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Assessment of platyhelminth diversity within amphibians of French Guiana revealed a new species of *Nanopolystoma* (Monogenea: Polystomatidae) in the caecilian *Typhlonectus compressicauda*

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Abstract: An expedition was undertaken to French Guiana in search of amphibian parasites. Of the 23 anuran species collected and screened for polystomes, the toad *Rhinella margaritifera* (Laurenti) was the sole species found to be infected with a polystome, namely *Wetapolystoma almae* Gray, 1983. Of the two caecilian species collected, a new species of *Nanopolystoma* du Preez, Huyse et Wilkinson, 2008 was discovered from the urinary bladder of the aquatic caecilian *Typhlonectus compressicauda* (Duméry et Bibon). The small size of the mature worm, two non-diverticulated caeca of equal length that are non-confluent posteriorly, vitelline follicles in two dense lateral fields, a single follicular testis in the middle of the body, small ovary and a single operculated egg in utero, vaginae present and the caecilian host allowed the identification of the specimen as *Nanopolystoma*. Larger body size, hamulus length, egg diameter and occurrence in the caecilian family Typhlonectidae distinguishes the new species from the two other known polystomes in *Nanopolystoma*; thus, the description of *Nanopolystoma tinsleyi* sp. n. is provided within this paper.

Keywords: *Nanopolystoma tinsleyi* sp. n., polystome, Cayenne caecilian, Typhlonectidae, Neotropical region

Scientists attending the First World Congress of Herpetology in 1989 first became concerned about a widespread decline in amphibian population numbers (Stuart et al. 2004). Since then, the number of studies and publications on amphibians increased considerably and the number of species known nearly doubled to the current figure of 7 044 (Frost 2013). In their comprehensive study to identify biodiversity hotspots for conservation priorities, Meyers et al. (2000) reported on the unique biodiversity of the Neotropical realm and the high levels of endemism. This biogeographical area hosts indeed the highest amphibian diversity in the world with 49% of all known amphibian species (Stuart et al. 2004, Vredenburg and Wake 2007), this biogeographical area may have played an important role in the evolution of anuran polystomes. According to Bentz et al. (2001, 2006) South America could be the centre of origin for the cosmopolitan and most diversified genus, i.e. *Polystoma*. In comparison to the South American anuran polystomes that were described mostly in the 1970s and 1980s (see Verneau 2004), *Nanopolystoma* from caecilians has only recently been described and consists of only two species, i.e. *Nanopolystoma brayi* du Preez, Huyse et Wilkinson, 2008 from the urinary bladder of *Caecilia cf. gracilis* Shaw from French Guiana and *N. lynchi* du Preez, Huyse et Wilkinson, 2008 from the urinary bladder and phallosome of *Caecilia cf. pachynema* Günther from an unknown locality in South America.

To increase understanding of global biodiversity, we conducted an expedition to French Guiana to investigate amphibian polystome diversity. In the present paper, a new polystomatid found during this survey is described.
MATERIALS AND METHODS

From 9 to 23 April 2012, three areas were surveyed in French Guiana for frogs and caecilians. They included ponds and marshes in and around Cayenne (4°52′49″; 52°20′6″W), a forest pond on the road to Kaw (4°40′12″N; 52°18′20″W), and several sites in the tropical forest of the Nouragues National Reserve (4°22′4″N; 52°40′29″W). Another area was surveyed in December 2011 by Philippe Gaucher, near St. Georges (3°53′11″N; 51°48′6″W).

Frogs were collected by hand at night with the aid of strong flashlights. They were sorted according to species and placed over night in plastic bags containing some water. The following day the water was poured through two plankton sieves with respective mesh size of 500 and 100 µm. If polystome eggs were retrieved from a particular species, frog specimens were placed individually in bags and kept one day more for a second round of egg screening. With the exception of endangered species, all infected specimens as well as one representative of each species per locality were euthanised using ethyl-4-aminobenzoate (MS222, Sigma, Johannesburg, South Africa) and dissected for parasites and host tissue collections using a Nikon SMZ 645 microscope (Nikon, Amsterdam, Netherlands).

