intern Med 2005;165(10):1179-1184.
- 22. Teo KK, Ounpuu S, Hawken S, Pandey MR, Valentin V, Hunt D, et al. Tobacco use and risk of myocardial infarction in 52 countries in The Interheart study: A case-control study. Lancet 2006;368(9536):647-658.
- 23. Tesoriero JM, Gieryic SM, Carrascal A, Lavigne HE. Smoking among HIV positive New Yorkers: Prevalence, frequency, and opportunities for cessation. AIDS Behav 2010;14(4):824-835.
- 24. Nahvi S, Cooperman NA. Review: The need for smoking cessation among HIV-positive smokers. AIDS Educ Prev 2009;21:14-27.
- 25. Rasmussen LD, Helleberg M, May MT, Afzal S, Kronborg G, Larsen CS, et al. Myocardial infarction among Danish HIV-infected individuals: Population-attributable fractions associated with smoking. Clin Infect Dis 2015;60(9):1415-1423.
- 26. Ohsawa M, Okayama A, Nakamura M, Onoda T, Kato K, Itai K, et al. CRP levels are elevated in smokers but unrelated to the number of cigarettes and are decreased by long-term smoking cessation in male smokers. Prev Med 2005;41(2):651-656.
- 27. Feldman JG, Goldwasser P, Holman S, DeHovitz J, Minkoff H. C-reactive protein is an independent predictor of mortality in women with HIV-1 infection. J Acquir Immune Defic Syndr 2003;32(2):210-214.
- 28. Reingold J, Wanke C, Kotler D, Lewis C, Tracy R, Heymsfield S, et al. Association of HIV infection and HIV/HCV coinfection with c-reactive protein levels: The Fat Redistribution and Metabolic change in HIV infection (FRAM) study. J Acquir Immune Defic Syndr 2008;48(2):142-148.
- 29. Sung JH, Lee JE, Samdarshi TE, Nagarajarao HS, Taylor JK, Agrawal KK, et al. C-reactive protein and subclinical cardiovascular disease among African Americans:(The Jackson Heart study). J Cardiovasc Med 2014;15(5):371.

- 30. Shikuma CM, Ribaudo HJ, Zheng Y, Gulick RM, Meyer WA, Tashima KT, et al. Change in high-sensitivity c-reactive protein levels following initiation of efavirenz-based antiretroviral regimens in HIV-infected individuals. AIDS Res Hum Retroviruses 2011;27(5):461-468.
- 31. Hillege HL, Fidler V, Diercks GF, van Gilst WH, de Zeeuw D, van Veldhuisen DJ, et al. Urinary albumin excretion predicts cardiovascular and noncardiovascular mortality in general population. Circulation 2002;106(14):1777-1782.
- 32. James MA, Fotherby MD, Potter JF. Screening tests for microalbuminuria in non-diabetic elderly subjects and their relation to blood pressure. Clin Sci 1995;88(2):185-190.
- 33. Mattock MB, Morrish NJ, Viberti G, Keen H, Fitzgerald AP, Jackson G. Prospective study of microalbuminuria as predictor of mortality in NIDDM. Diabetes 1992;41(6):736-41.
- 34. Roest M, Banga JD, Janssen WM, Grobbee DE, Sixma JJ, de Jong PE, et al. Excessive urinary albumin levels are associated with future cardiovascular mortality in postmenopausal women. Circulation 2001;103(25):3057-3061.
- 35. Cailhol J, Nkurunziza B, Izzedine H, Nindagiye E, Munyana L, Baramperanye E, et al. Prevalence of chronic kidney disease among people living with HIV/AIDS in Burundi: A cross-sectional study. BMC Nephrol 2011;12:40.
- 36. Emem CP, Arogundade F, Sanusi A, Adelusola K, Wokoma F, Akinsola A. Renal disease in HIV-seropositive patients in Nigeria: An assessment of prevalence, clinical features and risk factors. Nephrol Dial Transplant 2008;23(2):741-746.
- 37. Hermans MM, Henry R, Dekker JM, Kooman JP, Kostense PJ, Nijpels G, et al. Estimated glomerular filtration rate and urinary albumin excretion are independently associated with greater arterial stiffness: The HOORN study. J Am Soc Nephrol 2007;18(6):1942-1952.
- 38. Ngatchou W, Lemogoum D, Ndobo P, Yagnigni E, Tiogou E, Nga E, et al. Increased burden and severity of metabolic syndrome and arterial stiffness in treatment-naive HIV+ patients from Cameroon. Vasc Health Risk Manag 2013;9:509-516.

- 39. Van Vonderen MG, Hassink EA, Van Agtmael MA, Stehouwer CD, Danner SA, Reiss P, et al. Increase in carotid artery intima-media thickness and arterial stiffness but improvement in several markers of endothelial function after initiation of antiretroviral therapy. J Infect Dis 2009;199(8):1186-1194.
- 40. Gleason RL, Jr., Caulk AW, Seifu D, Parker I, Vidakovic B, Getenet H, et al. Current efavirenz (efv) or ritonavir-boosted lopinavir (lpv/r) use correlates with elevate markers of atherosclerosis in HIV-infected subjects in Addis Ababa, Ethiopia. PLoS One 2015;10(4):117-125.
- 41. Schoffelen AF, De Groot E, Tempelman HA, Visseren FL, Hoepelman AI, Barth RE. Carotid intima media thickness in mainly female HIV-infected subjects in rural South Africa: Association with cardiovascular but not HIV-related factors. Clin Infect Dis 2015;61(10):1606-1614.
- 42. Blom DJ. Cardiovascular risk assessment. S Afr Fam Prac 2011;53(2):121-128.
- 43. D'Agostino RB, Vasan RS, Pencina MJ, Wolf PA, Cobain M, Massaro JM, et al. General cardiovascular risk profile for use in primary care: The Framingham Heart study. Circulation 2008;117(6):743-753.
- 44. Ridker PM, Buring JE, Rifai N, Cook NR. Development and validation of improved algorithms for the assessment of global cardiovascular risk in women: The Reynolds risk score. JAMA 2007;297(6):611-619.
- 45. Ridker PM, Paynter NP, Rifai N, Gaziano JM, Cook NR. C-reactive protein and parental history improve global cardiovascular risk prediction: The Reynolds risk score for men. Circulation 2008;118(22):2243-2251.
- 46. Greenland P, Alpert JS, Beller GA, Benjamin EJ, Budoff MJ, Fayad ZA, et al. 2010 ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults: A report of the American College of Cardiology Foundation/American Heart Association task force on practice guidelines. J Am Coll Cardiol 2010;56(25):50-103.
- 47. Genest J, McPherson R, Frohlich J, Anderson T, Campbell N, Carpentier A, et al. 2009 Canadian cardiovascular society/Canadian guidelines for the diagnosis and treatment of dyslipidemia and prevention of cardiovascular disease in the adult 2009 recommendations. Can J Cardiol 2009;25(10):567-579.

- 48. Executive summary of the third report of The National Cholesterol Education Program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel iii). JAMA 2001;285(19):2486-2497.
- 49. Goff DC, Jr, Lloyd-Jones DM. The pooled cohort risk equations—black risk matters. JAMA Cardiol 2016;1(1):12-14.
- 50. Cook NR, Paynter NP, Eaton CB, Manson JE, Martin LW, Robinson JG, et al. Comparison of the Framingham and Reynolds risk scores for global cardiovascular risk prediction in The Multiethnic Women's Health Initiative. Circulation 2012;125(14):1748-1756.
- 51. Mancia G, Laurent S, Agabiti-Rosei E, Ambrosioni E, Burnier M, Caulfield MJ, et al. Reappraisal of European guidelines on hypertension management: A European society of hypertension task force document. Blood Press 2009;18(6):308-347.
- 52. Mancia G, De Backer G, Dominiczak A, Cifkova R, Fagard R, Germano G, et al. 2007 ESH-ESC practice guidelines for the management of arterial hypertension: ESH-ESC task force on the management of arterial hypertension. J Hypertens 2007;25(9):1751-1762.

## Chapter 2: Literature review

#### 1. Cardiovascular disease in Africa

Non-communicable diseases are predicted to become the leading cause of mortality in Africa by 2030 [1]. The World Health Organization (WHO) has identified cardiovascular disease (CVD) as being one of the main categories of NCD [2]. In a study conducted in an urban East African setting, the researchers found that the participants showed a high prevalence of cardiovascular risk, especially among the women [3]. Coronary heart disease (CHD) may be classified as a sub-category of CVD, mainly affecting the coronary arteries, and the mortality thereof is known to be higher in African-Americans than in white Americans [4].

Besides being a region with a high prevalence of CVD, sub-Saharan Africa is also known as a region with a high prevalence of human immunodeficiency virus (HIV) infection [5].

#### 2. Cardiovascular disease and Human Immunodeficiency Virus in Africa

According to the WHO, HIV continues to be a crucial global public health concern and has claimed roughly 35 million lives up until 2016 [6]. In 2015 there were approximately 19 million HIV-infected individuals in Eastern and Southern Africa [7] and it is predicted that over 10 million individuals aged older than 50 years will be living with HIV in sub-Saharan Africa by 2030 [8, 9].

Human immunodeficiency virus is described as a lentivirus (member of the *Retroviridae* family) that progresses to acquired immunodeficiency syndrome (AIDS) [10]. The immune system is demolished by retroviruses and the enzyme called reverse transcriptase is produced, which plays a role in the demolition of the human immune system [10].

Human immunodeficiency virus-1 is characterised by genetic heterogeneity, which is driven by several factors, including a lack of proofreading ability of the reverse transcriptase [11, 12], rapid in vivo HIV-1 turnover [13], host selective immune pressures [14] and recombination of events during the viral replication [15]. Due to the variability of HIV-1 variants, it may be divided into three main phylogenetic groups: group M (main), group O (outlier) and group N (non-M/non-O) [16-18]. Different subtypes of HIV-1 exist, whereas the main virus which prevails in South Africa is HIV-1 subtype C [19, 20]. Human immunodeficiency virus-1 subtype B is responsible for

the infections in North America, Europe and Australia and its genome differs from HIV-1 subtype C by as much as 30% [19, 21, 22].

HIV demolishes the T-helper cells, which play an essential role in immunity, and infects any cell that expresses CD4 proteins [10]. The CD4 T-helper cell count indicates the immune status and is associated with clinical manifestations of the HIV infection [23]. Numerous mechanisms are related to viral replication during HIV progression. During the primary infection, HIV-1 leads to strong T-cell responses which may persist during the phase of chronic infection and this may be due to the replication of the virus [24, 25]. Several studies suggest that HIV gene products may directly induce the activation of macrophages, lymphocytes and proinflammatory chemokine and cytokine production [14-16]. The protein gp120 activates cells or enhances their responsiveness to activation through binding to CD4 or other co-receptors [26-28].

The use of antiretroviral therapy (ART) is needed to decelerate the progression of HIV replication. ART may be associated with higher levels of inflammation, lower levels of high-density lipoprotein cholesterol (HDL-C) [29] and higher levels of triglycerides (TG) which may aggravate CVD risk in HIV-infected individuals and may be directly related to viral infection, ART use or both [30]. In contrast with the latter, Rajasuriar et al. (2015) concluded that early ART use may reduce inflammation, however, the timing and duration of ART use are important factors [31]. High-density lipoprotein cholesterol has anti-inflammatory characteristics through the inhibition of the expression of endothelial adhesion molecules [32-34]. Therefore lower levels of HDL-C may reduce the anti-inflammatory function of the molecule and lead to further aggravation of inflammation. Participants exhibiting high levels of TG frequently have additional risk factors, such as insulin resistance which may affect their sensitivity to atherosclerosis development [35].

Untreated HIV infection induces pro-atherogenic mechanisms in the body and speeds up the progression of atherosclerosis leading to the development of CVD [29]. According to Baker et al. (2015) accelerated cardiovascular risk is becoming more prevalent in the HIV-infected population [29] due to prolonged life expectancy and a higher prevalence of traditional lifestyle cardiovascular risk factors in the HIV-infected population, such as smoking [36, 37].

#### 3. Cardiovascular risk factors

#### 3.1 Non-modifiable cardiovascular risk factors

#### 3.1.1 Age

Age is known to be non-modifiable cardiovascular risk factor and Petoumenos et al. (2014) have found that CVD events in men increase with an increase in age [38]. A general "hallmark" seen in aging tissues is chronic inflammation [39]. Low-grade inflammation together with chronic —and systemic inflammation during the aging process can be described as a process called inflammaging [39].

Petoumenos et al. (2014) stated that the risk for developing CVD, myocardial infarction and CHD increase with twofold as the individual grows older [38]. As a person ages, arterial stiffness, measured by pulse wave velocity (PWV), increases with approximately 0.1 m/s per year [40]. As demonstrated by Lee et al. (2010), endothelial dysfunction, lower elastin levels, higher collagen levels, increased deposition of calcium and the growth of smooth muscle cells lead to vascular wall thickening and lower compliance of blood vessels [41] which may increase the risk for developing CVD.

In addition to inflammaging, HIV infection and the aging process share cellular immunological similarities [42]. It includes reduced naïve T-cell generation, T-cell receptor diversity, more memory T-cells and reduced function and shortened telomeres [42]. Early immune aging (continuous processes of antigen burden and immune activation) has been seen in the HIV-infected population [42].

Those infected with HIV are at increased risk of age-related non-AIDS mortality and morbidity compared to HIV-free controls [43]. It is hypothesised that HIV-infected individuals not only undergo chronological aging, but also biological aging which is mediated by increased cellular deterioration [44]. Chronological age is an indefinite measure of biological aging.

Triant et al. (2007) have shown diverging rates of myocardial infarction between HIV-infected and HIV-free participants with increasing age [45]. They also concluded that CVD will most likely become an increasing burden in HIV-infected individuals as they age and live longer [45].

#### 3.1.2 Sex and Ethnicity

Studies found that ethnicity is highly associated with the development of CVD [45]. Black individuals living in South Africa are more likely to develop hypertension, heart failure and atherosclerosis [46, 47] and it has been suggested that black Africans are less susceptible to angiotensin converting enzyme inhibitors, mainly due to the frequency of low renin hypertension, compared to white individuals [48]. The above mentioned may be the reason for black Africans developing hypertension and being more prevalent to develop CVD.

According to the Centers for Disease control and Prevention (CDC), black African American men accounted for one third of all HIV diagnoses in 2015 [49]. They also reported that African American women accounted for 11% of all HIV diagnoses in 2015 and black women were 16 times more likely to be diagnosed with HIV when compared to white women [49].

Njelekela et al. (2009) have found strong evidence of high cardiovascular risk in Tanzania, particularly among the women in their study group [3]. Receiving ART along with Framingham risk factors and other cardiovascular risk factors such as hypertension and renal disease may contribute to CVD development in both men and women [50].

#### 3.2 Modifiable cardiovascular disease risk factors

#### **3.2.1 Lipids**

Dyslipidaemia is known to be a predominant CVD risk factor, especially in the HIV-infected population [51]. Lipid profile alterations, indicating lower HDL-C and higher triglycerides levels, form part of the risk to develop CVD in HIV-infected individuals and may be due to the virus itself, ART or both [30]. Black individuals have lower levels of total cholesterol (TC), TG [30] and higher levels of HDL-C compared to other populations such as white individuals [52] and it may indicate a "protective" pattern against the development of atherosclerosis and ischaemic heart disease in black individuals [52]. However, in a study conducted by Fourie et al. (2010), the HDL-C levels of the black HIV-infected African participants were lower than those normally associated with increased cardiovascular risk [53]. Triglyceride and HDL-C levels are evaluated together and inversely correlated [54-57].

Total cholesterol includes low-density lipoprotein cholesterol (LDL-C), HDL-C and TG. Elevated serum TC levels are associated with the development of atherosclerosis and correlate with CHD [58]. Levels of TC above 11.60 mmol/L have been directly associated with a two-fold higher risk of developing hypertension in the general population [59] and may increase the risk for developing CVD.

Atherosclerosis is an inflammatory disease. Studies have shown that HDL-C has important anti-inflammatory effects and promotes these anti-inflammatory effects by inhibiting the expression of endothelial adhesion molecules [32-34]. The latter therefore explains why an increase in HDL-C levels with 0.06 mmol/L may reduce the risk of developing CVD by 2% [60].

Low-density lipoprotein cholesterol is seen as the main source of cholesterol build-up in the arteries and arterial blockage [61]. Three clinical trials showed that lower levels of LDL-C in diabetics reduced the incidence of CVD development [62-64]. Howard et al. (2000) have found that LDL-C is a powerful predictor of the development of CHD and have suggested that a 0.60 mmol/L increase in LDL-C would lead to a 12% increase in CVD [65].

Triglycerides are main constituents of natural oils and fats in the human body. A triglyceride molecule consists of three fatty acid groups and glycerol. Elevated TG levels are predictors of CVD development [66, 67] as studies been suggested that increased TG levels are major independent risk factor for the development of CHD [68-70]. Studies concluded that TG may contribute to the development of CVD since an association of TG with atherogenic lipoproteins exists [71].

Several studies have found that ART may contribute to developing hyperlipidaemia and/or lipodystrophy (abnormal conditions of the body's adipose tissue) [72, 73]. Nucleoside reverse transcriptase inhibitors (NRTI), most likely stavudine (which form part of the first line ART in South Africa), may lead to increased levels of TG, LDL-C and TC [74].

#### 3.2.2 Obesity

Obesity has been acknowledged for decades as a significant contributing factor to develop various chronic diseases such as hypertension and CVD [75-77]. An increased prevalence of obesity is also seen in low-and-middle income countries, and people classified as being middle-aged ( $\pm$  45 – 64 years of age) have the highest

prevalence of obesity [78]. African-American women are more likely to be obese [79] and it has been suggested that black South African women are also more likely to have a high prevalence of abdominal obesity [80]. McCormick et al. (2014) have also found in their study that HIV-infected women from sub-Saharan Africa are more prone to present with obesity than their male counterparts [81].

By 2008, an estimated 502 million adults were classified worldwide as being obese (BMI >30 kg/m²) and 14.6 million adults as being overweight [82]. Body mass index (BMI) has been used for the identification of individuals at risk of developing obesity and related conditions such as CVD [83]. Each five-unit increase in BMI was associated with an increase in CVD mortality of 29% in women and 34% in men [84]. Another measuring method is the waist circumference (WC) which is one of the most common proxy measures of the presence of visceral fat accumulation [85]. Central obesity, as indicated by increased WC measurements, is an important cardiovascular risk factor in populations globally, including those living in sub-Saharan Africa [86, 87].

In a study conducted by Ogunmola et al. (2014), the HIV-free individuals were more likely to be obese, compared to HIV-infected individuals [88]. A recent study found that the women attributed their weight gain to ART and it was suggested that weight gain in the HIV-infected women occurred as a result of successful ART [81]. An increase in BMI with an average of 2.4 kg/m² is seen during a six-month ART initiation period and 3.5 kg/m² after a year of ART use among HIV-infected individuals living in SA [89].

#### 3.2.3 Hypertension

Hypertension is the most significant cardiovascular risk factor accounting for more than half of cardiovascular morbidity and mortality [90] and is the main cause of 7.6 million deaths every year, around the globe [91]. An increase in arterial blood pressure may lead to organ damage via hemodynamic load. Thus hypertension and the resulting hemodynamic load may lead to functional and structural cardiac changes [92], such as left ventricular hypertrophy (LVH) which is associated with hypertension. Hypertension may be characterised by the remodelling of small and large arteries as well as endothelial dysfunction [93]. This may lead to reduced dilation capability of high resistance vessels and the development of plaque formation, reduced coronary reserve and stenosis [93].

A study conducted by Yang et al. (2016) found that hypertension prevalence was 53.2% in the elderly (65 years and older), of which only 55% were aware of their hypertension status [94]. The prevalence of hypertension is higher in developing countries, as these countries have higher hypertension risk factors such as urbanisation and lifestyle changes [94]. Lloyd-Sherlock et al. (2014) found that South Africa is the country with the highest prevalence of hypertension among the elderly [95]. Although studies found that hypertension is more prevalent in sub-Saharan African countries, a meta-analysis conducted by Dillon et al. (2013) showed that HIV-infected sub-Saharan Africans had lower systolic blood pressure (SBP) and diastolic blood pressure (DBP) levels than the HIV-free control group [96].

The Heart of Soweto Study found that the prevalence of hypertension was 56% for urban South African study participants [47].

#### 3.2.4 Hyperglycaemia

Diabetes mellitus is a significant contributor to the overall CVD burden [97]. It has been suggested by the International Diabetes Foundation that the global population of individuals classified as being diabetic will increase from 382 million in 2013 to 592 million in 2035 with the highest increase of 109% in sub-Saharan Africa where an estimated number of 19.8 million individuals with diabetes will double to 41.5 million [98].

A recent study conducted showed that diabetes accounted for nearly 8.6% of the total mortality rate in sub-Saharan Africa in 2013 [99]. Several studies have indicated that the risk of developing CVD is twice more likely to occur in individuals with diabetes and 80% of mortality in individuals with diabetes occur as a result of CVD [100, 101]. Diabetes has been considered a state of chronic inflammation [102] and there is evidence to suggest that immune activation may precede insulin resistance in both diabetic and pre-diabetic conditions and may be seen as the factor which initially increases the CVD risk during the process of the disease [103].