To search for caecilians, we first dug at numerous places in the forest, however, we did not find any terrestrial caecilians. Our focus then shifted to the aquatic caecilian, Tylphlocnemos compressicauda (Duméril et Bibron), known to occur in water bodies in French Guiana. Crayfish traps were modified to reduce the entrance size to about 80 mm in diameter. They were baited with fish or ox liver and set in such a way that the trap was above the water to allow captured non-target air breathing animals to surface. They were secured to vegetation and left overnight in about 20 distinct sites.

The GPS coordinates provided only for the localities where caecilians were caught: Site A, a pool of about 30 m in diameter (4°53′31″N; 52°20′47″W) where six traps were set overnight; site B, a big swamp (4°49′23″N; 52°20′28″W), where 11 traps were set overnight on a first occasion, and 17 at a later occasion; site C, the downstream part of a big river near St. Georges (3°53′11″N; 51°48′6″W), where specimens were collected by P. Gaucher using three funnel traps. Captured caecilians were kept individually in buckets containing water. The procedure for polystome egg screening was the same as described above. Eggs, were rinsed from the 100 µm sieve and transferred to a Petri dish for measurements and larval development with the aim to obtain oncomiracidia for morphological description. All specimens were euthanised using MS222 and dissected to verify that they were not infected with subadult polystomes.

The urinary bladder and all reproductive and excretory ducts were thoroughly screened for polystomes. After all visible parasites were removed, the bladder was placed in hot 70% ethanol and rigorously shaken to detach any small parasites that might have been overlooked. Live parasites were immediately placed in a drop of water on a slide and briefly heated from below with a butane lighter until they relaxed and stopped moving. They were fixed in 10% neutral buffered formalin under very gentle coverslip pressure. Specimens destined for permanent mounting were stained overnight in a weak aceticarmine solution, then were dehydrated and mounted in Canada balsam, whereas a few subadult specimens earmarked for molecular biology were kept in 70% ethanol.

RESULTS

A total of 124 frogs representing seven families and 23 species as well as nine caecilians representing two families and two species were screened for polystomes. Details about species classification and sampling localities are presented in Table 1. A single anuran species was infected, namely Rhinella margaritifera. Among the eight specimens of T. compressicauda collected, one at site A, four at site B and three at site C, a single caecilian from site B was infected with 14 polystomes. Prevalence was 25% for site B and 12.5% for the total sample. The single specimen of C. gracilis that was collected by P. Gaucher on a road following heavy rains some weeks prior to our visit was not infected.

Nanopolystoma tinsleyi sp. n.  
Figs. 1, 2, Table 2

Description (based on three egg-producing adult and eight subadult specimens; eggs harvested failed to develop; measurements expressed as range with mean and number of measurements in parentheses). Adult pyriform (Fig. 1). Average body length of mature specimens 2.8–3.1 mm (2.9 mm). Haptor 684–866 (774) long, 934–948 (941) wide; length: body length ratio of 0.27. Eyespots not observed in adults. Mouth subterminal, ventral, surrounded by false oral sucker. Pharynx spherical. Intestine bifurcate, caeca blind with neither diverticula nor anastomoses, left and right caeca of almost equal length, extending laterally along full length of body proper and extending to posterior medial position just anterior to haptor where two arms come very close to each other but do not join.

Genitointestinal canal present just before entry of posterior vitelline duct, connecting oviduct to caecum. Testis single, prominent, follicular, postovarian, median in midbody extending laterally to vitellarium, in length about 25% of body proper, dense masses of sperm in between testicular follicles (Fig. 1). Vas deferens widens anteriorly forming seminal vesicle, narrows at genital bulb to open at common genital opening. Genital bulb just behind intestinal bifurcation, armed with 16 genital spines; genital spine length 20 (19–20), strongly curved with branched root on proximal ends and sharp point distally pointing outwards, arranged in circle.