The prevalence of diabetes is higher in HIV-infected individuals, compared to HIV-free individuals [104]. It has also been found that HIV-infected individuals receiving ART have an increased incidence of high fasting glucose levels (hyperglycaemia) [104].

The formation of glycosylated haemoglobin A1c (HbA1c) occurs by glycation of lysine and valine residues within haemoglobin [105]. The value of HbA1c is the bound fraction

of haemoglobin molecules to glucose molecules [106]. Glycosylated haemoglobin A1c is used to monitor diabetes as it is an index of glycaemia [107, 108].

Glycosylated haemoglobin A1c levels are associated with a higher risk of CVD [109]. Adams et al. (2009) have concluded that the risk to develop CHD is higher in women compared to their male counterparts when HbA1c was less than 6% [109]. Hypertriglyceridemic HIV-infected African-Americans making use of highly active antiretroviral treatment have a higher risk to develop diabetes [110].

#### 3.2.5 Smoking

Smoking is one of the most important established modifiable CVD risk factors and smoking may lead to CVD development [111]. Smoking is also related to lower levels of HDL-C [112] and to the progression of atherosclerosis [113]. Smoking could promote atherosclerosis by leading to alterations in lipid profiles of smokers, such as higher levels of serum cholesterol, triglycerides and LDL-C levels and lower HDL-C levels [114]. Cigarette smoking may also lead to decreased amounts of paraoxonase which is known as an enzyme that protects against the oxidation of LDL-C [115].

Black South Africans are more likely to be smokers compared to other races [116], and individuals infected with HIV are two to three times more likely to be smokers compared to HIV-free controls [117, 118]. Cardiovascular risk and mortality, due to smoking, have been found to be substantial in HIV-infected participants [119, 120]. Smoking and myocardial infarction development are directly related among the HIV-infected population [119]. Three out of four myocardial infarction events are associated with smoking in the HIV-infected population, whereas only one out of four myocardial infarction events are attributable to smoking among the general population [119]. The cessation of smoking could prevent up to 40% and more of myocardial infarction risk among those infected with HIV [119].

Those living with HIV who are smokers lose more "life-years" to smoking than to the virus itself [120].

#### 3.2.6 Alcohol

Alcohol use is seen as a lifestyle cardiovascular risk factor and is also known to be a coping mechanism, especially among the HIV-infected African population [121]. Alcohol consumption is a key risk factor for developing hypertension and mortality in the HIV-infected population [122, 123].

A positive relationship between alcohol consumption and lipids, such as HDL-C, LDL-C and TG together with blood pressure exists [124]. The above mentioned risk variables are elevated in heavy drinking, increasing the risk for these people to suffer from myocardial infarction [124]. Alcohol consumption may also have harmful effects on the kidneys, as it may lead to higher blood pressure due to increased pressure on the walls of arteries by injurious kidneys [125]. High levels of gamma-glutamyltransferase were found in smokers with chronic kidney disease (CKD) and this finding was associated with alcohol consumption [126]. Gamma-glutamyltransferase (GGT) is an enzyme found in the liver and is used as a marker of high levels of alcohol intake.

Africans show higher levels of GGT when being compared to other ethnic groups [127]. According to Lee et al. (2007) GGT correlates positively with cardiovascular risk markers such as, blood pressure, diabetes mellitus, dyslipidaemia and obesity [128]. It was reported that HIV infection was associated with higher GGT in Africans [129].

In terms of atherosclerosis [130], GGT was considered to be proatherogenic in atherosclerotic plaques [131] and alcohol consumption is significantly associated with arterial stiffness [132].

#### 4. Inflammation, cardiovascular disease and human immunodeficiency virus

C-reactive protein (CRP) is one of the most common markers used to assess and diagnose inflammation [133]. C-reactive protein belongs to the pentraxin family of proteins and levels increase 1000-fold or more during the occurrence of injury, inflammation or tissue death [134].

Literature suggests that CRP is considered a biomarker for the progression of CVD [135-137] and elevated CRP levels have been linked to future CVD events [138]. Atherosclerosis is regarded as an inflammatory disease [139] and it has been suggested that CRP may be used as a marker of atherosclerotic burden in African-Americans [140].

HIV-infection is associated with chronic activation of the immune system and a proinflammatory states [141]. CRP is known to be an indicator of immune activation in response to damage due to inflammation or infection [134, 142-144] and it binds to pathogens which then leads to the activation of the complement system for the enhancement of opsonisation and clearance [145]. Studies found higher levels of CRP in those living with HIV [146, 147]. Elevated levels of CRP, in those infected with HIV, are associated with the progression of HIV and with mortality in women [146, 148]. IL-6 is a is an immune protein (marker of inflammation) and is released in response to inflammation and/or infection [149].

Interleukin 6 (IL-6) levels are higher in the HIV-infected population (due to the presence of inflammation and infection) and may lead to CVD development [150]. It has also been found that black individuals living in sub-Saharan Africa have higher levels of CRP and IL-6 compared to white individuals [151].

Cigarette smoke, LDL-C and HDL-C were predictive in the development of elevated CRP levels [152]. Sex differences in CRP levels have been reported in the general population whereas women that complied with National Cholesterol Education Program (NCEP) metabolic syndrome criteria, have been reported to have higher CRP levels compared to their male counterparts [153].

#### 5. End-organ damage and cardiovascular disease

#### 5.1 Chronic kidney disease

The prevalence of kidney insufficiency and CVD rises with age [154] and even in the absence of kidney disease it is thought that human kidney function declines by 10 ml/min/1.73m<sup>2</sup> per decade [155]. Chronic kidney disease seen in the HIV-infected population develops as a result of viral-related risk factors as well as risk factors for developing kidney disease. HIV associated nephropathy develops in individuals with high viremia, typically seen during the advanced HIV disease stage or during acute HIV infection [156, 157].

According to the literature, HIV is associated with focal segmental glomerulosclerosis (FSGS) which is mainly seen in individuals of African descent [158]. Ethnic disparities in the rate of kidney failure progression to end-stage kidney disease exist between the African-American population and white individuals [159]. Focal segmental glomerulosclerosis associated with HIV, may present with low levels of protein excretion in urine to severe proteinuria [158]. According to the literature HIV-associated FSGS rapidly progresses to end-stage renal disease [158].

The determination of renal function includes isotopic determination of the glomerular filtration rate (GFR) [160] and creatinine clearance (CrCl) with the Cockcroft-Gault

equation [161]. A decrease in GFR was previously associated with an increased risk of CVD development together with higher morbidity and mortality [162]. Elevated levels of serum creatinine were associated with cardiovascular events and mortality [154]. In the HIV-infected population, lower GFR and higher serum creatinine occur [163].

The introduction of ART dramatically changed the clinical picture of HIV progression and HIV associated CKD [158]. With the effective use of ART, renal disease may stabilise, the disease process may be reversed and even disappear [164]. However, receiving tenofovir (a nucleoside transcriptase inhibitor commonly used as a first-line ART agent) is characterized by a significant loss of kidney function in those living with HIV [165].

#### **5.2 Arterial stiffness**

Stiffness of large elastic blood vessels is mainly determined by the extracellular matrix components situated in the arterial wall. Smooth muscle cells contribute minimally to mechanical behaviour of large elastic arteries [166]. The most important extracellular matrix components seen in large elastic arteries are elastin and collagen [167]. Elastin is a protein which provides reversible extensibility during cyclic loading of the cardiac cycle [167], while strength and prevention of failure at high pressure are provided by collagen [168]. During the aging process and disease progression, the elastic fibres are degraded and fragmented, which may lead to stiffening of the arterial wall [169].

An increase in arterial stiffness, as assessed by the pulse wave velocity measurement, is directly associated with the progression of atherosclerosis [170]. Schutte et al. (2011) reported that black individuals living in South Africa have higher PWV values, compared to their white counterparts [171]. Ngatchou et al. (2013) reported that HIV-infected individuals not receiving ART show a higher prevalence of aortic stiffness which may be associated with CVD [172].

Van Vonderen et al. (2009) have found that HIV-infected individuals have lower compliance and distensibility components in the carotid, femoral and brachial arteries [173]. Moreover, in a recent study it has been found that an increase in the PWV is associated with the use of efavirenz, traditional cardiovascular risk factors such as smoking, dyslipidaemia and systemic inflammation (CRP). These researchers have also suggested that ART may play a role in arterial stiffening in those living with HIV [174].

#### 5.3 Left ventricular hypertrophy

Left ventricular hypertrophy may reflect increased work load to physiological adaption of the heart [175]. An increase in left ventricular mass is the principal factor in developing LVH and hypertension a powerful determinant of LVH [175]. Hypertension may be directly associated with deficient levels of nitric oxide [176], leading to the inability of the endothelium to function normally in maintaining normal vascular tone in preventing onset of vascular damage. Furthermore, dysfunctional endothelium leads to functional changes in vasculature [177] and alterations in the function of the heart. According to literature, LVH is an independent predictor of cardiovascular events [178]. It has been concluded that LVH is more prevalent in black Africans compared to their white counterparts [179]. Havranek et al. (2008) indicated that LVH contributes to the risk of cardiovascular mortality in African individuals compared to their white counterparts [180].

Mansoor et al. (2009) found a positive association between LVH and HIV [181]. Several autopsy studies have reported that HIV infection directly affects myocardial cells and is associated with local cytokine release and other factors leading to inflammation in those living with HIV [182]. It has also been found that individuals exposed to protease inhibitors show higher intraventricular septal -and posterior wall thickness compared to those not receiving protease inhibitor [183] which may lead to a higher prevalence of LVH.

#### 5.4 Atherosclerosis

Atherosclerosis may be described as a chronic immune-inflammatory fibroproliferative (proliferation of fibroblasts) disease which occurs in blood vessels [184-186]. Endothelial cells, leukocytes and intimal smooth muscle cells are few of the major determinants of the development of atherosclerosis. Atherosclerotic lesions initially develop with leaky, activated and dysfunctional endothelium [187]. Recruitment of circulating monocytes and T- lymphocytes are of the earliest cellular responses in atherogenesis [185, 186]. The perseverance of these cellular responses seems to underlie disease progression [185, 186]. During disease progression, the immune-inflammatory response is accompanied by the fibro proliferative response which is mediated by intimal smooth muscle cells and they are responsible for the healing and repair process after arterial injury [188]. Smooth muscle cells produce collagen-rich

matrix compartments which offer stability to plaques with the aim of protecting them against plaque rupture and thrombosis [189].

Structural changes in blood vessels could be identified by conducting a measurement of the arterial intima-media thickness of the common carotid, femoral and brachial arteries by making use of the non-invasive B-mode ultrasonography [190, 191]. From the literature, it is evident that the intima-media thickness (IMT) is strongly associated with future CVD events especially stroke and myocardial infarction [192] and IMT is also seen as a marker for atherosclerosis. In a study by Okeahialam et al. (2011), Nigerian participants, who were apparently non-diabetic and non-hypertensive, showed a high CVD with regard to their IMT values [193]. Schoffelen et al. (2015) concluded that IMT was thicker in HIV-infected South African women compared to men and IMT associated better with CVD risk factors [194]. According to Mangili et al. (2011) IMT is associated with HIV mortality [195].

In a cross-sectional study conducted by Pen et al. (2013), it was found that the Framingham risk score displayed a significant correlation with coronary atherosclerotic burden [196]. However, Parra et al. (2010) did not find any correlation between the Framingham risk score and sub-clinical atherosclerosis in HIV-infected individuals [197].

## 6. Cardiovascular risk assessment

Cardiovascular risk assessment may be used to select participants for intervention [198]. Cardiovascular risk assessment involves "trying to predict the future adverse outcomes" by the implementation of risk score models [198]. All risk scores are designed in the same manner, where multiple clinical variables are documented in a population of individuals who are free of clinical CVD [198]. These individuals are then followed up over many years and the cardiovascular events well-documented [198]. Finally, the dataset is analysed to identify clinical variables which may be associated with cardiovascular outcomes [198]. The final product consists of a mathematical risk model and equation [198]. Even though risk models may be seen as reliable tools for clinicians to assess cardiovascular risk, algorithms generally do not account for risk variables which change over time. For instance, smoking is a categorical variable classified as "yes" or "no" with little regard to the duration and dose of smoking. A person classified as being a non-smoker may have been a chain smoker for 30 years before quitting or exposed to second-hand smoking.

According to Blom (2011), models used to analyse the data generated have been fairly simplistic [198] and more sophisticated techniques may improve the performance of the various algorithms [198]. Blom has also concluded that cardiovascular risk assessment is a science in "rapid evolution" [198].

# 7. The Framingham risk score

The Framingham study was the first study of its kind to investigate risk factors in a well-constructed longitudinal cohort. It consisted of 5209 men and women who were aged between 30 and 62, free of CVD and recruited in 1948 from Framingham, Massachusetts [199].

In 1961, a paper published by Kannel and colleagues on "Factors of Risk" enabled physicians as well as scientists to be more assured that blood pressure and other risk factors lead to CHD development [200].

The Framingham study reported other risk equations forming a link of common risk factors with CHD, stroke, fatal and non-fatal CVD [201, 202]. Moreover, these publications led to more refined screening techniques and served as the "primary" of many clinical guidelines existing worldwide [203].

After the identification of individual risk factors for CHD development, the Framingham study enhances the methods which assess one's overall risk established from numerous risk factors [204]. The risk of CVD development is strongly influenced by a cluster of risk factors present, notably CVD history, age, sex, diabetes, smoking, blood pressure and the concentrations of blood lipids [205].

In 1967, an analysis of the Framingham cohort was published and this analysis included seven risk factors to create a risk function for assessment in men and women aged between 30 to 62 years, namely age, cholesterol, systolic blood pressure, weight, haemoglobin, cigarette smoking and electrocardiography (ECG) (as evidence of LVH) [206].

In the 1990s improvements were made with the addition of HDL-C and extended duration for allowing risk assessment in an even older population (up to 74 years of age). A point scoring system [201] was added in particular for the clinicians who wanted to calculate the risk and in 1998, LVH was excluded [207]. Further changes included the impact of blood pressure treatment on CVD risk and tools for computing

risk scores were added on the web for the ease of use among clinicians. The additions made it possible for clinicians to determine CHD development without the use of a calculator. The risk factors currently included in the Framingham risk score [208] are: age, TC, smoking, HDL-C and SBP, which are included in two separate models, one for men and one for women.

Dada et al. (2016) found that the female participants of their study did not show a high Framingham risk [209]. Bergersen et al. (2004) demonstrated that twice as many HIV-infected individuals receiving ART, had a 10-year Framingham risk of more than 20%, compared to the HIV-free controls [210].

## 8. The Reynolds risk score

Cardiovascular risk also relates to family history, inflammation (CRP) and HbA1c (among diabetics) [211, 212]. These cardiovascular risk factors are included in the Reynolds risk score, which is an alternative global risk model developed in 2007 for men [211], and women [212]. This risk model also includes the same risk factors as proposed in the Framingham risk score [213].

The Reynolds risk score [212] was developed in the Women's Health Study Cohort. Recruitment of female participants took place in a nationwide cohort of the United States and these women were 45 years and older and did not have CVD. Cook et al. (2012) concluded that the Reynolds risk score was better calibrated than the Framingham risk score and showed improved discrimination overall and in black and white women [214]. The Reynolds risk score for men was developed in 2008 from a prospective cohort of 10000 American men where the risk score also included family history of myocardial infarction and high levels of CRP as risk factors [211].

Both the Reynolds and Framingham risk models received class one recommendations from the American Heart Association as well as the American College of Cardiology [215] and both these risk models were endorsed as part of the national guidelines for CVD prevention programme running in Canada [216].

Both these cardiovascular risk models were mainly developed for white men and women [208, 212, 217] however, in a recent study it has been found that the application of the Framingham risk model to a large black population is applicable to black individuals and not easily improved on, which suggests that no unique risk model needs to be developed for black individuals [218]. Mashinya et al. (2015) also

concluded that there is no need to develop a race/ethnicity specific risk model for HIV infected Africans [219].

To the best of my knowledge, there are no studies incorporating the Reynolds risk score to assess a 10-year risk in the HIV-infected population, especially in women [220].

# 9. Cardiovascular risk assessment and end-organ damage

The literature found that organ damage has an independent prognostic significance, irrespective of whether it involves the function and/or structure of the blood vessels, kidney, brain or heart [93]. It has also been shown that, once organ damage has been detected, the patients usually have a high cardiovascular risk [221], thus a chance of having either a fatal or a morbid cardiovascular event over a period of 10 years [222].

Table 1: Cardiovascular disease risk factors

Modifiable	Africans compared to	HIV-infected individuals
cardiovascular disease	white individuals	compared to HIV-free
risk factors		individuals
Lipids	↑ HDL-C [52]	↓ HDL-C [30]
	↓ TG [30]	↑ TG [30]
	↓ TC [52]	
Obesity	↑ Women [80]	↑ Women [81]
Hypertension	↑ Elderly [95]	↓ SBP and DBP [96]
Glucose	↑ Diabetes [98]	↑ Diabetes [104]
		Hyperglycaemia [104]
Smoking	↑ Smoking [116]	↑ Smoking [117, 118]
Alcohol	↑ GGT [127]	↑ GGT [129]
Inflammation	↑ CRP [151]	↑ CRP [146, 147]
	↑ IL-6 [151]	↑ IL-6 [146, 147]
CKD	↑ CKD [223]	↓ GFR [163]
		↑ Serum creatinine [163]
		Albuminuria [224]
LVH	LVH [179]	LVH [181]
Arterial stiffness	↑ PWV [171]	↑ PWV [174]
Atherosclerosis	↑ IMT [193]	↑ IMT in women [194]
Framingham risk score	↓ Women [209]	↑ FRS [210]
Reynolds Risk Score	No studies incorporating	No studies incorporating
	this risk score in Africans	this risk score in HIV-
		infected populations [220]

HDL-C, High-density lipoprotein cholesterol; TG, triglyceride; TC, total cholesterol; SBP, systolic blood pressure; DBP, diastolic blood pressure; GGT, gamma-glutamyltransferase; CRP, c-reactive protein; IL-6, interleukin-6; CKD, chronic kidney disease; GFR, glomerular filtration rate; LVH, left ventricular hypertrophy; PWV, pulse wave velocity; IMT, intima-media thickness; FRS, Framingham risk score. ↑ indicates an increase and ↓ indicates decreased levels.

## **Summary**

A dual burden of non-communicable (CVD) and communicable (HIV) diseases exist in South Africa and literature suggests that HIV-infected individuals have higher cardiovascular risk than HIV-free individuals. It is important to identify risk in this

population. This study will be conducted in longitudinal context and few studies in this regard exist.

Overall literature regarding CVD in the HIV-infected African population is sparse. Health care systems may benefit from this data in order to make important decisions concerning the CVD risk among HIV-infected black South Africans.

#### Aims

- To determine (1) the prevalence of cardiovascular disease risk markers and (2) cardiovascular disease risk with the Framingham and Reynolds risk score models in a cohort infected with HIV for at least ten years, compared to a HIV-free control group.
- To determine associations of the risk scores with measures of end-organ damage in this cohort.

# **Objectives**

- To determine the levels of cardiovascular disease risk markers among HIV-infected participants and HIV-free controls at baseline (2005) and follow-up (2015) and to compare the 10-year cardiovascular disease risk score between HIV-infected and HIV-free participants using the Framingham and Reynolds risk score models;
- To determine the prevalence of co-morbidities (Table 4) among this cohort after ten years;
- To determine whether these risk scores associate with markers of end-organ damage [sub-clinical atherosclerosis (IMT), arterial stiffness (PWV), ECG derived left ventricular hypertrophy (Cornell product) and chronic kidney disease (CrCl)] in the HIV-infected and HIV-free group.

## **Hypotheses**

- HIV-infected participants have a higher 10-year cardiovascular disease risk compared to HIV-free participants in both risk score models.
- A high cardiovascular disease risk score correlates positively with markers of end-organ damage [increased sub-clinical atherosclerosis (IMT), increased

arterial stiffness (PWV), left ventricular hypertrophy (LVH) and chronic kidney disease (CrCl)] in the HIV-infected group.

## References

- 1. Marquez PV, Farrington JL. The challenge of non-communicable diseases and road traffic injuries in sub-Saharan Africa. An overview. Washington, DC: The World Bank. 2013.
- World Health Organization (WHO). Non-communicable diseases [Internet].
   [cited 2015]. Available from: http://www.who.int/mediacentre/factsheets/fs355/en/.
- 3. Njelekela MA, Mpembeni R, Muhihi A, Mligiliche NL, Spiegelman D, Hertzmark E, et al. Gender-related differences in the prevalence of cardiovascular disease risk factors and their correlates in urban Tanzania. BMC Cardiovasc Disord 2009;9:30.
- 4. Liao Y, Cooper RS. Continued adverse trends in coronary heart disease mortality among blacks, 1980-91. Public Health Reports 1995;110(5):572.
- 5. Williams DW, Eugenin EA, Calderon TM, Berman JW. Monocyte maturation, HIV susceptibility, and transmigration across the blood brain barrier are critical in HIV neuropathogenesis. J Leukoc Biol 2012;91(3):401-415.
- 6. World Health Organization (WHO). HIV/AIDS [Internet]. 2016 [cited 2016]. Available from: http://www.who.int/mediacentre/factsheets/fs360/en/.
- 7. Joined United Nations Programme on HIV/AIDS (UNAIDS). Global AIDS update [Internet]. 2016 [cited 2016]. Available from: http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf.
- 8. Mills EJ, Barnighausen T, Negin J. HIV and aging-preparing for the challenges ahead. N Engl J Med 2012;366(14):1270-1273.
- 9. Mathers CD, Loncar D. Projections of global mortality and burden of disease from 2002 to 2030. PLoS Med 2006;3(11):442.
- 10. Girard MP, Osmanov SK, Kieny MP. A review of vaccine research and development: The human immunodeficiency virus (HIV). Vaccine 2006;24(19):4062-4081.

- 11. De Coul ELO, Prins M, Cornelissen M, van der Schoot A, Boufassa F, Brettle RP, et al. Using phylogenetic analysis to trace HIV-1 migration among Western European injecting drug users seroconverting from 1984 to 1997. AIDS 2001;15(2):257-266.
- 12. Roberts JD, Bebenek K, Kunkel TA. The accuracy of reverse transcriptase from HIV-1. Science 1988;242(4882):1171-1173.
- 13. Ho DD, Neumann AU, Perelson AS, Chen W, Leonard JM, Markowitz M. Rapid turnover of plasma virions and CD4 lymphocytes in HIV-1 infection. Nature 1995;373(6510):123-126.
- 14. Michael NL. Host genetic influences on HIV-1 pathogenesis. Curr Opin Immunol 1999;11(4):466-474.
- 15. Temin HM. Retrovirus variation and reverse transcription: Abnormal strand transfers result in retrovirus genetic variation. Proc Natl Acad Sci 1993;90(15):6900-6903.
- 16. Ayouba A, Souquières S, Njinku B, Martin PM, Müller-Trutwin MC, Roques P, et al. HIV-1 group n among HIV-1-seropositive individuals in Cameroon. AIDS 2000;14(16):2623-2625.
- 17. Gürtler LG, Hauser PH, Eberle J, Von Brunn A, Knapp S, Zekeng L, et al. A new subtype of human immunodeficiency virus type 1 (MVP-5180) from Cameroon. J Virol 1994;68(3):1581-1585.
- 18. Simon F, Mauclère P, Roques P, Loussert-Ajaka I, Müller-Trutwin MC, Saragosti S, et al. Identification of a new human immunodeficiency virus type 1 distinct from group M and group O. Nat Med 1998;4(9):1032-1037.
- 19. Peeters M. The genetic variability of HIV-1 and its implications. Transfus Clin Biol 2001;8(3):222-225.
- 20. Jacobs GB, de Beer C, Fincham JE, Adams V, Dhansay MA, van Rensburg EJ, et al. Serotyping and genotyping of HIV-1 infection in residents of Khayelitsha, Cape Town, South Africa. J Med Virol 2006;78(12):1529-1536.
- 21. Freire E. Overcoming HIV-1 resistance to protease inhibitors. Drug Discov Today: Dis Mech 2006;3(2):281-286.

- 22. Gaschen B, Taylor J, Yusim K, Foley B, Gao F, Lang D, et al. Diversity considerations in HIV-1 vaccine selection. Science 2002;296(5577):2354-2360.
- 23. Manavi K. A review on infection with human immunodeficiency virus. Best Pract Res Clin Obstet Gynaecol 2006;20(6):923-940.
- 24. Betts MR, Ambrozak DR, Douek DC, Bonhoeffer S, Brenchley JM, Casazza JP, et al. Analysis of total human immunodeficiency virus (HIV)-specific CD4(+) and CD8(+) T-cell responses: Relationship to viral load in untreated HIV infection. J Virol 2001;75(24):11983-11991.
- 25. Papagno L, Appay V, Sutton J, Rostron T, Gillespie GM, Ogg GS, et al. Comparison between HIV- and CMV-specific T-cell responses in long-term HIV infected donors. Clin Exp Immunol 2002;130(3):509-517.
- 26. Merrill JE, Koyanagi Y, Chen IS. Interleukin-1 and tumor necrosis factor alpha can be induced from mononuclear phagocytes by human immunodeficiency virus type 1 binding to the CD4 receptor. J Virol 1989;63(10):4404-4408.
- 27. Rieckmann P, Poli G, Fox CH, Kehrl JH, Fauci AS. Recombinant gp120 specifically enhances tumor necrosis factor alpha production and IG secretion in B lymphocytes from HIV-infected individuals but not from seronegative donors. J lmmunol 1991;147(9):2922-2927.
- 28. Lee C, Liu QH, Tomkowicz B, Yi Y, Freedman BD, Collman RG. Macrophage activation through CCR5- and CXCR-mediated gp120 elicited signaling pathways. J Leukoc Biol 2003;74(5):676-682.
- 29. Baker JV, Lundgren JD. Cardiovascular implications from untreated human immunodeficiency virus infection. Eur Heart J 2011;32(8):945-951.
- 30. Dube MP, Lipshultz SE, Fichtenbaum CJ, Greenberg R, Schecter AD, Fisher SD. Effects of HIV infection and antiretroviral therapy on the heart and vasculature. Circulation 2008;118(2):36-40.
- 31. Rajasuriar R, Wright E, Lewin SR. Impact of antiretroviral therapy (ART) timing on chronic immune activation/inflammation and end-organ damage. Curr Opin HIV/AIDS 2015;10(1):35.

- 32. Cockerill GW, Rye KA, Gamble JR, Vadas MA, Barter PJ. High-density lipoproteins inhibit cytokine-induced expression of endothelial cell adhesion molecules. Arterioscler Thromb Vasc Biol 1995;15(11):1987-1994.
- 33. Cockerill GW, Saklatvala J, Ridley SH, Yarwood H, Miller NE, Oral B, et al. High-density lipoproteins differentially modulate cytokine-induced expression of eselectin and cyclooxygenase-2. Arterioscler Thromb Vasc Biol 1999;19(4):910-917.
- 34. Baker PW, Rye KA, Gamble JR, Vadas MA, Barter PJ. Ability of reconstituted high density lipoproteins to inhibit cytokine-induced expression of vascular cell adhesion molecule-1 in human umbilical vein endothelial cells. J Lipid Res 1999;40(2):345-353.
- 35. Ginsberg HN, Bonds DE, Lovato LC, Crouse JR, Elam MB, Linz PE, et al. Evolution of the lipid trial protocol of the action to control cardiovascular risk in diabetes (ACCORD) trial. Am J Cardiol 2007;99(12):56-67.
- 36. Savès M, Chêne G, Ducimetière P, Leport C, Le Moal G, Amouyel P, et al. Risk factors for coronary heart disease in patients treated for human immunodeficiency virus infection compared with the general population. Clin Infect Dis 2003;37(2):292-298.
- 37. High KP, Effros RB, Fletcher CV, Gebo K, Halter JB, Hazzard WR, et al. Workshop on HIV infection and aging: What is known and future research directions. Clin Infect Dis 2008;47(4):542-553.
- 38. Petoumenos K, Reiss P, Ryom L, Rickenbach M, Sabin CA, El-Sadr W, et al. Increased risk of cardiovascular disease (CVD) with age in HIV-positive men: A comparison of the D:A:D CVD risk equation and general population CVD risk equations. HIV Med 2014;15(10):595-603.
- 39. Franceschi C, Bonafe M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann N Y Acad Sci 2000;908:244-254.
- 40. Avolio AP, Chen SG, Wang RP, Zhang CL, Li MF, O'Rourke MF. Effects of aging on changing arterial compliance and left ventricular load in a Northern Chinese urban community. Circulation 1983;68(1):50-58.
- 41. Lee HY, Oh BH. Aging and arterial stiffness. Circ J 2010;74(11):2257-2262.

- 42. Desai S, Landay A. Early immune senescence in HIV disease. Curr HIV/AIDS Rep 2010;7(1):4-10.
- 43. Deeks SG. Immune dysfunction, inflammation, and accelerated aging in patients on antiretroviral therapy. Top HIV Med 2009;17(4):118-123.
- 44. Deeks SG. HIV infection, inflammation, immunosenescence, and aging. Annu Rev Med 2011;62:141-155.
- 45. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. J Clin Endocrinol Metab 2007;92(7):2506-2512.
- 46. Van Rooyen JM, Kruger HS, Huisman HW, Wissing MP, Margetts BM, Venter CS, et al. An epidemiological study of hypertension and its determinants in a population in transition: The THUSA study. J Hum Hypertens 2000;14(12):779-787.
- 47. Sliwa K, Wilkinson D, Hansen C, Ntyintyane L, Tibazarwa K, Becker A, et al. Spectrum of heart disease and risk factors in a black urban population in South Africa (The Heart of Soweto study): A cohort study. Lancet 2008;371(9616):915-922.
- 48. Major outcomes in high-risk hypertensive patients randomized to angiotensin-converting enzyme inhibitor or calcium channel blocker vs diuretic: The Antihypertensive and Lipid-lowering treatment to prevent Heart Attack Trial (ALLHAT). JAMA 2002;288(23):2981-2997.
- 49. Centers for Disaease Control and Prevention. HIV among African Americans [Internet]. 2017 [cited 2017]. Available from: https://www.cdc.gov/nchhstp/newsroom/docs/factsheets/cdc-hiv-aa-508.pdf.
- 50. Freiberg MS, Chang CC, Kuller LH, Skanderson M, Lowy E, Kraemer KL, et al. HIV infection and the risk of acute myocardial infarction. JAMA Intern Med 2013;173(8):614-622.
- 51. Grunfeld C, Delaney JA, Wanke C, Currier JS, Scherzer R, Biggs ML, et al. Pre-clinical atherosclerosis due to HIV infection: Carotid intima-medial thickness measurements from The FRAM study. AIDS 2009;23(14):1841.
- 52. Steyn K, Damasceno A. Lifestyle and related risk factors for chronic diseases. Disease and mortality in sub-Saharan Africa 2006;2:247-265.

- 53. Fourie CM, van Rooyen JM, Kruger A, Schutte AE. Lipid abnormalities in a never-treated HIV-1 subtype C-infected African population. Lipids 2010;45(1):73-80.
- 54. Godsland IF, Johnston DG, Chaturvedi N. Mechanisms of disease: Lessons from ethnicity in the role of triglyceride metabolism in ischemic heart disease. Nat Clin Pract Endocrinol Metab 2007;3(7):530-538.
- 55. Adiels M, Olofsson SO, Taskinen MR, Boren J. Overproduction of very low-density lipoproteins is the hallmark of the dyslipidemia in the metabolic syndrome. Arterioscler Thromb Vasc Biol 2008;28(7):1225-1236.
- 56. Choi SH, Ginsberg HN. Increased very low density lipoprotein (VLDL) secretion, hepatic steatosis, and insulin resistance. Trends Endocrinol Metab 2011;22(9):353-363.
- 57. Miller M, Stone NJ, Ballantyne C, Bittner V, Criqui MH, Ginsberg HN, et al. Triglycerides and cardiovascular disease a scientific statement from the American Heart Association. Circulation 2011;123(20):2292-2333.
- 58. Third report of the National Cholesterol Education Program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel iii) final report. Circulation 2002;106(25):3143-3421.
- 59. Pereira AC, Sposito AC, Mota GF, Cunha RS, Herkenhoff FL, Mill JG, et al. Endothelial nitric oxide synthase gene variant modulates the relationship between serum cholesterol levels and blood pressure in the general population: New evidence for a direct effect of lipids in arterial blood pressure. Atherosclerosis 2006;184(1):193-200.
- 60. Castelli WP, Anderson K, Wilson PW, Levy D. Lipids and risk of coronary heart disease. The Framingham study. Ann Epidemiol 1992;2(1-2):23-28.
- 61. Centers for Disaease Control and Prevention. LDL and HDL Cholesterol: "Bad" and "Good" Cholesterol [Internet]. 2017 [cited 2017]. Available from: https://www.cdc.gov/cholesterol/ldl hdl.htm.
- 62. Pyorala K, Pedersen TR, Kjekshus J, Faergeman O, Olsson AG, Thorgeirsson G. Cholesterol lowering with Simvastatin improves prognosis of diabetic patients with coronary heart disease. A subgroup analysis of the Scandinavian Simvastatin Survival study (4s). Diabetes Care 1997;20(4):614-620.

- 63. Sacks FM, Pfeffer MA, Moye LA, Rouleau JL, Rutherford JD, Cole TG, et al. The effect of Pravastatin on coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and recurrent events trial investigators. N Engl J Med 1996;335(14):1001-1009.
- 64. Downs JR, Clearfield M, Weis S, Whitney E, Shapiro DR, Beere PA, et al. Primary prevention of acute coronary events with Lovastatin in men and women with average cholesterol levels: Results of AFCAPS/TEXCAPS. Air force/Texas Coronary Atherosclerosis Prevention study. JAMA 1998;279(20):1615-1622.
- 65. Howard BV, Robbins DC, Sievers ML, Lee ET, Rhoades D, Devereux RB, et al. LDL cholesterol as a strong predictor of coronary heart disease in diabetic individuals with insulin resistance and low LDL: The Strong Heart study. Arterioscler Thromb Vasc Biol 2000;20(3):830-835.
- 66. Castelli WP. The triglyceride issue: A view from Framingham. Am Heart J 1986;112(2):432-437.
- 67. Castelli WP. Epidemiology of triglycerides: A view from Framingham. Am J Cardiol 1992;70(19):3-9.
- 68. Hokanson JE, Austin MA. Plasma triglyceride level is a risk factor for cardiovascular disease independent of high-density lipoprotein cholesterol level: A meta-analysis of population-based prospective studies. J Cardiovasc Risk 1996;3(2):213-219.
- 69. Schulte H, Cullen P, Assmann G. Obesity, mortality and cardiovascular disease in The Munster Heart study (PROCAM). Atherosclerosis 1999;144(1):199-209.
- 70. Sarwar N, Danesh J, Eiriksdottir G, Sigurdsson G, Wareham N, Bingham S, et al. Triglycerides and the risk of coronary heart disease: 10,158 incident cases among 262,525 participants in 29 Western prospective studies. Circulation 2007;115(4):450-458.
- 71. Talayero BG, Sacks FM. The role of triglycerides in atherosclerosis. Curr Cardiol Rep 2011;13(6):544-552.
- 72. Grunfeld C, Pang M, Doerrler W, Shigenaga JK, Jensen P, Feingold KR. Lipids, lipoproteins, triglyceride clearance, and cytokines in human immunodeficiency

- virus infection and the acquired immunodeficiency syndrome. J Clin Endocrinol Metab 1992;74(5):1045-1052.
- 73. Feingold KR, Krauss RM, Pang M, Doerrler W, Jensen P, Grunfeld C. The hypertriglyceridemia of acquired immunodeficiency syndrome is associated with an increased prevalence of low density lipoprotein subclass pattern B. J Clin Endocrinol Metab 1993;76(6):1423-1427.
- 74. Dube MP, Stein JH, Aberg JA, Fichtenbaum CJ, Gerber JG, Tashima KT, et al. Guidelines for the evaluation and management of dyslipidemia in human immunodeficiency virus (HIV)-infected adults receiving antiretroviral therapy: Recommendations of the HIV medical association of The Infectious Disease Society of America and the Adult AIDS Clinical Trials Group. Clin Infect Dis 2003;37(5):613-627.
- 75. Must A, Spadano J, Coakley EH, Field AE, Colditz G, Dietz WH. The disease burden associated with overweight and obesity. JAMA 1999;282(16):1523-1529.
- 76. Asfaw A. The effects of obesity on doctor-diagnosed chronic diseases in Africa: Empirical results from Senegal and South Africa. J Public Health Policy 2006;27(3):250-264.
- 77. Cappuccio FP, Kerry SM, Adeyemo A, Luke A, Amoah AG, Bovet P, et al. Body size and blood pressure: An analysis of Africans and the African diaspora Epidemiology 2008;19(1):38-46.
- 78. Swinburn BA, Sacks G, Hall KD, McPherson K, Finegood DT, Moodie ML, et al. The Global Obesity Pandemic: Shaped by global drivers and local environments. Lancet 2011;378(9793):804-814.
- 79. Adeboye B, Bermano G, Rolland C. Obesity and its health impact in Africa: A systematic review. Cardiovasc J Afr 2012;23(9):512-521.
- 80. Puoane T, Steyn K, Bradshaw D, Laubscher R, Fourie J, Lambert V, et al. Obesity in South Africa: The South African Demographic and Health Survey. Obes Res 2002;10(10):1038-1048.
- 81. McCormick CL, Francis AM, Iliffe K, Webb H, Douch CJ, Pakianathan M, et al. Increasing obesity in treated female HIV patients from sub-Saharan Africa: Potential causes and possible targets for intervention Front Immunol 2014;5:507.

- 82. Finucane MM, Stevens GA, Cowan MJ, Danaei G, Lin JK, Paciorek CJ, et al. National, regional, and global trends in body-mass index since 1980: Systematic analysis of health examination surveys and epidemiological studies with 960 country-years and 9.1 million participants. Lancet 2011;377(9765):557-567.
- 83. Bastien M, Poirier P, Lemieux I, Despres JP. Overview of epidemiology and contribution of obesity to cardiovascular disease. Prog Cardiovasc Dis 2014;56(4):369-381.
- 84. Dudina A, Cooney MT, Bacquer DD, Backer GD, Ducimetiere P, Jousilahti P, et al. Relationships between body mass index, cardiovascular mortality, and risk factors: A report from the score investigators. Eur J Cardiovasc Prev Rehabil 2011;18(5):731-742.
- 85. Czernichow S, Kengne AP, Stamatakis E, Hamer M, Batty GD. Body mass index, waist circumference and waist-hip ratio: Which is the better discriminator of cardiovascular disease mortality risk?: Evidence from an individual-participant meta-analysis of 82 864 participants from nine cohort studies. Obes Rev 2011;12(9):680-687.
- 86. Yusuf S, Hawken S, Ôunpuu S, Dans T, Avezum A, Lanas F, et al. Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (The Interheart study): Case-control study. Lancet 2004;364(9438):937-952.
- 87. De Koning L, Merchant AT, Pogue J, Anand SS. Waist circumference and waist-to-hip ratio as predictors of cardiovascular events: Meta-regression analysis of prospective studies. Eur Heart J 2007;28(7):850-856.
- 88. Ogunmola OJ, Oladosu OY, Olamoyegun AM. Association of hypertension and obesity with HIV and antiretroviral therapy in a rural tertiary health center in Nigeria: A cross-sectional cohort study. Vasc Health Risk Manag 2014;10:129-137.
- 89. Barth RE, van der Meer JT, Hoepelman AI, Schrooders PA, van de Vijver DA, Geelen SP, et al. Effectiveness of highly active antiretroviral therapy administered by general practitioners in rural South Africa. Eur J Clin Microbiol Infect Dis 2008;27(10):977-984.

- 90. Ezzati M, Vander Hoorn S, Lawes CM, Leach R, James WP, Lopez AD, et al. Rethinking the "diseases of affluence" paradigm: Global patterns of nutritional risks in relation to economic development. PLoS Med 2005;2(5):133.
- 91. Lawes CMM, Hoorn SV, Rodgers A. Global burden of blood-pressure-related disease, 2001. The Lancet 371(9623):1513-1518.
- 92. Schmieder R, Messerli F. Hypertension and the heart. J Hum Hypertens 2000;14(10/11):597.
- 93. Mancia G, Laurent S, Agabiti-Rosei E, Ambrosioni E, Burnier M, Caulfield MJ, et al. Reappraisal of European guidelines on hypertension management: A European society of hypertension task force document. Blood Press 2009;18(6):308-347.
- 94. Yang F, Qian D, Hu D. Prevalence, awareness, treatment, and control of hypertension in the older population: Results from the multiple national studies on ageing. J Am Soc Hypertens 2016;10(2):140-148.
- 95. Lloyd-Sherlock P, Beard J, Minicuci N, Ebrahim S, Chatterji S. Hypertension among older adults in low- and middle-income countries: Prevalence, awareness and control. Int J Epidemiol 2014;43(1):116-128.
- 96. Dillon DG, Gurdasani D, Riha J, Ekoru K, Asiki G, Mayanja BN, et al. Association of HIV and ART with cardiometabolic traits in sub-Saharan Africa: A systematic review and meta-analysis. Int J Epidemiol 2013;42(6):1754-1771.
- 97. Vos T, Barber RM, Bell B, Bertozzi-Villa A, Biryukov S, Bolliger I, et al. Global, regional, and national incidence, prevalence, and years lived with disability for 301 acute and chronic diseases and injuries in 188 countries, 1990 & 2013;2013: A systematic analysis for the global burden of disease study 2013. Lancet 386(9995):743-800.
- 98. Atlas ID. 6th edn. International diabetes federation 2013. 2015.
- 99. Kengne AP, Echouffo-Tcheugui JB, Sobngwi E, Mbanya JC. New insights on diabetes mellitus and obesity in Africa-part 1: Prevalence, pathogenesis and comorbidities. Heart 2013;99(14):979-983.