Two prominent vaginæ, on lateral margins about one third from anterior end at level of ovary; vaginal ducts descending to respective vitelline ducts. Main left and right vitelline ducts join medially to form vitelline reservoir with posterior duct connecting to oviduct. Vitellarium follicular, dense, from position of intestinal bifurcation confined to 2 lateral fields extending posteriorly to end of body proper, giving impression of a peripheral ring of vitelline follicles.

Ovary prominent, curved oval, to one side of body at level of egg; developing oocytes from tiny dots gradually increasing in size to fully formed oocytes. Oviduct leaves
Table 1. List of anurans and caecilians screened for polystomes.

<table>
<thead>
<tr>
<th>Family</th>
<th>Species</th>
<th>Locality</th>
<th>No. host examined</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bufonidae</td>
<td><em>Rhaebo guttatus</em> (Schneider)</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>Rhinella aff. castaneotica</em> (Caldwell)</td>
<td>Nouragues</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td><em>R. marina</em> (Linnaeus)</td>
<td>Cayenne</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>R. margaritifera</em> (Laurenti)</td>
<td>Cayenne</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td><em>R. quesita</em> (Linnaeus)</td>
<td>Nouragues</td>
<td>33</td>
</tr>
<tr>
<td></td>
<td><em>R. leucura</em> Fouquet, Gaucher, Blanc et Vélez-Rodrigues</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td>Centrolenidae</td>
<td><em>Vitreorana riteae</em> (Latz)</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td>Craugastoridae</td>
<td><em>Pristimantis chiastonotus</em> (Lynch et Hoogmoed)</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>P. zeuxotylus</em> (Lynch et Hoogmoed)</td>
<td>Nouragues</td>
<td>3</td>
</tr>
<tr>
<td>Dendrobatidae</td>
<td><em>Allobates aff. femoralis</em> (Boulenger)</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>A. granti</em> Kok, MacCulloch, Gaucher, Poelman, Bourne, Lathrop et Lengle</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>Dendrobates tnictorius</em> (Cuvier)Nouragues</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>Hylidae</td>
<td><em>Dendropsophus nanus</em> (Boulenger)</td>
<td>Cayenne</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td><em>D. nanus</em></td>
<td>Kav forest</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td><em>Hypisboas cf. cinerascens</em> (Spix)</td>
<td>Nouragues</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td><em>H. punctatus</em> (Schneider)</td>
<td>Cayenne</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td><em>Osteocephalus cf. taurus</em> Steinadchner</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>Phyllomedusa aff. toomopterna</em> Duméril et Bibron</td>
<td>Kav forest</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td><em>Scinax boesemanni</em> (Goin)</td>
<td>Cayenne</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td><em>S. boesemanni</em></td>
<td>Kav forest</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>S. ruber</em> (Laurenti)</td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>Trachycephalus coriaceus</em> (Peters)</td>
<td>Kav forest</td>
<td>13</td>
</tr>
<tr>
<td>Leptodactylidae</td>
<td><em>Leptodactylus pentadactylus</em> (Laurenti)</td>
<td>Cayenne</td>
<td>10</td>
</tr>
<tr>
<td></td>
<td><em>L. pentadactylus</em></td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>L. podicipinus</em> (Cope)</td>
<td>Kav forest</td>
<td>2</td>
</tr>
<tr>
<td>Pipidae</td>
<td><em>Pipa aspera Müller</em></td>
<td>Nouragues</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>P. pipa</em> (Linnaeus)</td>
<td>Cayenne</td>
<td>3</td>
</tr>
<tr>
<td>Typhlonectidae</td>
<td><em>Caecilia gracilis</em> Shaw</td>
<td>Kav forest</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>Typhlonectes compressicuda</em> sp. n.</td>
<td></td>
<td>8</td>
</tr>
</tbody>
</table>

1 six specimens infected with *Wetapolystoma almae* Gray, 1993; 2 one specimen infected with *Nanopolystoma tinsleyi* sp. n.

Ovary, ascends and receives duct from vitelline reservoir, forms ootype, surrounded by Mehlis’ gland. Uterus short, anterior to ovary, containing a single oval egg. Eggs 236–260 (249) × 168–193 (183; n = 9), yellowish-tan, operculated, oval, with operculated end flattened; no indication of intrauterine development. Oncomiracidium not known.