- 100. Stamler J, Vaccaro O, Neaton JD, Wentworth D. Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the multiple risk factor intervention trial. Diabetes Care 1993;16(2):434-444.
- 101. Woodward M, Zhang X, Barzi F, Pan W, Ueshima H, Rodgers A, et al. The effects of diabetes on the risks of major cardiovascular diseases and death in the Asia-Pacific region. Diabetes Care 2003;26(2):360-366.
- 102. Pickup JC, Mattock MB, Chusney GD, Burt D. NIDDM as a disease of the innate immune system: Association of acute-phase reactants and interleukin-6 with metabolic syndrome X. Diabetologia 1997;40(11):1286-1292.
- 103. Festa A, D'Agostino R, Jr., Howard G, Mykkanen L, Tracy RP, Haffner SM. Chronic subclinical inflammation as part of the insulin resistance syndrome: The Insulin Resistance Atherosclerosis study (IRAS). Circulation 2000;102(1):42-47.
- 104. Brown TT, Cole SR, Li X, et al. Antiretroviral therapy and the prevalence and incidence of diabetes mellitus in The Multicenter AIDS Cohort study. Arch Intern Med 2005;165(10):1179-1184.
- 105. Bansal B, Carvalho P, Mehta Y, Yadav J, Sharma P, Mithal A, et al. Prognostic significance of glycemic variability after cardiac surgery. Journal of Diabetes and Its Complications 30(4):613-617.
- 106. Motta M, Bennati E, Cardillo E, Ferlito L, Malaguarnera M. The value of glycosylated hemoglobin (HbA1c) as a predictive risk factor in the diagnosis of diabetes mellitus (DM) in the elderly. Arch Gerontol Geriatr 50(1):60-64.
- 107. International expert committee report on the role of the A1c assay in the diagnosis of diabetes. Diabetes Care 2009;32(7):1327-1334.
- 108. Sacks DB, Arnold M, Bakris GL, Bruns DE, Horvath AR, Kirkman MS, et al. Executive summary: Guidelines and recommendations for laboratory analysis in the diagnosis and management of diabetes mellitus. Clin Chem 2011;57(6):793-798.
- 109. Adams RJ, Appleton SL, Hill CL, Wilson DH, Taylor AW, Chittleborough CR, et al. Independent association of HbA1c and incident cardiovascular disease in people without diabetes. Obesity 2009;17(3):559-563.

- 110. Misra R, Chandra P, Riechman SE, Long DM, Shinde S, Pownall HJ, et al. Relationship of ethnicity and CD4 count with glucose metabolism among HIV patients on highly-active antiretroviral therapy (HAART). BMC Endocr Disord 2013;13(1):1-19.
- 111. Teo KK, Ounpuu S, Hawken S, Pandey MR, Valentin V, Hunt D, et al. Tobacco use and risk of myocardial infarction in 52 countries in the Interheart study: A case-control study. Lancet 2006;368(9536):647-658.
- 112. Imamura H, Teshima K, Miyamoto N, Shirota T. Cigarette smoking, high-density lipoprotein cholesterol subfractions, and lecithin: Cholesterol acyltransferase in young women. Metabolism 2002;51(10):1313-1316.
- 113. Howard G, Wagenknecht LE, Burke GL, Diez-Roux A, Evans GW, McGovern P, et al. Cigarette smoking and progression of atherosclerosis: The Atherosclerosis Risk In Communities (ARIC) study. JAMA 1998;279(2):119-124.
- 114. Craig WY, Palomaki GE, Haddow JE. Cigarette smoking and serum lipid and lipoprotein concentrations: An analysis of published data. BMJ 1989;298(6676):784-788.
- 115. Nishio E, Watanabe Y. Cigarette smoke extract inhibits plasma paraoxonase activity by modification of the enzyme's free thiols. Biochem Biophys Res Commun 1997;236(2):289-293.
- 116. Reddy P, Zuma K, Shisana O, Jonas K, Sewpaul R. Prevalence of tobacco use among adults in South Africa: Results from the first South African National Health and Nutrition Examination Survey. S Afr Med J 2015;105(8):648-655.
- 117. Tesoriero JM, Gieryic SM, Carrascal A, Lavigne HE. Smoking among HIV positive New Yorkers: Prevalence, frequency, and opportunities for cessation. AIDS Behav 2010;14(4):824-835.
- 118. Nahvi S, Cooperman NA. Review: The need for smoking cessation among HIV-positive smokers. AIDS Educ Prev 2009;21:14-27.
- 119. Rasmussen LD, Helleberg M, May MT, Afzal S, Kronborg G, Larsen CS, et al. Myocardial infarction among Danish HIV-infected individuals: Population-attributable fractions associated with smoking. Clin Infect Dis 2015;60(9):1415-1423.

- 120. Helleberg M, Afzal S, Kronborg G, Larsen CS, Pedersen G, Pedersen C, et al. Mortality attributable to smoking among HIV-1-infected individuals: A Nationwide, Population-based Cohort study. Clin Infect Dis 2013;56(5):727-734.
- 121. Braithwaite RS, Nucifora KA, Kessler J, Toohey C, Mentor SM, Uhler LM, et al. Impact of interventions targeting unhealthy alcohol use in Kenya on HIV transmission and AIDS-related deaths. Alcohol Clin Exp Res 2014;38(4):1059-1067.
- 122. I keda MLR, Barcellos NT, Alencastro PR, Wolff FH, Brandão A, Fuchs FD, et al. Association of blood pressure and hypertension with alcohol consumption in HIV-infected white and non-white patients. Sci World J 2013;2013:169825.
- 123. Long Y, Zeng F, Shi J, Tian H, Chen T. Gamma-glutamyltransferase predicts increased risk of mortality: A systematic review and meta-analysis of prospective observational studies. Free Rad Res 2014;48(6):716-728.
- 124. Kloner RA, Rezkalla SH. To drink or not to drink? That is the question. Circulation 2007;116(11):1306-1317.
- 125. Laurent S. Arterial stiffness in arterial hypertension. Curr Hypertens Rep 2006;8(3):179-180.
- 126. Noborisaka Y, Ishizaki M, Yamazaki M, Honda R, Yamada Y. Elevated serum gamma-glutamyltransferase (GGT) activity and the development of chronic kidney disease (CKD) in cigarette smokers. Nephro-urology monthly 2013;5(5):967-973.
- 127. Stewart SH, Connors GJ, Hutson A. Ethnicity and gamma-glutamyltransferase in men and women with alcohol use disorders. Alcohol Alcohol 2007;42(1):24-27.
- 128. Lee S-A, Wen W, Xiang Y-B, Barnes S, Liu D, Cai Q, et al. Assessment of dietary isoflavone intake among middle-aged Chinese men. J Nutr 2007;137(4):1011-1016.
- 129. Mokondjimobe E, Longo-Mbenza B, Mampouya-Arrouse P, Parra HJ, Diatewa M. Inflammatory status hepatic enzymes and serum creatinine in HIV-, HIV+ and HIV-TB co-infected adult central Africans. Intern J Med 2012;5:961-965.
- 130. Wannamethee S, Lennon L, Shaper A. The value of gamma-glutamyltransferase in cardiovascular risk prediction in men without diagnosed cardiovascular disease or diabetes. Atherosclerosis 2008;201(1):168-175.

- 131. Niemelä O. Biomarkers in alcoholism. Clinica Chimica Acta. 2007;377(1):39-49.
- 132. Sasaki S, Yoshioka E, Saijo Y, Kita T, Okada E, Tamakoshi A, et al. Relation between alcohol consumption and arterial stiffness: A cross-sectional study of middle-aged Japanese women and men. Alcohol 2013;47(8):643-649.
- 133. Pepys MB, Hirschfield GM. C-reactive protein: A critical update. Journal of Clinical Investigation. 2003;111(12):1805.
- 134. Pepys MB, Hirschfield GM. C-reactive protein: A critical update. J Clin Invest 2003;111(12):1805-1812.
- 135. McDermott MM, Green D, Greenland P, Liu K, Criqui MH, Chan C, et al. Relation of levels of hemostatic factors and inflammatory markers to the ankle brachial index. Am J Cardiol 2003;92(2):194-199.
- 136. Galante A, Pietroiusti A, Vellini M, Piccolo P, Possati G, De Bonis M, et al. Creactive protein is increased in patients with degenerative aortic valvular stenosis. J Am Coll Cardiol 2001;38(4):1078-1082.
- 137. Gunduz H, Akdemir R, Binak E, Tamer A, Keser N, Uyan C. Can serum lipid and CRP levels predict the" severity" of aortic valve stenosis? Acta cardiologica 2003;58(4):321-326.
- 138. Danesh J, Whincup P, Walker M, Lennon L, Thomson A, Appleby P, et al. Low grade inflammation and coronary heart disease: Prospective study and updated meta-analyses. BMJ 2000;321(7255):199-204.
- 139. Ross R. Atherosclerosis-an inflammatory disease. J Engl Med 1999;340: 2115-2126.
- 140. Sung JH, Lee JE, Samdarshi TE, Nagarajarao HS, Taylor JK, Agrawal KK, et al. C-reactive protein and subclinical cardiovascular disease among African Americans:(The Jackson Heart study). J Cardiovasc Med 2014;15(5):371.
- 141. Fauci AS, Pantaleo G, Stanley S, Weissman D. Immunopathogenic mechanisms of HIV infection. Ann Intern Med 1996;124(7):654-663.

- 142. Lau B, Sharrett AR, Kingsley LA, Post W, Palella FJ, Visscher B, et al. C-reactive protein is a marker for human immunodeficiency virus disease progression. Arch Intern Med 2006;166(1):64-70.
- 143. Zulu I, Hassan G, Njobvu L, Dhaliwal W, Sianongo S, Kelly P. Cytokine activation is predictive of mortality in Zambian patients with AIDS-related diarrhoea. BMC Infect Dis 2008;8(1):156.
- 144. Melchior J-C, Niyongabo T, Henzel D, Durack-Bown I, Henri S-C, Boulier A. Malnutrition and wasting, immunodepression, and chronic inflammation as independent predictors of survival in HIV-infected patients. Nutrition 1999;15(11):865-869.
- 145. Volanakis JE. Human c-reactive protein: Expression, structure, and function. Mol Immunol 2001;38(2):189-197.
- 146. Feldman JG, Goldwasser P, Holman S, DeHovitz J, Minkoff H. C-reactive protein is an independent predictor of mortality in women with HIV-1 infection. J Acquir Immune Defic Syndr 2003;32(2):210-214.
- 147. Reingold J, Wanke C, Kotler D, Lewis C, Tracy R, Heymsfield S, et al. Association of HIV infection and HIV/HCV coinfection with c-reactive protein levels: The Fat Redistribution and Metabolic change in HIV infection (fram) study. J Acquir Immune Defic Syndr 2008;48(2):142-148.
- 148. Lau B, Sharrett AR, Kingsley LA, Post W, Palella FJ, Visscher B, et al. C-reactive protein is a marker for human immunodeficiency virus disease progression. Arch Intern Med 2006;166(1):64-70.
- 149. Janeway CA TP, Walport M, Schlomchik MJ. 5th ed. New York: Garland Science, 2001.
- 150. Dolan SE, Hadigan C, Killilea KM, Sullivan MP, Hemphill L, Lees RS, et al. Increased cardiovascular disease risk indices in HIV-infected women. J Acquir Immune Defic Syndr 2005;39(1):44-54.
- 151. Mokhaneli MC, Fourie CM, Botha S, Mels CM. The association of oxidative stress with arterial compliance and vascular resistance in a bi-ethnic population: The SABPA study. Free Radic Res 2016;50(8):920-928.

- 152. Masiá M, Bernal E, Padilla S, Graells ML, Jarrín I, Almenar MV, et al. The role of c-reactive protein as a marker for cardiovascular risk associated with antiretroviral therapy in HIV-infected patients. Atherosclerosis 2007;195(1):167-171.
- 153. Saltevo J, Vanhala M, Kautiainen H, Kumpusalo E, Laakso M. Gender differences in c-reactive protein, interleukin-1 receptor antagonist and adiponectin levels in the metabolic syndrome: A population-based study. Diabet Med 2008;25(6):747-750.
- 154. Fried LF, Shlipak MG, Crump C, Kronmal RA, Bleyer AJ, Gottdiener JS, et al. Renal insufficiency as a predictor of cardiovascular outcomes and mortality in elderly individuals. J Am Coll Cardiol 2003;41(8):1364-1372.
- 155. Manjunath G, Tighiouart H, Ibrahim H, MacLeod B, Salem DN, Griffith JL, et al. Level of kidney function as a risk factor for atherosclerotic cardiovascular outcomes in the community. J Am Coll Cardiol 2003;41(1):47-55.
- 156. Winston JA, Klotman ME, Klotman PE. HIV-associated nephropathy is a late, not early, manifestation of HIV-1 infection. Kidney Int 1999;55(3):1036-1040.
- 157. Levin ML, Palella F, Shah S, Lerma E, Butter J, Kanwar YS. HIV-associated nephropathy occurring before HIV antibody seroconversion. Am J Kidney Dis 2001;37(5):39.
- 158. Kopp JB, Winkler C. HIV-associated nephropathy in African Americans. Kidney Int 2003;63:43-9.
- 159. Lucas GM, Lau B, Atta MG, Fine DM, Keruly J, Moore RD. Chronic kidney disease incidence, and progression to end-stage renal disease, in HIV-infected individuals: A tale of two races. J Infect Dis 2008;197(11):1548-1557.
- 160. Stevens LA, Coresh J, Greene T, Levey AS. Assessing kidney function—measured and estimated glomerular filtration rate. N Engl J Med 2006;354(23):2473-2483.
- 161. Cockcroft DW, Gault MH. Prediction of creatinine clearance from serum creatinine. Nephron 1976;16(1):31-41.
- 162. Collins AJ, Li S, Gilbertson DT, Liu J, Chen S-C, Herzog CA. Chronic kidney disease and cardiovascular disease in the medicare population: Management of

- comorbidities in kidney disease in the 21st century: Anemia and bone disease. Kidney Int 2003;64:24-31.
- 163. Jabłonowska E, Małolepsza E, Wójcik K. The assessment of renal function in HIV-positive patients before the introduction of antiretroviral therapy. HIV/AIDS Review 2010;9(2):45-47.
- 164. Winston JA, Bruggeman LA, Ross MD, Jacobson J, Ross L, D'agati VD, et al. Nephropathy and establishment of a renal reservoir of HIV type 1 during primary infection. N Engl J Med 2001;344(26):1979-1984.
- 165. Cooper RD, Wiebe N, Smith N, Keiser P, Naicker S, Tonelli M. Systematic review and meta-analysis: Renal safety of tenofovir disoproxil fumarate in HIV-infected patients. Clin Infect Dis 2010;51(5):496-505.
- 166. Faury G, Maher GM, Li DY, Keating MT, Mecham RP, Boyle WA. Relation between outer and luminal diameter in cannulated arteries. Am J Physiol 1999;277(5 Pt 2):1745-1753.
- 167. Mecham RP. Overview of extracellular matrix. Curr Protoc Cell Biol 1998:10.1.1-.10.1.6.
- 168. Fung Y. Biomechanics–mechanical properties of living tissues, 2nd edn.,1993. Springer-Verlag Inc., New York.
- 169. Greenwald S. Ageing of the conduit arteries. J Pathol 2007;211(2):157-172.
- 170. Willum-Hansen T, Staessen JA, Torp-Pedersen C, Rasmussen S, Thijs L, Ibsen H, et al. Prognostic value of aortic pulse wave velocity as index of arterial stiffness in the general population. Circulation 2006;113(5):664-670.
- 171. Schutte AE, Huisman HW, Schutte R, Van Rooyen JM, Malan L, Malan NT, et al. Arterial stiffness profiles: Investigating various sections of the arterial tree of African and Caucasian people. Clin Exp Hypertens 2011;33(8):511-517.
- 172. Ngatchou W, Lemogoum D, Ndobo P, Yagnigni E, Tiogou E, Nga E, et al. Increased burden and severity of metabolic syndrome and arterial stiffness in treatment-naive HIV+ patients from Cameroon. Vasc Health Risk Manag 2013;9:509-516.

- 173. Van Vonderen MG, Hassink EA, van Agtmael MA, Stehouwer CD, Danner SA, Reiss P, et al. Increase in carotid artery intima-media thickness and arterial stiffness but improvement in several markers of endothelial function after initiation of antiretroviral therapy. J Infect Dis 2009;199(8):1186-1194.
- 174. Gleason RL, Jr., Caulk AW, Seifu D, Parker I, Vidakovic B, Getenet H, et al. Current efavirenz (efv) or ritonavir-boosted lopinavir (lpv/r) use correlates with elevate markers of atherosclerosis in HIV-infected subjects in Addis Ababa, Ethiopia. PLoS One 2015;10(4):117-125.
- 175. Kahan T, Bergfeldt L. Left ventricular hypertrophy in hypertension: Its arrhythmogenic potential. Heart 2005;91(2):250-256.
- 176. Touyz RM, Schiffrin EL. Increased generation of superoxide by angiotensin ii in smooth muscle cells from resistance arteries of hypertensive patients: Role of phospholipase d-dependent nad (p) h oxidase-sensitive pathways. J Hypertens 2001;19(7):1245-1254.
- 177. Bleakley C, Hamilton PK, Pumb R, Harbinson M, McVeigh GE. Endothelial function in hypertension: Victim or culprit? J Clin Hypertens 2015;17(8):651-654.
- 178. Verdecchia P, Porcellati C, Reboldi G, Gattobigio R, Borgioni C, Pearson TA, et al. Left ventricular hypertrophy as an independent predictor of acute cerebrovascular events in essential hypertension. Circulation 2001;104(17):2039-2044.
- 179. Nabbaale J, Kibirige D, Ssekasanvu E, Sebatta ES, Kayima J, Lwabi P, et al. Microalbuminuria and left ventricular hypertrophy among newly diagnosed black African hypertensive patients: A cross sectional study from a tertiary hospital in Uganda. BMC Res Notes 2015;8(1):198.
- 180. Havranek EP, Froshaug DB, Emserman CD, Hanratty R, Krantz MJ, Masoudi FA, et al. Left ventricular hypertrophy and cardiovascular mortality by race and ethnicity. Am J Med 2008;121(10):870-875.
- 181. Mansoor A, Golub ET, Dehovitz J, Anastos K, Kaplan RC, Lazar JM. The association of HIV infection with left ventricular mass/hypertrophy. AIDS Res Hum Retroviruses 2009;25(5):475-481.

- 182. Barbaro G, Di Lorenzo G, Grisorio B, Barbarini G. Incidence of dilated cardiomyopathy and detection of HIV in myocardial cells of HIV-positive patients. N Engl J Med 1998;339(16):1093-1099.
- 183. Meng Q, Lima JA, Lai H, Vlahov D, Celentano DD, Strathdee S, et al. Use of HIV protease inhibitors is associated with left ventricular morphologic changes and diastolic dysfunction. J AIDS 2002;30(3):306-310.
- 184. Glass CK, Witztum JL. Atherosclerosis: The road ahead. Cell 2001;104(4):503-516.
- 185. Libby P. Inflammation in atherosclerosis. Nature 2002;420(6917):868-874.
- 186. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. N Engl J Med 2005;352(16):1685-1695.
- 187. Davies MJ, Woolf N, Rowles P, Pepper J. Morphology of the endothelium over atherosclerotic plaques in human coronary arteries. Br Heart J 1988;60(6):459-464.
- 188. Kragel AH, Reddy SG, Wittes JT, Roberts WC. Morphometric analysis of the composition of atherosclerotic plaques in the four major epicardial coronary arteries in acute myocardial infarction and in sudden coronary death. Circulation 1989;80(6):1747-1756.
- 189. Schwartz SM, Virmani R, Rosenfeld ME. The good smooth muscle cells in atherosclerosis. Curr Atheroscler Rep 2000;2(5):422-429.
- 190. Boutouyrie P. New techniques for assessing arterial stiffness. Diabetes & Metab 2008;34(1):21-26.
- 191. O'Leary DH, Polak JF, Kronmal RA, Manolio TA, Burke GL, Wolfson SK, Jr. Carotid-artery intima and media thickness as a risk factor for myocardial infarction and stroke in older adults. Cardiovascular health study collaborative research group. N Engl J Med 1999;340(1):14-22.
- 192. Lorenz MW, Markus HS, Bots ML, Rosvall M, Sitzer M. Prediction of clinical cardiovascular events with carotid intima-media thickness: A systematic review and meta-analysis. Circulation 2007;115(4):459-467.