Haptor with 3 pairs of laterally located suckers, diameter 164–187 (176), size and shape variable in adult specimens; hamulus body robust with deep cut between two roots (Fig. 2A), inner length 25–118 (66), outer length 133–175 (164), recurved hook 24–33 (28). In one of paratypes, one hamulus deformed with a second guard (Fig. 2A). Spherical sclerite droplets associated with ha-

Fig. 1. *Nanopolystoma tinsleyi* sp. n. from *Typhlonectes compressicuda*. Ventral view of holotype (NMB P357). Abbreviations: eg – egg; gb – genital bulb; gi – genitointestinal canal; ha – hamulus; hp – haptor; ic – intestinal caecum; mg – Mehlis’ gland; mh – marginal hooklets; mo – mouth; ov – ovary; ph – pharynx; su – sucker; sv – seminal vesicle; te – testis; vd – vas deferens; vg – vagina; vi – vitellarium.

Type host: *Typhlonectes compressicuda* (Duméril et Bibron).

Type locality: Swamp close to Cayenne, French Guiana.
Table 2. Diagnostic measurements for *Nanopolystoma tinsleyi* sp. n., *N. brayi* and *N. lynchi*. Range with mean in parentheses are given in micrometres (µm), except for body length.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Nanopolystoma tinsleyi sp. n.</th>
<th>N. brayi</th>
<th>N. lynchi</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>subadult</td>
<td>mature</td>
<td>mature</td>
</tr>
<tr>
<td>Number of specimens</td>
<td>8</td>
<td>3</td>
<td>4</td>
</tr>
<tr>
<td>Body length (mm)</td>
<td>0.7–2.1 (1.6)</td>
<td>2.8–3.1 (2.9)</td>
<td>1.1–1.4 (1.2)</td>
</tr>
<tr>
<td>Maximum width</td>
<td>346–703 (549)</td>
<td>1076–1153 (1123)</td>
<td>543–572 (555)</td>
</tr>
<tr>
<td>Haptor length</td>
<td>273–502 (425)</td>
<td>684–866 (774)</td>
<td>398–446 (415)</td>
</tr>
<tr>
<td>Haptor width</td>
<td>365–695 (545)</td>
<td>934–948 (941)</td>
<td>388–529 (457)</td>
</tr>
<tr>
<td>Width at vagina</td>
<td>452–694 (534)</td>
<td>971–1060 (1013)</td>
<td>514–534 (529)</td>
</tr>
<tr>
<td>False oral sucker width</td>
<td>76–137 (102)</td>
<td>167–180 (175)</td>
<td>-</td>
</tr>
<tr>
<td>Pharynx length</td>
<td>101–118 (113)</td>
<td>161–190 (174)</td>
<td>60–86 (68)</td>
</tr>
<tr>
<td>Ovary length</td>
<td>129–228 (184)</td>
<td>275–376 (328)</td>
<td>106–130 (119)</td>
</tr>
<tr>
<td>Uterine egg length</td>
<td>-</td>
<td>236–260 (249)</td>
<td>182–238 (221)</td>
</tr>
<tr>
<td>Uterine egg width</td>
<td>-</td>
<td>168–193 (183)</td>
<td>110–115 (114)</td>
</tr>
<tr>
<td>Testis length</td>
<td>81–486 (279)</td>
<td>385–577 (492)</td>
<td>-</td>
</tr>
<tr>
<td>Testis width</td>
<td>166–485 (390)</td>
<td>674–861 (764)</td>
<td>-</td>
</tr>
<tr>
<td>Genital bulb width</td>
<td>-</td>
<td>68–71 (70)</td>
<td>91–108 (97)</td>
</tr>
<tr>
<td>Genital spine number</td>
<td>-</td>
<td>16</td>
<td>16–19</td>
</tr>
<tr>
<td>Genital spine length</td>
<td>-</td>
<td>19–20 (20)</td>
<td>20–22 (21)</td>
</tr>
<tr>
<td>Haptoral sucker diameter</td>
<td>81–133 (109)</td>
<td>164–187 (176)</td>
<td>122–139 (129)</td>
</tr>
<tr>
<td>Hamulus length</td>
<td>90–136 (115)</td>
<td>133–175 (164)</td>
<td>74–110 (95)</td>
</tr>
<tr>
<td>Hamulus hook</td>
<td>21–29 (23)</td>
<td>24–33 (28)</td>
<td>26</td>
</tr>
<tr>
<td>Marginal hooklet length</td>
<td>-</td>
<td>18.4–20.3 (19.6)</td>
<td>19–20 (20)</td>
</tr>
</tbody>
</table>