- 193. Okeahialam BN, Alonge BA, Pam SD, Puepet FH. Carotid intima media thickness as a measure of cardiovascular disease burden in Nigerian Africans with hypertension and diabetes mellitus. Intern J Vasc Med 2011;2011:327171.
- 194. Schoffelen AF, De Groot E, Tempelman HA, Visseren FL, Hoepelman AI, Barth RE. Carotid intima media thickness in mainly female HIV-infected subjects in rural South Africa: Association with cardiovascular but not HIV-related factors. Clin Infect Dis 2015;61(10):1606-1614.
- 195. Mangili A, Polak JF, Quach LA, Gerrior J, Wanke CA. Markers of atherosclerosis and inflammation and mortality in patients with HIV infection. Atherosclerosis 2011;214(2):468-473.
- 196. Pen A, Yam Y, Chen L, Dennie C, McPherson R, Chow BJ. Discordance between Framingham risk score and atherosclerotic plaque burden. Eur Heart J 2013;34(14):1075-1082.
- 197. Parra S, Coll B, Aragones G, Marsillach J, Beltran R, Rull A, et al.

  Nonconcordance between subclinical atherosclerosis and the calculated

  Framingham risk score in HIV-infected patients: Relationships with serum markers of oxidation and inflammation. HIV Med 2010;11(4):225-231.
- 198. Blom DJ. Cardiovascular risk assessment. S Afr Fam Pract 2011;53(2):121-128.
- 199. Dawber TR, Meadors GF, Moore FE, Jr. Epidemiological approaches to heart disease: The Framingham study. Am J Public Health Nations Health 1951;41(3):279-281.
- 200. Kannel WB, Dawber TR, Kagan A, Revotskie N, Stokes J, 3rd. Factors of risk in the development of coronary heart disease--six year follow-up experience. The Framingham study. Ann Intern Med 1961;55:33-50.
- 201. Anderson KM, Odell PM, Wilson PW, Kannel WB. Cardiovascular disease risk profiles. Am Heart J 1991;121:293-298.
- 202. Wolf PA, D'Agostino RB, Belanger AJ, Kannel WB. Probability of stroke: A risk profile from the Framingham study. Stroke 1991;22(3):312-318.

- 203. Bitton A, Gaziano TA. The Framingham heart study's impact on global risk assessment. Prog Cardiovasc Dis 2010;53(1):68-78.
- 204. Giampaoli S, Palmieri L, Mattiello A, Panico S. Definition of high risk individuals to optimise strategies for primary prevention of cardiovascular diseases. Nutr Metab Cardiovasc Dis 2005;15(1):79-85.
- 205. Jackson R. Guidelines on preventing cardiovascular disease in clinical practice. BMJ 2000;320(7236):659-661.
- 206. Truett J, Cornfield J, Kannel W. A multivariate analysis of the risk of coronary heart disease in Framingham. J Chronic Dis 1967;20(7):511-524.
- 207. Wilson PWF, D'Agostino RB, Levy D, Belanger AM, Silbershatz H, Kannel WB. Prediction of coronary heart disease using risk factor categories. Circulation 1998;97(18):1837-1847.
- 208. D'Agostino RB, Vasan RS, Pencina MJ, Wolf PA, Cobain M, Massaro JM, et al. General cardiovascular risk profile for use in primary care: The Framingham heart study. Circulation 2008;117(6):743-753.
- 209. Dada AS, Ajayi DD, Areo PO, Raimi TH, Emmanuel EE, Odu OO, et al. Metabolic syndrome and Framingham risk score: Observation from screening of low-income semi-urban African women. Medicines 2016;3(2):15.
- 210. Bergersen BM, Sandvik L, Bruun JN, Tonstad S. Elevated Framingham risk score in HIV-positive patients on highly active antiretroviral therapy: Results from a Norwegian study of 721 subjects. Eur J Clin Microbiol Infect Dis 2004;23(8):625-630.
- 211. Ridker PM, Paynter NP, Rifai N, Gaziano JM, Cook NR. C-reactive protein and parental history improve global cardiovascular risk prediction: The Reynolds risk score for men. Circulation 2008;118(22):2243-2251.
- 212. Ridker PM, Buring JE, Rifai N, Cook NR. Development and validation of improved algorithms for the assessment of global cardiovascular risk in women: The Reynolds risk score. JAMA 2007;297(6):611-619.
- 213. Bitton A, Gaziano T. The Framingham heart study's impact on global risk assessment. Prog Cardiovasc Dis 2010;53(1):68-78.

- 214. Cook NR, Paynter NP, Eaton CB, Manson JE, Martin LW, Robinson JG, et al. Comparison of the Framingham and Reynolds risk scores for global cardiovascular risk prediction in The Multiethnic Women's Health Initiative. Circulation 2012;125(14):1748-1756.
- 215. Greenland P, Alpert JS, Beller GA, Benjamin EJ, Budoff MJ, Fayad ZA, et al. 2010 ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults: A report of the American College of Cardiology Foundation/American Heart Association task force on practice guidelines. J Am Coll Cardiol 2010;56(25):50-103.
- 216. Genest J, McPherson R, Frohlich J, Anderson T, Campbell N, Carpentier A, et al. 2009 Canadian cardiovascular society/Canadian guidelines for the diagnosis and treatment of dyslipidemia and prevention of cardiovascular disease in the adult 2009 recommendations. Can J Cardiol 2009;25(10):567-579.
- 217. Executive summary of the third report of The National Cholesterol Education Program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel iii). JAMA 2001;285(19):2486-2497.
- 218. Goff DC, Jr, Lloyd-Jones DM. The pooled cohort risk equations—black risk matters. JAMA Cardiol 2016;1(1):12-14.
- 219. Mashinya F, Alberts M, Van geertruyden J-P, Colebunders R. Assessment of cardiovascular risk factors in people with HIV infection treated with ART in rural South Africa: A cross sectional study. AIDS Res Ther 2015;12(1):1-42.
- 220. Adekunle R, Bagchi S. Review of cardiovascular disease in HIV-infected women. J AIDS Clin Res 2016;7(3):2-12.
- 221. Mancia G, De Backer G, Dominiczak A, Cifkova R, Fagard R, Germano G, et al. 2007 ESH-ESC practice guidelines for the management of arterial hypertension: ESH-ESC task force on the management of arterial hypertension. J Hypertens 2007;25(9):1751-1762.
- 222. Shlomai G, Grassi G, Grossman E, Mancia G. Assessment of target organ damage in the evaluation and follow-up of hypertensive patients: Where do we stand? J Clin Hypertens 2013;15(10):742-747.

- 223. Stanifer JW, Jing B, Tolan S, Helmke N, Mukerjee R, Naicker S, et al. The epidemiology of chronic kidney disease in sub-Saharan Africa: A systematic review and meta-analysis. Lancet Glob Health 2014;2(3):174-181.
- 224. Ando M, Yanagisawa N, Ajisawa A, Tsuchiya K, Nitta K. Urinary albumin excretion within the normal range is an independent risk for near-term development of kidney disease in HIV-infected patients. Nephrol Dial Transplant 2011;26(12):3923-3929.

# Chapter 3: Methodology

#### **Materials and methods**

## Study design and population

This study is embedded in the South African arm of a longitudinal multi-national study, the Prospective Urban and Rural Epidemiological (PURE) study conducted in the North-West province. This study is known for assessing lifestyle changes and causes of development of cardiovascular disease (CVD) [1]. The PURE-South African study of the North-West province started in 2005 (baseline) and consists of a follow-up period of 10 years (2015) which targeted urban and rural areas in low and middle-income countries such as South Africa [1]. The inclusion criteria consisted of volunteers older than 35 years of age who did not receive any chronic medication and did not have any self-reported diseases.

A number of 2010 individuals participated during baseline, of which 322 participants were newly identified as being infected with human immunodeficiency virus (HIV). During follow-up, 100 participants of the 322 HIV-infected participants took part in the 10-year follow-up study of which 29 participants were excluded due to incomplete data sets. The remaining 71 participants, who were infected with HIV for at least 10 years, were matched to 71 HIV-free controls according to age, sex and locality.

## Ethical considerations

Taking part in this study was completely voluntary and the participants could withdraw at any time. The involved procedures were explained to each participant in their own language, followed by the signing of an informed consent form. Each participant was allowed to ask questions when needed. The Ethics Committee of the North-West University, Potchefstroom campus approved the protocol of the PURE-South African study in 2005 (04M10) and 2015 (0016-10-A1) which complies with the Declaration of Helsinki. This study protocol was also approved (NWU-00019-16-A1).

## Experimental protocol

Permission for undertaking the PURE study was obtained from the provincial Department of Health, the local authorities and the tribal chief from the specific rural area. The experimental protocol for the data collection at follow-up was consistent with the protocol for the data collection during baseline. In short, lifestyle data (including self-reported tobacco use, alcohol intake and medical history) of the participants was obtained by trained field workers, in the participant's own language. During individual

post-counselling, each participant was informed about his/her HIV status, blood pressure levels and fasting glucose levels, followed by the referral of the infected participants to the local clinic or hospital for further follow-up and CD4 cell count determination. The fieldworkers signed a confidentiality agreement to protect the privacy of the participants.

## Anthropometric measurements

Calibrated instruments were used to measure the participant's height to the nearest 0.1 cm (Invicta Stadiometer, IP 1465, London, UK) at baseline and follow-up (Leicester height measure, Seca, Birmingham, UK). Weight was measured to the nearest 0.01 kg (Precision Health scale; A & D Company, Tokyo, Japan) at baseline and follow-up. Waist circumference (WC) was measured between the lowest rib and the lateral iliac crest and recorded to the nearest 0.1 cm with a non-stretchable metal tape (Holtain, Crymych, UK) at baseline [2] and with a steel tape (Lufkin, Cooper Tools, Apex NC, USA) at follow-up. The body mass index (BMI) of each participant was calculated (at baseline and follow-up) with the formula: weight (kg)/ height (m²). The above measurements were conducted by trained researchers in a private room.

## Cardiovascular measurements

Systolic and diastolic blood pressure (SBP and DBP) as well as the heart rate of the participants were measured with the validated OMRON HEM-757 (Omron Healthcare, Kyoto, Japan) device during baseline and with a validated OMRON MI 6 (Omron Healthcare, Kyoto, Japan) device during follow-up, according to appropriate and standardised methods. Each participant was fitted with the correct cuff size. The participant had to be calm and rested for more than/at least five minutes before the measurement, should not have smoked, should not have conducted any form of exercise or eaten during the last 30 minutes and should not have climbed stairs during the last 15-30 minutes before completing this measurement. The participant should have been seated in a supine upright position with his/her arm supported at heart level. After the first measurement was conducted, the procedure was repeated five minutes apart and the last measurement was used (a total of two measurements were conducted).

The carotid-femoral pulse wave velocity (cfPWV) was measured non-invasively on the right side of each participant in a supine position. The carotid pulse wave velocity

(cfPWV), which is seen as the golden standard for the measurement of arterial stiffness [3], was measured. The validated SphygmoCor device (ATCor Medical Pty Ltd, Sydney, Australia) used superficial pulses to measure the PWV. The intima-media thickness (IMT) was measured non-invasively with the SonoSite Micromaxx ultrasound system (SonoSite, Inc., Bothel, WA, USA) with a 6-13 MHz linear array transducer on a selected segment of maximum ten millimeter with good image in each subject. The cardiovascular measurements were conducted by researchers in controlled and private conditions. A standard 12-lead electrocardiography (ECG) was recorded during resting conditions (PC 1200, v5.030, Norav Medical, Yokneam, Israel). Electrocardiography left ventricular hypertrophy (ECG-LVH) was determined using the Cornell product [4-6]. The golden standard for the measurement of left ventricular hypertrophy is echocardiography. However, literature suggests that both ECG-LVH and Echocardiography left ventricular hypertrophy (echo-LVH) are equally predictive of incident heart failure and can be used interchangeably in heart failure risk-prediction models [7]. In a middle-income country such as South Africa the measurement of echo-LVH is not always feasible; we therefore used the ECG-LVH in this study.

Plaque scores were not determined in this study. Although ECG LVH is not seen as the golden standard it remains an appropriate measurement.

## Blood, serum and plasma samples

Blood samples from fasting participants were collected by a registered nurse with a sterile winged infusion set and syringes. The blood was drawn from the antebrachial vein and the preparation of both the serum and the plasma was carried out according to standardised methods, snap frozen on dry ice and stored in the laboratory at -80°C. In the case of blood collection in rural areas, serum and plasma were snap frozen and stored at -20°C for a maximum of five days. All the samples were transported to the laboratory and stored in a freezer at -80°C which was/is connected to an alarm system via cell phone. This alarm notified the researcher on duty of any malfunction regarding the freezer, so that the samples were always protected. Mid-stream spot urine samples were collected and frozen at -80°C.

## Biochemical analyses

Total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C), triglycerides (TG), gamma-glutamyltransferase (GGT) and C-reactive protein (CRP) levels were analysed at baseline (Konelab20i™ auto-analyser, Thermo Fisher Scientific Oy, Vantaa, Finland) and at follow-up by particle enhanced turbidimetric assay (Cobas Integra 400 Roche® Clinical System, Roche Diagnostics, Indianapolis, IN) from serum samples. The Friedewald formula [8] was used to calculate low-density lipoprotein cholesterol (LDL-C) levels. Albumin and creatinine levels were determined (Cobas Integra 400 Roche® Clinical System, Roche Diagnostics, Indianapolis, IN) and urinary albumin creatinine ratio (uACR) was calculated. Creatinine clearance (CrCl) was calculated with the Cockcroft-Gault formula: CrCl in ml/min= [(140-age) x (weight in kg) x (0.85 if female)]/(72 x creatinine in mmol/l) [9], at both baseline and follow-up. The glycosylated haemoglobin (HbA1c) levels were determined from blood samples collected in tubes with ethyl-enediamin-e-tetracetic acid. The D-10 haemoglobin testing system from Bio-Rad (#220-0101), which is based on ion-exchange high performance liquid chromatography, was used both at baseline and follow-up.

Glucose levels (blood collected in fluoride tubes) were determined at baseline (Vitros DT6011 Chemistry Analyser; Ortho-Clinical Diagnostics, Rochester, New York, USA) and at follow-up by an enzymatic reference method with hexokinase (Cobas Integra 400 Roche® Clinical System, Roche Diagnostics, Indianapolis, IN).

The HIV status of the participants was determined after informed consent was given, in private, by a trained researcher/counsellor. At both baseline and follow-up, the First Response (PMC Medical, Daman, India) rapid HIV card test was used. If the participant tested positive at baseline, the test was repeated for confirmation by using the Pareekshak card test (BHAT Bio-tech, India) and at follow-up Abon card test (Biopharm Corporation Limited Hanyzhou, China). Feedback regarding the HIV status and post-counselling was given individually in a private room by trained counsellors. Participants who already tested positive at baseline were not tested again. For this study, only participants who were identified as being HIV-infected at baseline were included. The HIV-infected participants were treated with a combination pill containing efavirenz, tenofovir and emtricitabine at follow-up. The CD4 counts were determined (in whole blood) by the National Health Laboratory using flow cytometric analysis (Beckman COULTER® EPICS® XLTM, Fullerton, USA) at baseline and at follow-up

with finger-prick blood and a point-of-care device, PIMA<sup>TM</sup> CD4 (Alere, Jena, Germany).

# Risk analyses

The Framingham [10] and Reynolds [11] risk scores were calculated according to the risk score models in Excel spreadsheets at both baseline and follow-up. Both these risk models consisted of two separate models - one for men and one for women. The risk scores were determined as follows:

Table 2: The Framingham risk table implemented for South Africans [10]

Women

Total

cholesterol (mmol/L)

≤4.09 4.10-5.10 5.11-6.20 6.21-7.20

≥7.21

Age (years)	Points
20-34	-7
35-39	-3
40-44	0
45-49	3
50-54	6
55-59	8
60-64	10
65-69	12
70-74	14
75-79	16

40-49

0

6 8

10

Age 20-39

13

	i
-	
)	

60-69

3

0

50-59

0 2 4

High-density (mmol/L)	lipoprotein	cholesterol	Points
≥1.60			-1
1.30-1.59			0
1.00-1.29			1
<1			2

Systolic blood pressure (mmHg)	Untreated	Treated
<120	0	0
120-129	1	3
130-139	2	4
140-159	3	5
≥160	4	6

Smoking status	20-39	40-49	50-59	60-69	70-79
Non-	0	0	0	0	0
smoker					
Smoker	9	7	4	2	1

Total	10-years cardiovascular disease risk (%)
points	
<9	<1
9	1
10	1
11	1
12	1
13	2
14	2
15	3
16	4
17	5
18	6
19	8
20	11
21	14
22	17
23	22
24	27
>25	≥30

Modified from: Blom (2011) [10]

Men

Age (years)	Points
20-34	-9
35-39	-4
40-44 45-49	0
	3
50-54	6
55-59	8
60-64	10
65-69	11
70-74	12
75-79	13

	Age				
Total cholesterol (mmol/L)	20- 39	40- 49	50- 59	60- 69	70-79
≤4.09	0	0	0	0	0
4.10-5.10	4	3	2	1	0
5.11-6.20	7	5	3	1	0
6.20-7.20	9	6	4	2	1
≥7.21	11	8	5	3	1

High-density (mmol/L)	lipoprotein	cholesterol	Points
≥1.60			-1
1.30-1.59			0
1.00-1.29			1
<1			2

Systolic blood pressure (mmHg)	Untreated	Treated
<120	0	0
120-129	0	1
130-139	1	2
140-159	1	2
≥160	2	3

Smoking status	20-39	40-49	50-59	60-69	70-79
Non-	0	0	0	0	0
smoker					
Smoker	8	5	3	1	1

Total	10-years cardiovascular disease risk (%)
points	
<0	<1
0	1
1	1
2	1
3	1
4	1
5	2
6	2
7	3
8	4
9	5
10	6
11	8
12	10
13	12
14	16
15	20
16	25
≥17	≥30

The Reynolds risk score models [11]

Women [11]

10-year CVD risk (%) =  $[1 - 0.98634^{(exp [B-22.325])}] \times 100\%$  where

B =  $0.0799 \times age + 3.137 \times natural logarithm (SBP) + 0.180 \times natural logarithm (hs-CRP) + 1.382 \times natural logarithm (TC) - 1.172 \times natural logarithm (HDL) + 0.134 \times haemoglobin A<sub>1c</sub> (%) (if diabetic) + 0.818 (if current smoker) + 0.438 (if family history of premature MI).$ 

Men [11]

10-year CVD risk (%) =  $[1 - 0.908^{(exp [B-30.651])}] \times 100\%$  where

B = 4.034 x natural logarithm (age) + 2.562 natural logarithm (SBP) + 0.800 x natural logarithm (TC) – 0.760 x natural logarithm (HDL) + 0.352 (if current smoker) + 0.108 x natural logarithm (hs-CRP) + 0.487 (if family history of premature MI) + 0.098 x haemoglobin A<sub>1c</sub> (%) (if diabetic).

#### Co-morbidity prevalence

The prevalence of co-morbidities was assessed as follows: Obesity, body mass index (BMI) >30 kg/m²; Overweight, BMI >25 kg/m²; Central obesity (WC: men ≥102 cm, women ≥88 cm); Hypertension [systolic blood pressure (SBP) ≥140 mmHg and diastolic blood pressure (DBP) ≥90 mmHg]; TC >5.1 mmol/L; LDL-C >3 mmol/L; HDL-C (men <1mmol/L, women <1.2 mmol/L); Microalbuminuria (ACR 3-30 mg/mmol); eGFR <60 ml/min; ECG derived LVH (Cornell product >244 mV/ms) [12] Diabetes (Glucose >7 mmol/L); cfPWV >10 m/s; sub-clinical atherosclerosis (IMT >0.9 mm) [13] TG >1.7 mmol/L; Atherogenic dyslipidaemia (TG ≥2.31 mmol/L and HDL-C ≤0.88 mmol/L) [14] TG: HDL-C ≥1.49 mmol/L [15] CRP >3 mg/L [16] GGT (men ≥80 U/L, women ≥50 U/L) [17] AIDS (CD4 <200 cells/mm³) [18] CrCl <50 ml/min [19].

#### Statistical analysis

The statistical analysis was performed by using Statistica® 13 (StatSoft, Inc., Tulsa, OK, USA). Basic descriptive statistics were used to determine normal distribution of the data, logarithmic transformation was applied and presented as geometric mean with 5<sup>th</sup> and 95<sup>th</sup> percentiles if skewed. Groups were compared using independent t-tests and Chisquare tests as appropriate. Associations of measures of end-organ damage (PWV, IMT,

CrCl and Cornell product) with the risk scores were determined by making use of Pearson and partial correlations. Odds ratios with cut-off values and 95% confidence intervals (CI), were calculated. Since very few participants met the cut-off values for measures of end-organ damage and risk, the median values were used as cut-off values for the calculation of odds ratios. Median values were as follows: PWV median=8.15 m/s, IMT median=0.37 mm, CrCl median=103 ml/min, Cornell product median=64.6 mV/ms, CHD median=1% and CVD median=0.85%.

#### References

- 1. Teo K, Chow CK, Vaz M, Rangarajan S, Yusuf S. The Prospective Urban Rural Epidemiology (PURE) study: Examining the impact of societal influences on chronic noncommunicable diseases in low-, middle-, and high-income countries. Am Heart J 2009;158(1):1-7.
- 2. Marfell-Jones MJ, Stewart A, de Ridder J. International standards for anthropometric assessment. Lower Hutt, New Zealand: International Society for the Advancement of Kinanthropometry, 2011: 2011.
- 3. Laurent S, Cockcroft J, Van Bortel L, Boutouyrie P, Giannattasio C, Hayoz D, Pannier B, Vlachopoulos C, Wilkinson I, Struijker-Boudier H. on behalf of the European Network for non-invasive investigation of large arteries. Abridged version of the expert consensus document on arterial stiffness. Artery Res 2007;1:2–12.
- 4. Dahlöf B, Devereux RB, Julius S, Kjeldsen SE, Beevers G, de Faire U, et al. Characteristics of 9194 patients with left ventricular hypertrophy The LIFE study. Hypertension 1998;32(6):989-997.
- 5. Julius S, Alderman MH, Beevers G, Dahlöf B, Devereux RB, Douglas JG, et al. Cardiovascular risk reduction in hypertensive black patients with left ventricular hypertrophy: The LIFE study. J Am Coll Cardiol 2004;43(6):1047-1055.
- 6. Okin PM, Roman MJ, Devereux RB, Kligfield P. Electrocardiographic identification of increased left ventricular mass by simple voltage-duration products. J Am Coll Cardiol 1995;25(2):417-423.
- 7. Almahmoud MF, O'Neal WT, Qureshi W, Soliman EZ. Electrocardiographic Versus Echocardiographic Left Ventricular Hypertrophy in Prediction of Congestive Heart Failure in the Elderly. Clin Cardiol 2015;38(6):365-370.
- 8. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem 1972;18(6):499-502.
- 9. Cockcroft DW, Gault MH. Prediction of creatinine clearance from serum creatinine. Nephron 1976;16(1):31-41.

- 10. Blom DJ. Cardiovascular risk assessment. S Afr Fam Prac 2011;53(2):121-128.
- 11. Dolman RC. The role of diet in cardiovascular disease in black south africans: Both sides of the story [doctorate thesis]. North-West University. 2013, 50-51.
- 12. Seedat Y, Rayner B, Veriava Y. South African hypertension practice guideline 2014. Cardiovasc J Afr 2014;25(6):288-294.
- 13. Mancia G, Fagard R, Narkiewicz K, Redon J, Zanchetti A, Böhm M, et al. 2013 ESH/ESC guidelines for the management of arterial hypertension: The task force for the management of arterial hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC). Blood Press 2013;22(4):193-278.
- 14. Aguiar C, Alegria E, Bonadonna RC, Catapano AL, Cosentino F, Elisaf M, et al. A review of the evidence on reducing macrovascular risk in patients with atherogenic dyslipidaemia: A report from an expert consensus meeting on the role of fenofibrate-statin combination therapy. Atheroscler Suppl 2015;19:1-12.
- 15. Van Rooyen J, Fourie C, Steyn H, Koekemoer G, Huisman H, Schutte R, et al. Cardiometabolic markers to identify cardiovascular disease risk in HIV-infected black South Africans. S Afr Med J 2014;104(3):195-199.
- 16. Jiménez MC, Rexrode KM, Glynn RJ, Ridker PM, Gaziano JM, Sesso HD. Association between high-sensitivity c-reactive protein and total stroke by hypertensive status among men. J Am Heart Assoc 2015;4(9):20-73.
- 17. T Pisa P, H Vorster H, Kruger A, Margetts B, T Loots D. Association of alcohol consumption with specific biomarkers: A cross-sectional study in South Africa. J Health Popul Nutr 2015;33(1):146-156.
- 18. Ferrer E, Curto J, Esteve A, Miro J, Tural C, Riera S, et al. Progression to AIDS or death in HIV-infected patients initiating cART with CD4< 200 cells/µl: The role of CD4 and viral load changes during follow-up. J Int AIDS Soc 2012;15(6):18148.
- 19. Meintjes G, Maartens G. Guidelines for antiretroviral therapy in adults. S Afr J HIV Med 2012;13(3):114-133.

## Chapter 4: Manuscript for publication

#### INSTRUCTIONS TO AUTHORS: Journal: Heart, Lung and Circulation

• Complete manuscript. (1) Title page, (2) abstract and keywords if required, (3) text, (4) acknowledgments, (5) disclosures if required, (6) references, (7) tables (each complete with title) (8) figures and (9) figure legends.

#### Essential title page information

**Title.** Concise and informative. Titles are often used in information-retrieval systems.

**Author names and affiliations.** Clearly indicate given name(s) and family name(s) of each author. Present the authors' affiliation addresses below the names. Indicate all affiliations with a lower-case superscript letter immediately after the author's name and in front of the appropriate address. Provide the city and state and country name of each affiliation.

**Corresponding author.** Ensure that the full contact address and e-mail address is given.

#### Article structure

*Introduction*. State objectives of the work and provide an adequate background.

**Material and methods**. Provide sufficient detail to allow the work to be reproduced. Methods already published should be indicated by a reference: only relevant modifications should be described.

**Results**. Clear and concise.

**Discussion**. Explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is often appropriate. Avoid extensive citations and discussion of published literature.

**Conclusions**. The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion or Results and Discussion section.

- Abstract. A concise and factual abstract is required.
- Acknowledgements. Collate acknowledgements in a separate section at end
  of the article before the references and do not, therefore, include them on the
  title page, as a footnote to the title or otherwise.
- Formatting of funding sources. List funding sources with Grant numbers.
   No detail needed.

• **Nomenclature and units**. Use the international system of units (SI). If other quantities are mentioned, give their equivalent in SI.

#### • References

**Reference style**. Vancouver referencing style. Consecutive numbers in square brackets to be used to indicate references in the text, as part of the text. Endnotes should be placed at the end of the manuscript following the Acknowledgements.



# Cardiovascular disease risk assessment in HIV-infected black South Africans: A longitudinal study

Marlene Duvenhage<sup>a</sup>, Carla Maria Theresia Fourie<sup>a, b\*</sup> and Johannes

Marthinus van Rooyen<sup>a, b</sup>

<sup>a</sup>Hypertension in Africa Research Team (HART); North-West University (Potchefstroom Campus); Potchefstroom; South Africa

<sup>b</sup>South African Medical Research Council Unit for Hypertension and Cardiovascular Disease

#### \*Corresponding author:

CMT Fourie, RN, PhD

Hypertension in Africa Research Team (HART)

Private Bag X6001

North-West University (Potchefstroom Campus)

Potchefstroom

2522

South Africa

Tel: +2718 299 2080 Fax: +2718 285 2432

E-mail: carla.fourie@nwu.ac.za

#### **Abstract**

#### Introduction:

Cardiovascular risk is increased in human immunodeficiency virus (HIV)-infected individuals and may be due to risk factors and/or antiretroviral treatment (ART). Africans may be burdened with both CVD and HIV. Those infected with HIV also show higher prevalence of measures of end-organ damage. We aimed to assess the 10-year coronary heart disease (CHD) and cardiovascular risk in a cohort infected with HIV. Furthermore, we determined whether high risk may be associated with measures of end-organ damage.

#### Methods:

In 71 HIV-infected participants and 71 controls (matched according to age, sex and locality) cardiovascular risk markers, co-morbidities, CHD and cardiovascular risk and associations of risk with measures of end-organ damage were determined.

#### Results:

The CHD and cardiovascular risk did not differ between the HIV-infected and their matched controls. Prevalence of obesity, diabetes and microalbuminuria were higher among the controls at follow-up. A borderline negative correlation (p=0.053) was seen between CrCl and CHD at follow-up among those infected.

#### Conclusion:

The HIV-infected participants did not have a higher CHD or CVD risk compared to their matched HIV-free controls, even though 80% of the HIV-infected participants were treated. No associations between risk scores and measures of end-organ damage were found.

**Keywords:** Cardiovascular disease, Human immunodeficiency virus, Risk assessment, Africans, End-organ damage

#### Introduction

Cardiovascular disease (CVD) is considered a leading cause of morbidity and mortality worldwide and especially among Africans [1]. Furthermore, African-Americans are known to have a higher risk of developing coronary heart disease (CHD) compared to other ethnicities [2]. Besides being burdened by non-communicable diseases (NCD), Southern and Eastern Africa are known as the regions that are home to the highest number of people living with HIV/AIDS [3]. The introduction of antiretroviral therapy (ART) has improved the prognosis for many HIV-infected individuals [4]. However, ART is associated with a disturbance in lipid metabolism which may lead to dyslipidaemia [5] and hypertension [6], significant CVD risk factors [7]. Human immunodeficiency virus-infected individuals are more likely to show elevated levels of inflammation [8] and the inflammatory marker C-reactive protein (CRP) is known to be a contributing risk marker to the progression of CVD in the general population [9] and in the HIV-infected population [10].

In a study conducted in Cameroon, treatment naïve HIV-infected individuals showed early onset of aortic stiffness [11] which may be directly associated with atherosclerosis and CVD [12]. Chronic kidney disease (CKD) is seen as a risk factor for CVD [13]. CKD may be promoted through various mechanisms; heart failure promoting decline in kidney function and atherosclerosis leading to renovascular disease [13]. Chronic kidney disease has long been recognised in the HIV-infected population especially those of African descent [14]. Microalbuminuria, indicating CKD, may be an early marker of CVD risk in the HIV-infected population [15]. Left ventricular hypertrophy (LVH), a predictor of CHD [16] and CVD mortality [17], are also more prevalent in the HIV-infected population [18].

A traditional risk factor contributing to the development of both CHD [19] and CVD [20] is smoking. This behaviour is aggravated in the HIV-infected population [21].

In light of the above the need for risk assessment in the African HIV-infected population burdened by both CHD and CVD is important. Cardiovascular risk assessment was described by Blom (2011) as "predicting the future" by implementing risk scores [22]. The Framingham Risk Score has been known as the "heart" of risk assessment for decades [9] and estimates the 10-year risk of coronary endpoints (myocardial infarction and coronary death) [22]. The Reynolds Risk Score, assessing 10-year CVD risk, incorporates CRP as a risk marker [23]. Therefore, we firstly aimed to assess the

10-year CHD (Framingham) and CVD (Reynolds) risk in an African HIV-infected cohort, and secondly to determine the associations of CHD and CVD risk with measures of end-organ damage.

#### **Materials and methods**

#### Study design and population

This study is embedded in the South African arm of the PURE Prospective Urban and Rural Epidemiological (PURE) study. Data were collected in the North-West Province during 2005 (baseline) when 2010 individuals participated, of which 322 were newly identified as being HIV-infected. During 2015 (follow-up) 100 of the 322 HIV-infected participants took part in the 10-year follow-up study of whom 29 participants were excluded due to incomplete data. The remaining 71 HIV-infected participants were matched to 71 HIV-free controls according to age, sex and locality.

#### Experimental protocol

Procedures regarding the experimental protocol were in accordance with those of Fourie et al. (2015) [24].

#### Ethical considerations

The Ethics Committee of the North-West University approved the protocol of the PURE-SA study in 2005 and 2015 which complies with the Declaration of Helsinki.

#### Anthropometric measurements

We used standardised procedures to measure height, weight and waist circumference (WC) during baseline and follow-up.

#### Cardiovascular measurements

Blood pressure measurements during baseline were in accordance with those of Fourie et al. (2015) [24] as it was calculated with the validated OMRON HEM-757 device and at follow-up the validated OMRON MI 6 device was used. The carotid-femoral pulse wave velocity (cfPWV) was measured with the validated SphygmoCor device (ATCor Medical Pty Ltd, Sydney, Australia) during follow-up. The intima-media thickness (IMT) measurements were in accordance with those of Schutte et al. (2012) where it was measured with the SonoSite Micromaxx ultrasound system (SonoSite, Inc., Bothel, WA, USA) with a 6-13 MHz linear array transducer on a selected segment

of maximum ten millimeter with good image in each subject during follow-up [25]. Electrocardiogram (ECG) measurements were performed [26] and ECG ventricular mass was determined with the Cornell product [27].

#### Biochemical analyses

Total cholesterol (TC), HDL-C (high-density lipoprotein cholesterol), triglycerides (TG), gamma-glutamyltransferase (GGT) and hs-CRP levels were analysed at baseline (Konelab20i™ auto-analyser, Thermo Fisher Scientific Oy, Vantaa, Finland) and at follow-up by particle enhanced turbidimetric assay (Cobas Integra 400 Roche® Clinical System, Roche Diagnostics, Indianapolis, IN) from serum samples. Low-density lipoprotein cholesterol (LDL-C) levels were determined with the Friedewald formula [28]. Urinary albumin and creatinine (uACR) levels were determined. Creatinine clearance (CrCl) was calculated with the Cockcroft-Gault formula [29]. The glycosylated haemoglobin A1c (HbA1c) levels were determined. The D-10 haemoglobin testing system was utilised at both baseline and follow-up. Glucose levels were determined at baseline and at follow-up.

At both baseline and follow-up, the First Response (PMC Medical, Daman, India) rapid HIV card test was used to determine the HIV-statuses. If the participant tested positive at baseline, the test was repeated for confirmation by using the Pareekshak card test (BHAT Bio-tech, India) and at follow-up Abon card test (Biopharm Corporation Limited Hanyzhou, China). The CD4 counts were determined (in whole blood) by the National Health Laboratory using flow cytometric analysis (Beckman COULTER® EPICS® XLTM, Fullerton, USA) at baseline and at follow-up with finger-prick blood and a point-of-care device, PIMA<sup>TM</sup> CD4 (Alere, Jena, Germany). Only participants who were identified as being HIV-infected at baseline were included in our study.

#### Risk analysis

The Framingham Risk Score [22] and the Reynolds Risk Score [23] were calculated according to the risk score models in Excel spreadsheets. Both these risk models consist of separate models for men and women.

#### Statistical analyses

The statistical analyses was performed by using Statistica<sup>®</sup> 13 (StatSoft, Inc., Tulsa, OK, USA). Basic descriptive statistics were used to determine normal distribution of the data, logarithmic transformation were applied and presented as geometric mean with 5<sup>th</sup> and

95<sup>th</sup> percentiles if skewed. Groups were compared using independent t-tests and Chisquare tests.

Associations of measures of end-organ damage (PWV, IMT, CrCl and Cornell product) with the risk scores were determined by making use of Pearson and partial correlations. Odds ratios with median values and 95% confidence intervals (CI) were calculated. Since very few participants met the cut-off values for measures of end-organ damage and risk, the median was used as cut-off value for the calculation of odds ratios. Median values were as follows: PWV median=8.15 m/s, IMT median=0.37 mm, CrCl median=103 ml/min, Cornell product median=64.6 mV/ms, CHD median=1% and CVD median=0.85%.

#### Results

From Table 3 the characteristics of the HIV-free control group and the HIV-infected group at baseline and 10-year follow-up are evident. The HIV-infected group had significantly lower HDL-C (p<0.01) and CrCl (p=0.02) levels at baseline, compared to the HIV-free group. At follow-up the HIV-infected group showed significantly lower BMI (p<0.01), WC (p<0.01) and HbA1c (p=0.01) compared to the HIV-free control group.

At follow-up the CD4 counts of those infected with HIV were higher compared to baseline (p=0.03).

The prevalence of co-morbidities among the HIV-free and HIV-infected groups at the 10-year follow-up study is reported in Table 4. This table shows that more HIV-free participants were overweight, had diabetes, had lower HDL-C (men) and had a higher prevalence of microalbuminuria compared to the HIV-infected group.

Table 3: Characteristics of HIV-free and HIV-infected participants at baseline and 10-year follow-up

		Baseline 2005			Follow-up 2015	
	HIV-free	HIV-infected	p-value	HIV-free	HIV-infected	р
	(n=71)	(n=71)		(n=71)	(n=71)	
Demographic variables						
Age (years)	$43 \pm 0.44$	$43 \pm 5.85$	0.95	$53 \pm 5.32$	$53 \pm 5.43$	0.80
Sex male, <i>n</i> (%)	15 (21)	15 (21)	-	15 (21)	15 (21)	-
Locality, n (%)*	34 (48)	34 (48)	-	34 (48)	34 (48)	-
Anthropometric variables						
BMI (kg/m²)	24.2 (17.8-36.1)	22.9 (16.7-35.5)	0.16	25.9 (18.0-36.1)	23.1 (16.1-37.5)	< 0.01
WC (cm)	79.8 ± 10.8	75.4 ± 10.1	0.54	89.4 ± 13.7	83.0 ± 14.5	< 0.01
Cardiovascular variables						
SBP (mmHg)	124 ± 20.2	123 ± 20.8	0.80	129 ± 22.5	127 ± 24.3	0.73
DBP (mmHg)	84.1 ± 14.8	83.9 ± 13.6	0.49	85.5 ± 13.1	83.2 ± 13.4	0.30
Biochemical variables						
TC (mmol/L)	5.40 ± 1.44	4.36 ± 1.23	0.20	4.66 ± 1.22	$4.48 \pm 1.03$	0.35
HDL-C (mmol/L)	1.59 (0.74-2.98)	1.19 (0.55-2.32)	<0.01	1.29 (0.74-2.27)	1.30 (0.74-2.48)	0.90
LDL-C (mmol/L)	3.09 ± 1.17	2.51 ± 1.00	0.20	2.83 ± 1.07	2.58 ± 0.86	0.13
TG (mmol/L)	1.12 (0.59-2.43)	1.05 (0.50-2.34)	0.45	1.14 (0.50-3.17)	1.10 (0.50-3.11)	0.73
HbA1c (mmol/L)	5.57 (5.00-6.30)	5.49 (4.90-6.20)	0.46	5.82 (5.00-8.80)	5.47 (4.80-6.20)	0.01
Glucose (mmol/L)	5.00 ± 1.80	4.70 ± 0.80	0.23	5.50 ± 1.80	5.12 ± 0.83	0.11
CRP (mg/L)	1.85 (0.10-15.3)	2.29 (0.26-47.7)	0.42	2.85 (0.44-33.9)	4.22 (0.54-44.7)	0.06
GGT (U/L)	53.9 (19.0-339)	44.3 (16.0-228)	0.16	39.1 (11.2-225)	52.7 (15.4-342)	0.07
Lifestyle variables	,	` ,		,	,	
Tobacco use, n (%)	25 (35)	28 (39)	0.60	25 (35)	28 (39)	0.60
Alcohol use, n (%)	22 (31)	25 (35)	0.59	22 (31)	25 (35)	0.59
HT med, n (%)	8 (11)	8 (11)	-	16 (23)	12 (17)	0.40
CHOL med, $n$ (%)	- '	- ` ′	-	2 (3)	1 (1)	0.56
CD4 (cells/mm <sup>3</sup> )	-	362 ± 181	-	-	500 ± 258	-
ART, n (%)	-	-	-	-	57 (80)	-
ART duration, n (%)					, ,	
>5 years	-	-	-	-	31 (44)	-
<5 years	-	-	-	-	32 (45)	-
Fam hist. CHD, n (%)	23 (32)	32 (45)	0.12	-	-	-
Risk scores	,	,				
CHD, (%)	$2.63 \pm 3.30$	1.80 ± 2.08	0.08	$3.65 \pm 4.39$	$3.22 \pm 3.37$	0.51
CVD, (%)	1.37 ± 1.78	1.88 ± 4.01	0.33	$3.98 \pm 3.82$	$4.27 \pm 6.64$	0.75
Measures of end-organ dan	nage					
cfPWV (m/s)*	-	-	=	8.38 ± 1.77	8.27 ± 1.85	0.71
IMTf (mm)	-	_	-	$0.39 \pm 0.09$	$0.38 \pm 0.12$	0.53
uACR (mg/mmol)	1.13 ± 1.96	$5.23 \pm 20.7$	0.10	11.43 ± 48.7	$2.52 \pm 3.30$	0.14
CrCl (ml/min)	99.5 (61.7-193)	87.0 (51.1-156)	0.02	107 (71.5-194)	98.0 (55.8-167)	0.09
eGFR (ml/min)	122 ± 34.6	114 ± 37.0	0.20	132 (89.0-215)	136 (94.6-200)	0.62
Cornell product (mV/ms		-	-	56.8 (12.7-179)	65.9 (19.4-247)	0.29

n indicates the number of participants; locality \*(urban); BMI, body mass index; WC, waist circumference; SBP, systolic blood pressure; DBP, diastolic blood pressure; TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; TG, triglycerides; HbA1c, glycosylated haemoglobin; CRP, C-reactive protein; GGT, gamma-glutamyltransferase; HT med, hypertension medication; CHOL med, cholesterol medication; ART, antiretroviral treatment; Fam hist. CHD, family history coronary heart disease; CHD, coronary heart disease (Framingham Risk Score); CVD, cardiovascular disease (Reynolds Risk Score); cfPWV, carotid-femoral pulse wave velocity \*(adjusted for mean arterial pressure); IMTf, intima-media thickness-far wall mean; uACR, urinary albumin creatinine ratio; CrCl, creatinine clearance; eGFR, estimated glomerular filtration rate. Data are expressed as mean with standard deviation or geometric mean with 5<sup>th</sup> and 95<sup>th</sup> percentiles. P-values were obtained with independent t-tests, categorical variables with Chi-square tests and PWV p-value with ANCOVA. P-values ≤0.05 are regarded as significant.