Fig. 2. *Nanopolystoma tinsleyi* sp. n. from *Typhlonectes compressicauda*. A – hamulus from holotype NMB P357 (a) and paratypes NMB P 358–360 (b, c); B – marginal hooklets 1 from holotype and paratypes. Abbreviations: X – outer length; Y – inner length; Z – hook length.

(4°49′23″N; 52°20′28″W; 28 m a.s.l.).

**Site:** Urinary bladder.

**Deposition of specimens:** Eleven worms of which three were sexually mature were mounted as permanent slides. The remaining three subadult specimens were fixed for further molecular studies. Deposited specimens include the holotype (NMB P357) and three paratypes (NMB P358-360) in the Parasitic Worm Collection, National Museum, Bloemfontein, South Africa and one paratype (IPCAS M-556) in the Helminthological Collection, Institute of Parasitology, Biology Centre of ASCR, České Budějovice, Czech Republic. The remainder of specimens was deposited in the collec-
Ethenology: This species is named after Dr. Richard Tin-
sley in acknowledgement of his tremendous contribution to
knowledge of polystomatin flatworms.

Remarks. With an average total length of 2.9 mm
Nanopolystoma tinsleyi sp. n. is about double the size of
the other two known species, specifically 1.2 mm for
N. brayi and 1.7 mm for N. lynchi (Table 2). This signifi-
cant difference in the total length applies for most body
measurements. Where the hamulus hook length is basi-
cally identical for N. tinsleyi, N. brayi and N. lynchi, the
total hamulus length for N. tinsleyi, i.e. 164 µm in average
(133–175 µm), is significantly greater than in N. brayi,
and N. lynchi, i.e. 104 µm (81–121 µm) (Table 2). For both
N. brayi and N. lynchi, the hook length expressed as a percentage of the hamulus
length was 27%, whereas it is only 17% for N. tinsleyi.
The egg length for N. tinsleyi, i.e. 249 µm (236–260 µm),
is greater than in N. brayi, i.e. 221 µm (182–238 µm),
and N. lynchi, i.e. 218 µm (151–262 µm), though it is
not significant. In contrast, egg width of 183 (168–193)
for N. tinsleyi is significantly larger than in N. brayi, i.e.
114 µm (110–115 µm) and substantially larger than in
N. lynchi, i.e. 123 µm (110–173 µm) (Table 2). The geni-
tal spine number of 16 for N. tinsleyi does separate it from
N. lynchi with 10–12 genital spines (Table 2).

Finally, even if it is tentative to consider the host spe-
cies as a criterion to describe new parasite species, T. com-
pressicauda, which belongs to the Typhlonectidae, differs
in morphological, biological (Wilkinson and Nussbaum
2006) and phylogenetic (Zhang and Wake 2009, Pyron
and Wiens 2011) features from Caecilia gracilis and Caecil-
ia cf. pachynema (Caeciliidae), which are the respec-
tive hosts for N. brayi and for N. lynchi.

Although unpublished sequences 18S of rDNA (Gen-
Bank Acc. No. KM282386), 28S rDNA (GenBank Acc.
No. KM282387) and cox1 (GenBank Acc. No.
KM282388) of N. tinsleyi confirmed that the genus Na-
нополостoma is a valid taxon, the lack of DNA for the two
other known species in the genus prevented to unravel
their phylogenetic relationship.

DISCUSSION

Nanopolystoma tinsleyi sp. n. fits the generic crite-
rion of LDP at the North-West University, South Africa.

Madapolystoma du Preez, Raharivololoniaaina, Verneau,
Vences, 2010. The absence of a uterus implies that eggs
cannot be accumulated and thus intrauterine development
is not possible. Therefore, the presence of adult and sub-
adult specimens of N. tinsleyi in the same host suggests
that adult hosts can be re-infected. In a vast swamp
the chances of a free-swimming oncomiracidium to locate
a suitable host is so remote that one can assume that host
behaviour will be instrumental in facilitating infection
and reinfection. It was reported that T. compressicauda
spends daytime in a communal burrow and only leaves at
night to feed (AmphibiaWeb 2014). This might provide
the opportunity for polystome eggs to be deposited and
to hatch in the burrow, which may facilitate infection of
a community of caecilians. However, this needs to be fur-
ther investigated.

Hamuli of N. tinsleyi as well as those of N. brayi and
N. lynchi have the same basic shape with a very deep inci-
sion between the handle and the guard. In the original de-
scription of both N. brayi and N. lynchi, most of the hamu-
li were not oriented flat (du Preez et al. 2008). However, the
two flat oriented hamuli reported for N. lynchi (see du Preez
et al. 2008) had the same shape as reported here for N. tin-
sleyi. The incision between the handle and the guard of
the hamuli is the deepest reported for any known polystome.
Euzet et al. (1974) recognised the potential of the hamulus
shape as a taxonomic character for polystomes and took
four measurements per hamulus. Murith (1981) separated
several African polystomes based on the hamulus shape.
Since hamulus shape varies considerably between different
polystome genera, an in-depth study on the variation and
significance of this morphological structure is needed.

We have very good reasons to believe that the 108
species of amphibian polystomes currently known under-
represent global polystome diversity. Du Preez (1996) re-
ported 27 anuran species from Vernon Crookes Reserve,
a tiny coastal reserve in the KwaZulu-Natal Province of
South Africa. Of these, no less than 10 species hosted
polystomes. Du Preez (2011) reported that 11 of the 49
species of grass frog Ptychadena Boulenger known from
Africa are hosts for polystomes. Since several species of
Ptychadena have not yet been screened for polystomes, it
is likely that more polystome species remain to be discov-
ered. In a recent study conducted in the Okumu Reserve
in Nigeria, researchers revealed no less than ten differ-
ent polystome species including one undescribed species
from ten different host species in a fairly small geographical
area within the National Park (M.S.O. Aisien, Univer-
sity of Benin City, Benin City, Nigeria – pers. comm.).
Currently, there are 51 polystome species known from
amphibians in the Ethiopian Realm and 21 from the Neo-
tropical Realm. Though we found only two infected am-
phibian species during our field investigations in French
Guiana (Table 1), we can assume that the exceptionally
rich amphibian fauna of the Neotropical Realm may be
host to a larger diversity of polystomes.
The Polystomoidae includes the three known polystome genera from chelonians, namely *Polystomoides* Ward, 1917, *Polystomoidella* Price, 1939 and *Neopolystoma* Price, 1939, as well as *Nanopolystoma* based on morphological similarities that it shares with the chelonian polystomes (see du Preez et al. 2008). All known species of the Polystomoidae also have a non-sanguinivorous diet and it is believed that they feed on mucous and/or epithelial cells (Allen and Tinsley 1989). Du Preez and Moeng (2004) stated that an epithelial diet, which is common in polystomes of chelonians, Australian lungfish and hippopotamus, might reflect a common ancestry. Verneau et al. (2002) showed, based on a molecular phylogenetic study of polystomes covering 11 polystome genera, that chelonian and amphibian polystomes form two distinct sister clades. Because the lack of blood pigments in the intestinal caeca of *Nanopolystoma* also indicates a non-sanguinivorous diet, it should therefore be very interesting to investigate its phylogenetic position within the Polystomatidae as it might provide insights into the origin of chelonian polystomes (see Verneau et al. 2002). Examining *Nanopolystoma* oncomiracidia might also supply interesting features on polystome evolution as the ciliated cell pattern and chaetotaxy are distinct for polystome genera.

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