Table 4: Prevalence of co-morbidities among HIV-free controls and HIV-infected participants at 10-year follow-up

		Follow-up 2015		
	Total	HIV-free	HIV-infected	р
	Group			
	(n=142)	(n=71)	(n=71)	
Cardiovascular comorbidities				
Obesity, BMI >30 kg/m <sup>2</sup> , n / total (%) [30]	34/141 (24)	20/70 (29)	14 (20)	0.22
Overweight, BMI >25 kg/m <sup>2</sup> , $n$ / total (%) [30]	63/141 (45)	38/70 (54)	25 (35)	0.02
Central obesity [30]	` '	, ,	. ,	
Men ≥102 cm, n / total (%)	2/30 (7)	1/15 (7)	1/15 (7)	-
Women ≥88 cm, <i>n</i> / total (%)	52/112 (46)	31/56 (55)	21/56 (38)	0.06
Hypertension, $n(\%)^{[30]}$	28 (20) ` ′	16 (23)	12 (17) ´	0.40
Diabetes, <i>n</i> (%) [31]	4 (3)	4 (6)	0 (0)	0.04
TC >5.1 mmol/L, n (%) [30]	44 (31)	24 (34)	20 (28)	0.47
LDL-C >3 mmol/L, $n$ (%) [30]	49 (35)	28 (40)	21 (30)	0.22
HDL-C [30]	(00)	_ ( ' ' ' ' '	_: (==)	•
Men <1 mmol/L, n / total (%)	8/30 (27)	7/15 (47)	1/15 (7)	0.013
Women <1.2 mmol/L, n / total (%)	46/112 (41)	21/56 (38)	25/56 (45)	0.44
TG >1.7 mmol/L, n / total (%) [32]	26/141 (18)	13/70 (19)	13 (18)	0.97
TG: HDL-C $\geq$ 1.49 mmol/L, $n$ / total (%) [33]	24/141 (17)	12/70 (17)	12/70 (17)	0.97
Atherogenic dyslipidaemia, $n / \text{total } (\%)$ [32]	2 // ( /	.2,70 ()	12/10 (11)	0.01
(TG ≥2.31 mmol/L and HDL-C ≤0.88 mmol/L)	5/141 (4)	4/70 (6)	1 (1)	0.17
CRP >3 mg/L, $n$ (%) [34]	79 (56)	35 (49)	44 (62)	0.13
GGT [35]	10 (00)	00 (10)	11 (02)	0.10
Men ≥80 U/L, <i>n</i> / total (%)	11/30 (37)	5/15 (33)	6/15 (40)	0.70
Women ≥50 U/L, n / total (%)	40/112 (36)	18/56 (32)	22/56 (39)	0.43
AIDS (CD4 <200 cells/mm <sup>3</sup> ), n / total (%) [36]	8/68 (12)	0 (0)	8/68 (12)	-
Risk assessment	0/00 (12)	0 (0)	0/00 (12)	
CHD (Framingham Risk Score)				
Low (<10%), n (%)	128 (90)	63 (89)	65 (92)	0.57
Medium (10-20%), <i>n</i> (%)	13 (9)	7 (10)	6 (8)	0.57
High (>20%), n (%)	1 (1)	1 (1)	0 (0)	-
CVD (Reynolds Risk Score)	' (')	' (')	0 (0)	
Low (<10%), n (%)	132 (93)	66 (93)	66 (93)	0.29
Medium (10-20%), <i>n</i> (%)	8 (6)	5 (7)	3 (4)	0.29
High (>20%), n (%)	2 (1)	0 (0)	2 (3)	0.29
Measures of end-organ damage	2 (1)	0 (0)	2 (3)	0.23
Stiffness (cfPWV >10 m/s), n / total (%) [31]	15/126 (12)	8/65 (12)	7/61 (11)	0.89
SC atherosclerosis (IMTf $>0.9$ mm), $n$ (%) [31]	0 (0)	0 (0)	0 (0)	-
Microalbuminuria, $n / \text{total } (\%)^{[30]}$	4/130 (3)	4/64 (6)	0 (0)	0.04
CrCl <50 ml/min, $n$ / total (%) [37]	4/130 (3) 1/141 (1)		1(1)	0.04
		0 (0)		
eGFR <60 ml/min, n (%) [30]	0 (0)	0 (0)	0 (0)	- 0.20
ECG LVH (Cornell product >244 mV/ms), $n$ (%) [30]		1/60 (2)	3/59 (5)	0.30

n indicates the number of participants; BMI, body mass index; TC, total cholesterol; LDL-C, low-density lipoprotein cholesterol; HDL-C, high-density lipoprotein cholesterol; TG, triglycerides; CRP, C-reactive protein; GGT, gamma-glutamyltransferase; CHD, coronary heart disease; CVD, cardiovascular disease; cfPWV, carotid-femoral pulse wave velocity; SC atherosclerosis (IMTf), sub-clinical atherosclerosis intima-media thickness-far wall mean; CrCl, creatinine clearance; eGFR, estimated glomerular filtration rate; ECG LVH, electrocardiographic derived left ventricular hypertrophy. P-values were obtained with Chisquare tests and p-values ≤0.05 are regarded as significant.

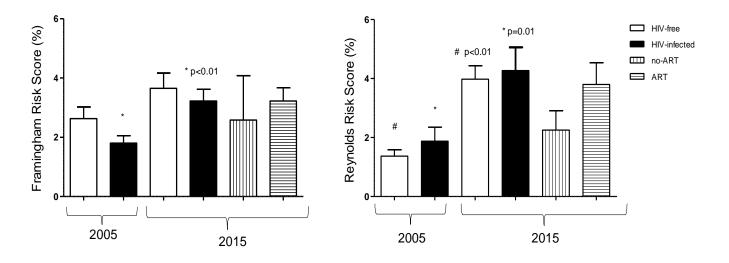


Figure 1: Baseline and follow-up Reynolds (CVD) and Framingham (CHD) risk scores in HIV-free and HIV-infected groups with the influence of treatment

Both the Framingham Risk Score (CHD risk) and Reynolds Risk Score (CVD risk) were determined for the HIV-free controls and HIV-infected group at baseline and follow-up. No differences were seen between the HIV-free and HIV-infected group with either the risk scores. Compared to baseline, the Framingham Risk Score was higher in the HIV-infected group at follow-up, however both the HIV-free controls and HIV-infected group had a higher Reynolds Risk Score at follow-up than at baseline. No differences were seen between the no-ART and ART groups (Figure 1).

Figure 2 reports the Pearson correlations between IMTf and CrCl with CHD risk and Cornell product with CVD risk in the HIV-free and HIV-infected group at 10-year follow-up. A borderline negative correlation (p=0.053) was seen between CrCl and CHD in the HIV-infected group at follow-up.

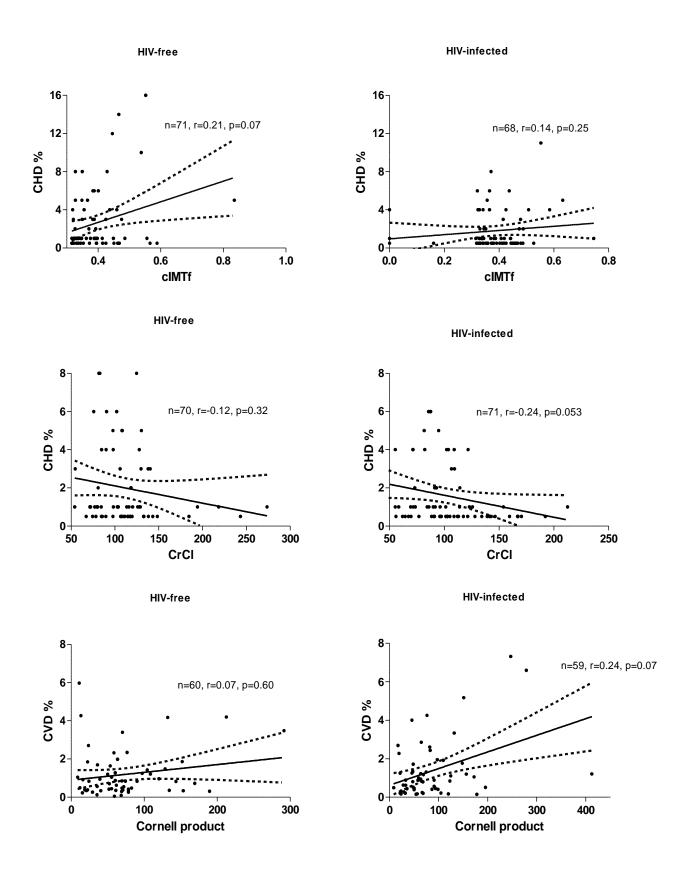


Figure 2: Scatterplots indicating the Pearson correlations between (intima-media thickness-far wall mean) IMTf and (creatinine clearance) CrCl with the Framingham Risk Score (CHD) and Cornell product with the Reynolds Risk Score (CVD) in the HIV-free and HIV-infected group at 10-year follow-up

Table 5: Partial correlations of 10-year Framingham (CHD) Risk Score and Reynolds (CVD) Risk Score with measures of end-organ damage at 10-year follow-up in the HIV-free and HIV-infected group

Coronary heart disease (Framingham Risk Score)				
	cfPWV (m/s)	IMTf (mm)	CrCl (ml/min)	CP (mV/ms)
Group				
HIV-free	r=-0.13, p=0.31	r=0.07; p=0.58	r=-0.07; p=0.58	r=-0.17; p=0.22
HIV-infected	r=-0.22; p=0.60	r=-0.08; p=0.82	r=0.16; p=0.67	r=-0.48; p=0.34
	Card	iovascular disease (Reyno	olds Risk Score)	·
	cfPWV (m/s)	IMTf (mm)	CrCl (ml/min)	CP (mV/ms)
Group	-			
HIV-free	r=0.06; p=0.65	r=0.03; p=0.82	r=-0.14; p=0.28	r=-0.30; p=0.03
HIV-infected	r=-0.82; p=0.01	r=-0.21; p=0.56	r=-0.25; p=0.49	r=-0.31; p=0.55

CHD, coronary heart disease; CVD, cardiovascular disease; cfPWV, carotid-femoral pulse wave velocity; IMTf, intimamedia thickness (far wall mean); CrCl, creatinine clearance; CP, Cornell product. P-values ≤0.05 are regarded as significant

Table 5 reports the partial correlations of the risk scores (CHD -and CVD) respectively with measures of end-organ damage. In the HIV-infected group we adjusted for the risk variables: age, sex, WC, SBP, CRP, CD4 and ART use. Cardiovascular risk correlated negatively with cfPWV in the HIV-infected group. In sensitivity analyses we replaced ART with tobacco use or alcohol use, however the results did not differ. In the HIV-free group we adjusted for the risk variables: age, sex, WC, SBP, CRP, tobacco use and alcohol use. Cardiovascular risk correlated negatively with Cornell product in the HIV-free group.

Table 6: Odds Ratios of 10-year risk scores and measures of end-organ damage in the HIV-free and HIV-infected participants

Coronary heart disease (Framingham Risk Score)				
	cfPWV (95% CI)	IMTf (95% CI)	CrCl (95% Cl)	CP (95% CI)
Group				
HIV-free	0.55 (0.19-1.57)	1.08 (0.40-2.93)	0.46 (0.16-1.31)	0.90 (0.31-2.65)
HIV-infected	1.23 (0.45-3.36)	0.60 (0.23-1.58)	0.39 (0.15-1.03)	2.17 (0.76-6.20)
	Cardio	vascular disease (Reyno	lds Risk Score)	
	cfPWV (95% CI)	IMTf (95% CI)	CrCl (95% Cl)	CP (95% CI)
Group				
HIV-free	0.74 (0.28-1.96)	1.68 (0.65-4.30)	0.57 (0.22-1.47)	0.87 (0.31-2.39)
HIV-infected	1.23 (0.45-3.36)	0.61 (0.23-1.61)	0.55 (0.21-1.44)	1.42 (0.51-3.96)

OR, odds ratios; cfPWV, carotid-femoral pulse wave velocity; IMTf, intima-media thickness-far wall mean; CrCl, creatinine clearance; CP, Cornell product. Significance is indicated by  $^{\star}$ 

The odds ratios of measures of end-organ damage and 10-year CHD -and CVD risk in the HIV-free and HIV-infected group are shown in Table 6. cfPWV was adjusted for mean arterial pressure (MAP). No significant odds were found for having a higher than median CHD or CVD risk with higher than median PWV, IMT and Cornell product or lower than median CrCl.

#### Discussion

To the best of our knowledge, we are the first to investigate CHD (Framingham Risk Score) and CVD (Reynolds Risk Score) risk and the association thereof with measures of end-organ damage in HIV-infected South Africans. The main finding of this study was that the HIV-infected participants did not have a higher CHD or CVD risk when compared to the age, sex and locality matched HIV-free control group; neither at baseline nor at 10-year follow-up. Furthermore, antiretroviral treatment did not seem to affect the CHD nor the CVD risk among those infected with HIV.

Although the Framingham risk model was mainly developed for risk prediction among white populations, it previously performed well to predict CHD risk in African American populations [38]. The Reynolds risk model also showed improved discrimination overall and especially in black and white women [39]. Mashinya et al. (2015) concluded that there is no need to develop a race/ethnicity specific risk model for HIV infected Africans [40].

In a cross-sectional study by Mashinya et al. (2015) the HIV-infected Africans in their study were classified as having a low 10-year Framingham Risk Score [40] which is in accordance with our findings.

ART may lead to lower immune activation [41] and this is seen in this study with the higher CD4 counts in the HIV-infected group (treated and not treated) at follow-up (p=0.03). Besides lowering the immune activation, ART may decrease the inflammatory response in those living with HIV [41]. However, in this study the CRP levels did not differ at baseline (all participants were ART naïve), while the CRP levels tended to be higher at follow-up (p=0.06) with 80% of the participants being on treatment.

Several cross-sectional studies reported that higher CRP levels were associated with an increase in IMT [42, 43] which correlated with arterial stiffness [11]. This correlation may be attributed to viral replication leading to an increase in inflammation and endothelial activation associated with atherosclerotic lesions and CVD [43]. We found no indication of atherosclerosis or arterial stiffness (PWV <10 m/s) in the HIV-infected group. Awotedu et al. (2015) found an increase in stiffness in their HIV-infected treatment-naïve group. They speculated that inflammation, induced by HIV, and increased collagen production, might have led to lower quantities of normal elastin,

and not receiving treatment may result in higher arterial stiffness especially in the elderly [44].

Left ventricular hypertrophy is a maladjusted response to chronic pressure overload and one of the most important risk factors in individuals with hypertension, especially high SBP [45]. Neither blood pressure nor prevalence of hypertension or ECG derived LVH differed between those infected with HIV and the HIV-free in this study. Our findings are in accordance with those of Ogunmola et al. (2014) where no differences in blood pressure or associations with hypertension were found between HIV-infected and HIV-free Nigerians [46].

Human immunodeficiency virus associated nephropathy describes glomerular and end stage renal disease in those infected with HIV-1 and may present with excretion of low levels of protein in urine (<2 g/day) or acute nephrotic proteinuria [47]. The HIV-infected group of this study were ART-naïve at baseline and started with ART during the course of the 10 years. Although the use of ART dramatically improved renal function, the participants were treated with a tenofovir-containing regimen which may result in declining kidney function with long-term use [48]. Although 44% of the HIV infected participants of this study received the tenofovir based regimen for more than five years, our results did not indicate kidney dysfunction or a prevalence of lower than normal CrCl at 10-year follow-up.

As ART lowers inflammation levels in those living with HIV [41] it will return the lipid profile towards normal, resulting in an increase in HDL-C levels [49]. The latter is seen in this study where lower levels of HDL-C were seen among those infected at baseline, but not at follow-up.

The HIV-infected group of this study were treated with efavirenz, emtricitabine and tenofovir containing regimen and may explain the resemblance of our results to those of Ogunmola et al. (2014) who have found a low prevalence of obesity in the HIV-infected group [46]. Metabolic complications usually arise with the use of protease inhibitors (PI) [50]. A lower prevalence of diabetes (also known as a metabolic complication), was reported among the HIV-infected individuals in our study.

Smoking [51] and alcohol consumption [52] are well-established traditional cardiovascular risk factors. Although it was found that HIV-infected individuals are

more likely to smoke [53] and drink excessive alcohol [54], tobacco and alcohol use did not differ between the HIV-infected and HIV-free control group of this study.

This study should be interpreted within the context of its strengths and limitations. Longitudinal studies assessing the 10-year CHD and cardiovascular risk among South Africans are sparse. To the best of our knowledge, the present study is the first to investigate the association between established cardiovascular risk scores and measures of end-organ damage in HIV-infected South Africans and matched HIV-free controls. This study is a well-controlled study and gold standard measurements were conducted with validated apparatus where possible. As it was not possible to measure left ventricular hypertrophy with echocardiography, the Cornell product was used to measure ECG-LVH. Limitations of this study is that this study group was relatively small, however the HIV-infected participants were matched to the HIV-free participants according to age, sex an locality and the HIV-infected participants were known to be infected for at least 10 years. Another limitation is that the plaque scores were not indicated with the measurement of the intima media thickness.

#### Conclusion

To conclude, despite their HIV-status of at least 10 years and 80% being treated, as well as an increase during the 10 years in the CHD risk only among those infected, the HIV-infected participants of our cohort did not have higher CHD or CVD risk when compared to the matched HIV-free participants. Furthermore, no indication of a higher prevalence of end-organ damage was detected among those infected with HIV, which correlates with the findings of the risk score models. Should our results be confirmed in larger studies, the studies would be of great interest to South Africa burdened by the high prevalence of both HIV and CVD.

#### **Conflict of interest**

Authors declared no conflict of interest.

#### Acknowledgements

This work was financially supported by SANPAD (South Africa - Netherlands Research Program on Alternatives in Development), Population Health Research Institute (PHRI), and the Medical Research Council (MRC) of South Africa, South African National Research Foundation North-West University, and the National Research

Foundation, South African Research Chairs Initiative (NRF-SARChI). Opinions expressed and conclusions arrived at, are those of the authors and are not necessarily to be attributed to the NRF.

We thank Dr. L Kruger, the PURE-SA research team, the field workers and office staff in the Africa Unit for Transdisciplinary Health Research (AUTHeR), North-West University, Potchefstroom, South Africa, as well as Dr S Yusuf (PURE-International) and the PURE project staff at the PHRI, Hamilton Health Sciences and McMaster University, ON, Canada.

#### References

- World Health Organization (WHO). Cardiovascular diseases (CVDs)
   [Internet]. 2016 [cited 2016]. Available from:
   <a href="http://www.who.int/mediacentre/factsheets/fs317/en/">http://www.who.int/mediacentre/factsheets/fs317/en/</a>
- 2. Kershaw KN, Osypuk TL, Do DP, De Chavez PJ, Roux AVD. Neighborhood-level racial/ethnic residential segregation and incident cardiovascular disease: The Multi-Ethnic study of Atherosclerosis. Circulation 2015;131(2):141-148.
- 3. Joined United Nations Programme on HIV/AIDS (UNAIDS). Global AIDS update [Internet]. 2016 [cited 2016]. Available from: <a href="http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf">http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf</a>.
- 4. Herbst AJ, Cooke GS, Bärnighausen T, KanyKany A, Tanser F, Newell M-L. Adult mortality and antiretroviral treatment roll-out in rural Kwazulu-Natal, South Africa. Bull World Health Organ 2009;87(10):754-762.
- 5. Zhou DT, Kodogo V, Chokuona KF, Gomo E, Oektedalen O, Stray-Pedersen B. Dyslipidemia and cardiovascular disease risk profiles of patients attending an HIV treatment clinic in Harare, Zimbabwe. HIV AIDS 2015;7:145-155.
- 6. Peck RN, Shedafa R, Kalluvya S, Downs JA, Todd J, Suthanthiran M, et al. Hypertension, kidney disease, HIV and antiretroviral therapy among Tanzanian adults: A cross-sectional study. BMC Med 2014;12(1):1-11.
- 7. Ezzati M, Vander Hoorn S, Lawes CM, Leach R, James WP, Lopez AD, et al. Rethinking the "diseases of affluence" paradigm: Global patterns of nutritional risks in relation to economic development. PLoS Med 2005;2(5):133.
- 8. Lau B, Sharrett AR, Kingsley LA, Post W, Palella FJ, Visscher B, et al. C-reactive protein is a marker for human immunodeficiency virus disease progression. Arch Intern Med 2006;166(1):64-70.
- 9. Yousuf O, Mohanty BD, Martin SS, Joshi PH, Blaha MJ, Nasir K, et al. High-sensitivity c-reactive protein and cardiovascular disease: A resolute belief or an elusive link? J Am Coll Cardiol 2013;62(5):397-408.

- 10. Kuller LH, Tracy R, Belloso W, De Wit S, Drummond F, Lane HC, et al. Inflammatory and coagulation biomarkers and mortality in patients with HIV infection. PLoS Med 2008;5(10):203.
- 11. Ngatchou W, Lemogoum D, Ndobo P, Yagnigni E, Tiogou E, Nga E, et al. Increased burden and severity of metabolic syndrome and arterial stiffness in treatment-naive HIV+ patients from Cameroon. Vasc Health Risk Manag 2013;9:509-516.
- 12. Willum-Hansen T, Staessen JA, Torp-Pedersen C, Rasmussen S, Thijs L, Ibsen H, et al. Prognostic value of a rtic pulse wave velocity as index of arterial stiffness in the general population. Circulation 2006;113(5):664-670.
- 13. Levin A, Djurdjev O, Barrett B, Burgess E, Carlisle E, Ethier J, et al. Cardiovascular disease in patients with chronic kidney disease: Getting to the heart of the matter. Am J Kidney Dis 2001;38(6):1398-1407.
- 14. Freedman BI, Soucie JM, Stone SM, Pegram S. Familial clustering of endstage renal disease in blacks with HIV-associated nephropathy. Am J Kidney Dis 1999;34(2):254-258.
- 15. Kurella M, Lo JC, Chertow GM. Metabolic syndrome and the risk for chronic kidney disease among nondiabetic adults. J Am Soc Nephrol 2005;16(7):2134-2140.
- 16. East MA, Jollis JG, Nelson CL, Marks D, Peterson ED. The influence of left ventricular hypertrophyon survival in patients with coronaryartery disease: Do race and gender matter? J Am Coll Cardiol 2003;41(6):949-954.
- 17. Foraker RE, Rose KM, Kucharska-Newton AM, Ni H, Suchindran CM, Whitsel EA. Variation in rates of fatal coronary heart disease by neighborhood socioeconomic status: The atherosclerosis risk in communities surveillance (1992–2002). Ann Epidemiol 2011;21(8):580-588.
- 18. Mansoor A, Golub ET, Dehovitz J, Anastos K, Kaplan RC, Lazar JM. The association of HIV infection with left ventricular mass/hypertrophy. AIDS Res Hum Retroviruses 2009;25(5):475-481.
- 19. Labarthe D. Epidemiology and prevention of cardiovascular diseases: A global challenge: Jones & Bartlett Publishers; 2010. 710p.

- 20. Ezzati M, Henley SJ, Thun MJ, Lopez AD. Role of smoking in global and regional cardiovascular mortality. Circulation 2005;112(4):489-497.
- 21. Grinspoon S, Carr A. Cardiovascular risk and body-fat abnormalities in HIV-infected adults. New Engl J Med 2005;352(1):48-62.
- 22. Blom DJ. Cardiovascular risk assessment. S Afr Fam Pract 2011;53(2):121-128.
- 23. Dolman RC. The role of diet in cardiovascular disease in black South Africans: Both sides of the story: North-West University; 2013, 50.
- 24. Fourie C, Schutte A, Smith W, Kruger A, van Rooyen J. Endothelial activation and cardiometabolic profiles of treated and never-treated HIV-infected Africans. Atherosclerosis 2015;240(1):154-160.
- 25. Schutte AE, Schutte R, Huisman HW, Van Rooyen JM, Fourie CMT, Malan NT, et al. Are behavioural risk factors to be blamed for the conversion from optimal blood pressure to hypertensive status in black South Africans? A 5-year prospective study. Int J Epidemiol 2012;41(4):1114-1123.
- 26. Schutte AE, Schutte R, Huisman HW, Van Rooyen JM, Fourie CM, Malan NT, et al. Blood pressure variability is significantly associated with ECG left ventricular mass in normotensive Africans: The SABPA study. Hypertens Res 2011;34(10):1127-1134.
- 27. Julius S, Alderman MH, Beevers G, Dahlöf B, Devereux RB, Douglas JG, et al. Cardiovascular risk reduction in hypertensive black patients with left ventricular hypertrophy: The LIFE study. J Am Coll Cardiol 2004;43(6):1047-1055.
- 28. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem 1972;18(6):499-502.
- 29. Cockcroft DW, Gault MH. Prediction of creatinine clearance from serum creatinine. Nephron 1976;16(1):31-41.
- 30. Seedat Y, Rayner B, Veriava Y. South African hypertension practice guideline 2014. Cardiovasc J Afr 2014;25(6):288-294.

- 31. Mancia G, Fagard R, Narkiewicz K, Redon J, Zanchetti A, Böhm M, et al. 2013 ESH/ESC guidelines for the management of arterial hypertension: The task force for the management of arterial hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC). Blood Press 2013;22(4):193-278.
- 32. Aguiar C, Alegria E, Bonadonna RC, Catapano AL, Cosentino F, Elisaf M, et al. A review of the evidence on reducing macrovascular risk in patients with atherogenic dyslipidaemia: A report from an expert consensus meeting on the role of fenofibrate-statin combination therapy. Atheroscler Suppl 2015;19:1-12.
- 33. Van Rooyen J, Fourie C, Steyn H, Koekemoer G, Huisman H, Schutte R, et al. Cardiometabolic markers to identify cardiovascular disease risk in HIV-infected black South Africans. S Afr Med J 2014;104(3):195-199.
- 34. Jiménez MC, Rexrode KM, Glynn RJ, Ridker PM, Gaziano JM, Sesso HD. Association between high-sensitivity c-reactive protein and total stroke by hypertensive status among men. J Am Heart Assoc 2015;4(9):e002073.
- 35. T Pisa P, H Vorster H, Kruger A, Margetts B, T Loots D. Association of alcohol consumption with specific biomarkers: A cross-sectional study in South Africa. J of Health Popul Nutr 2015;33(1):146-156.
- 36. Ferrer E, Curto J, Esteve A, Miro J, Tural C, Riera S, et al. Progression to AIDS or death in HIV-infected patients initiating cART with CD4< 200 cells/µl: The role of CD4 and viral load changes during follow-up. J Intern AIDS Soc 2012;15(4):1-2.
- 37. Meintjes G, Maartens G. Guidelines for antiretroviral therapy in adults. S Afr J Med 2012;13(3):114-133.
- 38. D'Agostino Sr RB, Grundy S, Sullivan LM, Wilson P. Validation of the Framingham coronary heart disease prediction scores: Results of a multiple ethnic groups investigation. JAMA 2001;286(2):180-187.
- 39. Cook NR, Paynter NP, Eaton CB, Manson JE, Martin LW, Robinson JG, et al. Comparison of the Framingham and Reynolds risk scores for global cardiovascular

- risk prediction in the Multiethnic Women's Health Initiative. Circulation 2012;125(14):1748-1756.
- 40. Mashinya F, Alberts M, Van geertruyden J-P, Colebunders R. Assessment of cardiovascular risk factors in people with HIV infection treated with ART in rural South Africa: A cross sectional study. AIDS Res Ther 2015;12(1):1-42.
- 41. Borges ÁH, Dubrow R, Silverberg MJ. Factors contributing to risk for cancer among HIV-infected individuals, and evidence that earlier cART will alter this risk. Curr Opin HIV/AIDS 2014;9(1):34-40.
- 42. Triant VA, Meigs JB, Grinspoon SK. Association of c-reactive protein and HIV infection with acute myocardial infarction. J Acquir Immune Defic Syndr.2009;51(3):268-273.
- 43. Ross AC, O'Riordan MA, Storer N, Dogra V, McComsey GA. Heightened inflammation is linked to carotid intima-media thickness and endothelial activation in HIV-infected children. Atherosclerosis 2010;211(2):492-498.
- 44. Awotedu KO, Longo-Mbeza B, Awotedu AA, Ekpebegh C. Arterial stiffness in hiv patients in a semi urban area of South Africa. Clin Microbiol 2015;4(3):207.
- 45. Katholi RE, Couri DM. Left ventricular hypertrophy: Major risk factor in patients with hypertension: Update and practical clinical applications. Int J Hypertens 2011(2011):495349.
- 46. Ogunmola OJ, Oladosu OY, Olamoyegun AM. Association of hypertension and obesity with HIV and antiretroviral therapy in a rural tertiary health center in Nigeria: A cross-sectional cohort study. Vasc Health Risk Manag 2014;10:129-137.
- 47. Kopp JB, Winkler C. HIV-associated nephropathy in African Americans. Kidney Int 2003;63:43-49.
- 48. Gallant JE, Parish MA, Keruly JC, Moore RD. Changes in renal function associated with tenofovir disoproxil fumarate treatment, compared with nucleoside reverse-transcriptase inhibitor treatment. Clin Infect Dis 2005;40(8):1194-1198.
- 49. Choy E, Sattar N. Interpreting lipid levels in the context of high-grade inflammatory states with a focus on rheumatoid arthritis: A challenge to conventional cardiovascular risk actions. Ann Rheum Dis 2009;68(4):460-469.

- 50. Group DS. Class of antiretroviral drugs and the risk of myocardial infarction. N Engl J Me. 2007;2007(356):1723-1735.
- 51. Teo KK, Ounpuu S, Hawken S, Pandey MR, Valentin V, Hunt D, et al. Tobacco use and risk of myocardial infarction in 52 countries in the Interheart study: A case-control study. Lancet 2006;368(9536):647-658.
- 52. Schutte AE, Schutte R, Huisman HW, Van Rooyen JM, Fourie CM, Malan NT, et al. Are behavioural risk factors to be blamed for the conversion from optimal blood pressure to hypertensive status in black South Africans? A 5-year prospective study. Int J Epidemiol 2012;41:1114-1123.
- 53. De Gaetano Donati K, Cauda R, Iacoviello L. HIV infection, antiretroviral therapy and cardiovascular risk. Mediterr J Hematol Infect Dis 2010;2(3):2010034.
- 54. Braithwaite RS, Nucifora KA, Kessler J, Toohey C, Mentor SM, Uhler LM, et al. Impact of interventions targeting unhealthy alcohol use in Kenya on HIV transmission and AIDS-related deaths. Alcohol Clin Exp Res 2014;38(4):1059-1067.

# Chapter 5: General conclusions and recommendations

#### Introduction

In this chapter, the main findings are summarised, compared to the relevant literature, discussed and concluded in order to point out the differences in the coronary heart disease (CHD) risk score (Framingham) and the cardiovascular risk score (Reynolds) between human immunodeficiency virus (HIV)-infected and HIV-free Africans matched according to age, sex and locality. Associations of the risk scores with measures of end-organ damage are shown. Recommendations for future studies are also given.

#### Hypotheses and comparison to relevant literature

The risk scores did not differ between the HIV-infected and HIV-free participants, despite their HIV status of at least 10 years and 80% being treated with antiretroviral treatment (ART). No indication of a higher prevalence of end-organ damage was detected among the HIV-infected participants, which correlates with the findings of the risk score models. Therefore,

the first hypothesis namely: HIV-infected participants have a higher 10-year cardiovascular disease risk compared to HIV-free participants in both risk score models is rejected because the HIV-infected participants did not show higher risk when compared to the HIV-free controls.

Vos et al. (2017) have found in a study executed in a study group of black participants living in Limpopo, that the Framingham risk did not differ between the HIV-infected and HIV-free group and also did not differ between the treatment-naïve and treated participants [1]. This is in accordance to the findings of the present study, whereas the Framingham risk score between the HIV-infected and HIV-free group did not differ.

There are no studies making use of the Reynolds risk score to assess a 10-year CVD risk in HIV-infected individuals, especially in women [2]. However, according to the literature, the Reynolds risk model showed improved overall discrimination and especially in black and white women [3] and therefore there is no need to develop a race/ethnicity specific risk model for HIV infected Africans [4].

The second hypothesis, namely: A high cardiovascular disease risk score correlates with markers of end-organ damage [increased sub-clinical atherosclerosis (IMT),

increased arterial stiffness (PWV), left ventricular hypertrophy (LVH) and chronic kidney disease (CrCl)] is also rejected.

Parra et al. (2010) did not find any correlation between the Framingham risk score and sub-clinical atherosclerosis in the HIV-infected individuals [5]. This is in accordance with our results, as no significant associations of the risk scores to atherosclerosis were found. Also, no associations of risk scores with arterial stiffness were found. According to the literature, atherosclerosis correlates with arterial stiffness [6]. No associations of risk scores to left ventricular hypertrophy (LVH) or creatinine clearance (CrCl) were seen in both study groups.

#### **Discussion of main findings**

According to the literature, Africans have a high prevalence of cardiovascular risk [7]. During 2015, Eastern and Southern Africa had 19 million individuals infected with HIV [8]. Studies predicted that over 10 million individuals aged older than 50 years will be HIV-infected in sub-Saharan Africa by 2030 [9, 10]. Human immunodeficiency virus infected individuals may show higher cardiovascular risk due to infection, immune activation and the use of ART [11].

In a study conducted by Bergersen et al. (2004), twice as many HIV-infected individuals receiving ART, had a 10-year Framingham risk score of more than 20%, compared to their HIV-free counterparts [12]. However, our results differ from those of Bergersen et al. (2004), as the Framingham risk score of the HIV-infected participants and HIV-free participants did not differ. It was found in a recent study that the HIV-infected African individuals who were treated did not show a higher Framingham risk score [1, 4] which is more in line with our results.

The HIV-infected participants and the HIV-free participants had no difference in the Reynolds risk score in this study. As mentioned, there are no studies incorporating the Reynolds risk score to assess CVD risk in HIV-infected individuals [2], therefore this risk score was included in our study since it is the only risk score that includes C-reactive protein (CRP) as a CVD risk marker [13] and CRP is known to be higher in the HIV-infected population [14, 15].

No associations between the risk scores and measures of end-organ damage were found.

As mentioned, Parra et al. (2010) did not find any correlation between the Framingham risk score and sub-clinical atherosclerosis in the HIV-infected individuals [5]. No indication of atherosclerosis or increased arterial stiffness in the HIV-infected group of this study was found. Several studies reported that higher levels of CRP were associated with an increase in IMT [16, 17] which correlated with arterial stiffness [6]. Because the HIV-infected participants of this study did not show higher levels of CRP, this may be the reason why no prevalence of atherosclerosis or arterial stiffness in the HIV-infected participants was observed.

The prevalence of electrocardiography (ECG) derived LVH did not differ between the HIV-infected and HIV-free participants in this study. Left ventricular hypertrophy is a response to high systolic blood pressure (SBP) [18]. The HIV-infected participants did not have a higher prevalence of hypertension when compared to the HIV-free participants.

The results did not indicate kidney dysfunction or a prevalence of lower than normal CrCl at 10-year follow-up, although nearly half of our HIV-infected participants were treated with tenofovir and this ART is known to be associated with a decrease in kidney function.

#### Conclusion

The HIV-infected participants of this cohort of whom 80% were using ART and were infected for at least 10 years, did not have higher CHD or CVD risk scores when compared to the HIV-free participants. No indication of a higher prevalence of measures of end-organ damage was detected among those infected with HIV, which correlated with the findings of the risk score models.

#### Chance and confounding

It is of importance to critically reflect on some of the factors that may have confounded the results of this study. After 2005, participants commenced with treatment as their CD4 cell count declined below 200 cells/mm<sup>3</sup>. Although the duration of treatment is shown in Table 1 (Chapter 4), the exact duration (months) of each participant's treatment could not be determined.

The study did not test for any opportunistic infections (this was only reported on questionnaires) and final mortality and events data were not available.

Statistical results were evaluated from a physiological perspective and statistical significance does not necessarily indicate physiological significance.

#### Recommendations to future research

- Studies should be conducted where all treated participants receive treatment for a longer period than 5 years.
- A larger experimental and control group, matched according to age, sex and locality, should be employed. Due to incomplete data sets, 29 of the participants had to be excluded which might have reduced the power of the study sample.
- The influence of ART on vascular function needs to be determined to assess
  whether being treated is the reason that we found differences in risk scores
  between the HIV-infected and HIV-free participants and no association of risk
  scores with measures of end-organ damage in this population.
- Similar studies in other provinces of South Africa should be conducted.

#### References

- 1. Vos A, Devillé W, Barth R, Klipstein-Grobusch K, Tempelman H, Venter F, et al. HIV infection and cardiovascular risk profile in a rural South African population: The Ndlovu Cohort study. BMJ Glob Health 2017;2(10):1043-1050.
- 2. Adekunle R, Bagchi S. Review of cardiovascular disease in HIV-infected women. J AIDS Clin Res 2016;7(557):2-12.
- 3. Cook NR, Paynter NP, Eaton CB, Manson JE, Martin LW, Robinson JG, et al. Comparison of the Framingham and Reynolds risk scores for global cardiovascular risk prediction in the Multiethnic Women's Health Initiative. Circulation 2012;125(14):1748-1756.
- 4. Mashinya F, Alberts M, Van geertruyden J-P, Colebunders R. Assessment of cardiovascular risk factors in people with HIV infection treated with ART in rural South Africa: A cross sectional study. AIDS Res Ther 2015;12(1):42-52.
- 5. Parra S, Coll B, Aragones G, Marsillach J, Beltran R, Rull A, et al. Nonconcordance between subclinical atherosclerosis and the calculated Framingham risk score in HIV-infected patients: Relationships with serum markers of oxidation and inflammation. HIV Med 2010;11(4):225-231.
- 6. Ngatchou W, Lemogoum D, Ndobo P, Yagnigni E, Tiogou E, Nga E, et al. Increased burden and severity of metabolic syndrome and arterial stiffness in treatment-naive HIV+ patients from Cameroon. Vasc Health Risk Manag 2013;9:509-516.
- 7. Njelekela MA, Mpembeni R, Muhihi A, Mligiliche NL, Spiegelman D, Hertzmark E, et al. Gender-related differences in the prevalence of cardiovascular disease risk factors and their correlates in urban Tanzania. BMC Cardiovasc Disord 2009;9(30):1-21.
- 8. Joined United Nations Programme on HIV/AIDS (UNAIDS). Global AIDS update [Internet]. 2016 [cited 2016]. Available from:

  <a href="http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf">http://www.unaids.org/sites/default/files/media\_asset/global-AIDS-update-2016\_en.pdf</a>.

- 9. Mills EJ, Barnighausen T, Negin J. HIV and aging--preparing for the challenges ahead. N Engl J Med 2012;366(14):1270-1273.
- 10. Mathers CD, Loncar D. Projections of global mortality and burden of disease from 2002 to 2030. PLoS Med 2006;3(11):442.
- 11. Bloomfield GS, Hogan JW, Keter A, Holland TL, Sang E, Kimaiyo S, et al. Blood pressure level impacts risk of death among HIV seropositive adults in Kenya: A retrospective analysis of electronic health records. BMC Infect Dis 2014;14:284.
- 12. Bergersen BM, Sandvik L, Bruun JN, Tonstad S. Elevated Framingham risk score in HIV-positive patients on highly active antiretroviral therapy: Results from a Norwegian study of 721 subjects. Eur J Clin Microbiol Infect Dis 2004;23(8):625-630.
- 13. Dolman RC. The role of diet in cardiovascular disease in black south africans: Both sides of the story: North-West University; 2013, 50.
- 14. Feldman JG, Goldwasser P, Holman S, DeHovitz J, Minkoff H. C-reactive protein is an independent predictor of mortality in women with HIV-1 infection. J Acquir Immune Defic Syndr 2003;32(2):210-214.
- 15. Reingold J, Wanke C, Kotler D, Lewis C, Tracy R, Heymsfield S, et al. Association of HIV infection and HIV/HCV coinfection with c-reactive protein levels: The Fat Redistribution and Metabolic change in HIV infection (FRAM) study. J Acquir Immune Defic Syndr 2008;48(2):142-148.
- 16. Triant VA, Meigs JB, Grinspoon SK. Association of c-reactive protein and HIV infection with acute myocardial infarction. J Acquir Immune Defic Syndr 2009;51(3):268-273.
- 17. Ross AC, O'Riordan MA, Storer N, Dogra V, McComsey GA. Heightened inflammation is linked to carotid intima-media thickness and endothelial activation in HIV-infected children. Atherosclerosis 2010;211(2):492-498.
- 18. Katholi RE, Couri DM. Left ventricular hypertrophy: Major risk factor in patients with hypertension: Update and practical clinical applications. Int J Hypertens 2011(2011):495349

### **Appendices**

#### Untitled

ORIGINALITY REPORT

7%

3%

4%

0%

SIMILARITY INDEX

INTERNET SOURCES

**PUBLICATIONS** 

STUDENT PAPERS

#### PRIMARY SOURCES

1

dspace.nwu.ac.za

Internet Source

3%

Bitton, A.. "The Framingham Heart Study's Impact on Global Risk Assessment", Progress in Cardiovascular Diseases, 201007/08

Publication

3

"2016 ACR/ARHP Annual Meeting Abstract Supplement", Arthritis & Rheumatology, 2016

2%

**EXCLUDE QUOTES** 

ON

EXCLUDE BIBLIOGRAPHY ON

EXCLUDE MATCHES < 2%

#### DECLARATION

I, C Vorster (ID: 710924 0034 084), Language editor and Translator, and member of the South African Translators' Institute (SATI member number 1003172), herewith declare that I did the language editing of a dissertation written by ms Marlene Duvenhage (student number 23440848).

Title of the article: Cardiovascular disease risk assessment in HIVinfected black South Africans: A longitudinal study

6 Youber	28 June 2017		
C Vorster	Date		